Structure-Activity Relationships in Human Toll-like Receptor 7-Active Imidazoquinoline Analogues

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Abstract
Engagement of toll-like receptors serve to link innate immune responses with adaptive immunity and can be exploited as powerful vaccine adjuvants for eliciting both primary and anamnestic immune responses. TLR7 agonists are highly immunostimulatory without inducing dominant proinflammatory cytokine responses. A structure-activity study was conducted on the TLR7-agonistic imidazoquinolines, starting with 1-(4-amino-2-((ethylamino)methyl)-1H-imidazo[4,5-c]quinolin-1-yl)-2-methylpropan-2-ol as a lead. Modifications of the secondary amine of the C2 ethylaminomethylene sidechain are poorly tolerated. The 4-amino group must be retained for activity. Replacement of the imidazole ring of the scaffold with triazole or cyclic urea led to complete loss of activity. A systematic exploration of N1-benzyl-C2-alkyl substituents showed a very distinct relationship between alkyl length and TLR7-agonistic potency with the optimal compound bearing a C2-n-butyl group. Transposition of the N1 and C2 substituents led to the identification of an extremely active TLR7-agonistic compound with an EC50 value of 8.6 nM. The relative potencies in human TLR7-based primary reporter gene assays were paralleled by interferon-α induction activities in whole human blood models.

Keywords
Toll-like receptor; TLR7; Imidazoquinoline; Vaccine adjuvants; Interferon; Innate immunity

Introduction
Toll-like receptors (TLRs) are pattern recognition receptors that recognize specific molecular patterns present in molecules that are broadly shared by pathogens, but are structurally distinct from host molecules. There are 10 TLRs in the human genome. The ligands for these receptors are highly conserved microbial molecules such as lipopolysaccharides (LPS) (recognized by TLR4), lipopeptides (TLR2 in combination with TLR1 or TLR6), flagellin (TLR5), single stranded RNA (TLR7 and TLR8), double stranded RNA (TLR3), CpG motif-containing DNA (recognized by TLR9), and profilin present on uropathogenic bacteria (TLR 11). TLR1, -2, -4, -5, and -6 respond to extracellular stimuli, while TLR3, -7, -8 and -9 respond to intracytoplasmic PAMPs, being associated with the endolysosomal compartment.

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Supporting Information: Characterization of intermediates and final compounds (1H, 13C, mass spectra). This material is available free of charge via the Internet at http://pubs.acs.org.
The activation of TLRs by their cognate ligands leads to activation of innate immune effector mechanisms, including the production of pro-inflammatory cytokines, and up-regulation of MHC molecules and co-stimulatory signals in antigen-presenting cells as well as activating natural killer (NK) cells. The consequence of activation of the innate immune system mobilizes and amplifies specific adaptive immune responses involving both T- and B-cell effector functions.\(^5\) Thus, TLR stimuli serve to link innate and adaptive immunity\(^5\) and can therefore be exploited as powerful adjuvants in eliciting both primary and anamnestic immune responses.

We have recently examined representative members of virtually the entire compendium of known TLR agonists in a series of hierarchical assays including primary TLR-reporter assays, secondary indices of immune activation such as cytokine induction and activation of lymphocytic subsets in whole human blood, and tertiary screens characterizing transcriptomal activation patterns with a view to identifying optimal immunostimulatory chemotypes.\(^8\) Of all the innate immune stimuli examined, we found that TLR7 agonists were extraordinarily immunostimulatory, stimulating virtually all subsets of lymphocytes, and yet without inducing dominant proinflammatory cytokine responses (unlike TLR4 -5 or -8 agonists, which were proinflammatory and therefore may exert systemic toxicity).\(^8\) We therefore became especially desirous of evaluating TLR7-active compounds as potential vaccine adjuvants.

Long before endosomal TLR7 was discovered to serve as the primary sensor for short, single-stranded, GU-rich RNA sequences (ssRNA), mainly of viral origin,\(^9\)-\(^11\) a number of small molecules were synthesized and evaluated in the 1970s and '80s for antiviral activities owing to their pronounced Type I interferon (IFN-\(\alpha\) and -\(\beta\)) inducing properties.\(^12\)-\(^16\) Although the mechanisms of innate immune stimulation of several of these compounds (such as tilorone\(^14\) and bromopirone\(^16\)) remain yet to be formally elucidated, members of the 1\(H\)-imidazo[4,5-c]quinolines were found to be good Type I IFN inducers in human cell-derived assays,\(^17\) and FDA approval was obtained in 1997 for Imiquimod (1) for the treatment of basal cell carcinoma and actinic keratoses.\(^18\) It was not until 2002, however, that the mechanistic basis of IFN induction by the imidazoquinolines was found to be a consequence of TLR7 engagement and activation.\(^19\) Other than the original landmark studies performed by investigators at 3M Pharmaceuticals,\(^17\) structure-activity relationships of the imidazoquinoline chemotype remains poorly explored, perhaps attributable in part to recent interest in the 8-hydroxy-adenine compounds as alternate TLR7-agonists,\(^20\)-\(^23\) which appear to lack emetic side-effects observed in ferrets upon oral administration.\(^22\)

An aspect of our recent work in this area\(^8;24;25\) focuses on developing adjuvants for transcutaneous vaccines (needle-free vaccine patches).\(^26\)-\(^28\) Given that imiquimod is already approved for topical use, it was of particular interest not only to explore chemical space around the imidazoquinolines, but also to carefully ‘immunophenotype’ such compounds in human TLR7-based assays to verify that they did not have any TLR8-driven proinflammatory properties. One of our goals was also to learn from such structure-activity studies optimal positions on the scaffold that would tolerate the introduction of electrophilic or photoactivatable labels for purposes of developing self-adjuvanting vaccine constructs\(^29;30\) by covalently modifying protein antigens. We report here the synthesis and evaluation of a focused library of variously substituted 1\(H\)-imidazo[4,5-c]quinolines, and the identification of a novel, pure TLR7 agonist whose potency is 250 times that of imiquimod.

**Results and Discussion**

As alluded to earlier, long before the fact that imidazoquinolines specifically engaged and activated TLR7 was known, it was recognized that these compounds were potently adjuvantic\(^31\)-\(^34\) in addition to displaying anti-viral effects via the induction of IFN-\(\alpha/\beta\).\(^35;36\) In human blood stimulated ex vivo with TLR7 agonists, upregulation of the surface expression...
of costimulatory molecules such as CD40, CD80, and CD86 occurred in both CD11c–
plasmacytoid DCs and CD11c+ myeloid DCs; furthermore, the Th1 stimulatory ability of both 
DC subsets was enhanced in response to TLR7 ligands. Because TLR7 agonists transmit a 
T-helper-like signal to antibody-producing B cells, it is a highly effective adjuvant even for 
synthetic peptides that lack T-cell epitopes, possibly even replacing the function of T-helper 
cells, and providing a potential T-cell-independent vaccination strategy.

Our initial interest in exploring TLR7 agonists as vaccine adjuvants has been greatly reinforced 
by our recent observations that pure TLR7 agonists, unlike other TLR ligands, are potently 
immunostimulatory without activating inflammatory programs in human whole blood model 
systems. We elected to choose 1-(4-amino-2-((ethylamino)methyl)-1H-imidazo[4,5-c] 
quinoxalin-1-yl)-2-methylpropan-2-ol, 2 as our point of departure in examining structure-activity 
relationships in the imidazoquinolines, rather than imiquimod because we had earlier 
established that this analogue was more active than the parent compound, and was also verified 
to be a pure TLR7 agonist in human primary cells. Furthermore, the only report on a detailed examination of SAR on the imidazoquinolines 
by the 3M group had used indirect bioassays (indirect assay for IFN-α using inhibition of 
virus-induced cytopathic effect) which predated the discovery and development of TLR7-
specific assays. In the studies reported below, all compounds have been evaluated in human 
TLR7 and TLR8 reporter gene assays; highly active compounds were further examined for 
IFN-α induction in whole human blood using analyte-specific immunoassays.

Recent studies show that the TLR7 agonistic imidazoquinolines are indeed potently adjuvantic 
in vaccine constructs; however, the route of administration appears to be crucial since 
intradermal, but not subcutaneous administration of the vaccine adjuvanted with 2 is 
immunogenic and protective. Epidermal Langerhans cells as well as dermal dendritic cells 
(myeloid and plasmacytoid subsets) function as specialized professional antigen presenting 
cells in the skin. The plasmacytoid dendritic cell subset appears to be the target cell-type 
for TLR7 agonists. We hypothesized that the C2 ethylaminomethylene substituent of 2 renders the molecule sufficiently polar as to hinder transcutaneous penetration to effectively 
access and stimulate plasmacytoid dendritic cells. It was therefore desirable to evaluate more 
hydrophobic analogues of 2 as potential candidate transcutaneous vaccine adjuvants.

Unlike TLR2, TLR3, and TLR4 for which crystal structures are available as complexes with 
their cognate ligands, a detailed structural characterization of the mode of binding of the 
imidazoquinolines to TLR7 is not yet available. Therefore, our strategy in examining structure-activity 
relationships was focused rather on exploring chemical space systematically around the 
validated lead compound 2, toward realizing the two major objectives mentioned earlier. 
We began with derivatization of the secondary amine of the C2 ethylaminomethylene sidechain 
in an effort to examine whether this could be a potential site for introducing photoactivable or 
electrophilic functionalities. We found that modifications on this substituent are poorly 
tolerated, with only the N-ethyl analogue 3 (Scheme 1) retaining partial activity, and acyl (4, 5) and guanidine (6) derivatives being bereft of activity (Table 1). The loss of activity 
appeared not to be a consequence of altered basicity of the amine; surmising that it may, in 
part, be due either to the hydrophobicity of the substituent, or the conformal constraint imposed 
by the amide or guanidine, we asked if a polar spacer group, such as triethyleneglycol (11) 
may mitigate loss of activity, but we were proven wrong because 11 was also found to be 
completely inactive. Concurrent with the syntheses of the above compounds, we also 
synthesized the N-propylenesiothiocyanate derivative (14) via the N-propyleneamino (13) and 
N-ethylenenitrile (12) precursors. Compound 14 was not tested in reporter gene assays because we anticipated spurious results on account of its reactivity; the reaction of 14 with the spermine 
derivative (used as a model amine) afforded the expected thiourea adduct (16) which, 
disappointingly, was also inactive. These results conclusively showed that the secondary amine
on the C2 substituent was not amenable to modifications. Replacement of the terminal ethyl group on the C2 substituent with an n-undecanoyl group in 22 resulted in complete loss of activity (Table 1), signaling that the length of the C2 substituent was crucial; this was to become dramatically apparent when 32, with a pentyl substituent on C2 was evaluated later (see below).

Desiring more lipophilic analogues which are expected to permeate the dermal barrier better, we sought to examine N^1-benzyl analogues (Scheme 3) noting that similar substitutions have resulted in augmented activity in the 8-hydroxy-adenine series.^{20-23} Compound 32 was the first to be synthesized in this series which, to our pleasant surprise showed an activity twenty-fold greater than 2. Coupled with our observation that 22 was completely inactive, we sought to systematically explore SAR of the N^1-benzyl C2 alkyl substituents. As depicted in Fig. 2, a very distinct relationship between alkyl length and TLR7-agonistic potency was observed, with the optimal compound being 31 (C2-n-butyl), with an EC_{50} of 59 nM (Table 1). n-Butylene (38) and n-butylene (43) analogues showed decrease in potency. It is noteworthy that a C2-n-butyl, N^1-phenyl analogue synthesized by the 3M group was not the most active; our results indicate that an N^1-benzyl substituent is preferred.

In recognition that bioactivity readouts using cell-culture systems may not always reflect with fidelity in vivo behavior owing to differential plasma protein binding (that we ourselves have observed and characterized),^{46} it was important to verify that the activity profiles observed in TLR7-specific reporter gene assays (Table 1) was also seen in whole blood with its full complement of plasma proteins. It was gratifying that we observed the expected 31 ≫ 2 ≈ 34 activity profile in whole blood IFN-α induction experiments (Fig. 3).

Bioisosteric replacements of the C4-NH$_2$ group with -NMe, -OH and -SMe groups on the imiquimod scaffold have been explored earlier,^{17} and it was of interest to examine if the previously-described SAR would hold true in our analogues, and also to test whether the introduction of phenyl, -NHNH$_2$ and -NHOH groups would alter TLR7 activity (Scheme 4). The 4-Cl precursor (44) and the desamino analogue 50, as well as the 4-OH (46) and 4-Ph (47) analogues were completely inactive, while the 4-NHOH (48) and 4-NHNNH$_2$ (49) compounds were substantially weaker (Table 1) than their 4-NH$_2$ counterparts (31, 32). These results indicate the importance of the preservation of the NH$_2$ group on C4 for maximal activity.

Up to this point in our SAR studies, the most active and chemically distinct species were compound 2, and the N^1-benzyl-C2-n-butyl analogue 31. We were curious to examine whether transposing the N^1 and C2 substituents on these molecules would result in enhanced activity. Fortuitously, both the ‘hybrids’ (52, 54; Scheme 5) were very active. 54 was found to be extraordinarily potent with an EC$_{50}$ of 8.6 nM (Fig. 4, Table 1).

Finally, we attempted to modify the imidazole ring of the scaffold. The triazole compounds 55 and 56, as well as the cyclic urea 57 were completely inactive, which emphasizes the role of the recognition of the imidazole ring system by TLR7. Mention must also be made that these studies have been useful in delineating possible sites for introducing labeling functionalities. Preliminary studies (data not shown) show that the N^1-benzyl substituent may be appropriate for introducing arylazido functional groups, while the quinoline ring may be amenable to introducing electrophilic groups. These results will be communicated in a future publication.

All of the C2-alkyl compounds reported herein were tested for TLR8 activity using the thiazoloquinoline CL075 (3M002) as a reference TLR8 agonist,^{47,48} and none had any appreciable activity in these assays. It should also be pointed out that while potencies of compounds are conventionally represented as EC$_{50}$ (or IC$_{50}$) values, that metric alone appears to be inadequate in rank-ordering the activities of the active compounds tested. An inspection of Fig. 4 indicates, for instance, that while the EC$_{50}$ values of both imiquimod and 2 are

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comparable (~ 2 μM), compound 2 induces a more robust response than imiquimod. Similarly, the area-under-curve estimates of 54 (analogous to combined ‘avidity’ and affinity measures of antigen-antibody interactions)\(^{49,50}\) indicate that consideration of the EC\(_{50}\) values alone may underestimate its potency.

Given that TLR7 is sequestered in an acidic endolysosomal compartment, and these compounds are weak bases and are therefore expected to accumulate in that acidic organelle, we wondered if the observed SAR could simply be a consequence of differential hydrophobicity of the compounds, modulating uptake into the cell and entry into the endolysosomal compartment activity, rather than governing specific molecular interactions with TLR7. A comparison of observed retention times on C\(_{18}\)-reverse-phase mass-chromatograms (a measure of hydrophobicity)\(^{51,52}\) with the potencies of the active compounds indicate a distinct deviation from the trend of 54, suggesting specific and idiosyncratic effects, rather than bulk-effects (Fig. 5).

These structure-activity studies have been instructive and rewarding in that they not only complement the seminal work at 3M, but also have led to the discovery of highly-potent, lipophilic, human TLR7 agonist, 54. We note, not without irony, that this highly potent compound differs from its parent molecule by only a replacement of a secondary amine by a methylene unit. The oral bioavailability of 54 is predicted to be good, and it is possible that the systemic IFN-α inducing effects of this imidazoquinoline may find additional uses in the treatment of conditions such as hepatitis,\(^{53}\) chronic myelogenous\(^{54}\) and hairy cell leukemias.\(^{55}\) However, for purposes of developing dermal vaccine adjuvants, 31 may be more efficacious for reasons discussed earlier. Compounds 54 and 31 are being exhaustively immunoprofiled and characterized in a variety of assays.

**Experimental Section**

**Chemistry**

All of the solvents and reagents used were obtained commercially and used as such unless noted otherwise. Moisture- or air-sensitive reactions were conducted under nitrogen atmosphere in oven-dried (120 °C) glass apparatus. The solvents were removed under reduced pressure using standard rotary evaporators. Flash column chromatography was carried out using RediSep Rf ‘Gold’ high performance silica columns on CombiFlash Rf instrument unless otherwise mentioned, while thin-layer chromatography was carried out on silica gel CCM pre-coated aluminum sheets. Purity for all final compounds was confirmed to be greater than 97% by LC-MS using a Zorbax Eclipse Plus 4.6 mm × 150 mm, 5 μm analytical reverse phase C\(_{18}\) column with H\(_2\)O-isopropanol or H\(_2\)O-CH\(_3\)CN gradients and an Agilent ESI-TOF mass spectrometer (mass accuracy of 3 ppm) operating in the positive ion (or negative ion, as appropriate) acquisition mode.

**Synthesis of Compound 3: 1-(4-Amino-2-((diethylamino)methyl)-1\(H\)-imidazo[4,5-c]quinolin-1-yl)-2-methylpropan-2-ol**

To a solution of trifluoroacetate salt of 2 (75 mg, 0.14 mmol) in anhydrous MeOH, were added acetaldehyde (6.6 mg, 0.15 mmol), 12-14 drops of acetic acid and sodium cyanoborohydride (10 mg, 0.15 mmol). After 4 hours the solvent was removed using vacuum and the residue was purified using column chromatography (7% MeOH/dichloromethane) to obtain the compound 3 (16 mg, 34%). \(^1\)H NMR (500 MHz, MeOD) \(\delta\) 8.36 (d, \(J = 7.8\) Hz, 1H), 7.73 (dd, \(J = 8.4, 1.0\) Hz, 1H), 7.69 – 7.64 (m, 1H), 7.52 (dd, \(J = 8.4, 7.2, 1.2\) Hz, 1H), 4.83 (s, 2H), 4.75 – 4.63 (m, 2H), 3.42 (dd, \(J = 14.2, 7.0\) Hz, 4H), 1.39 – 1.29 (m, 6H), 1.21 (dd, \(J = 13.5, 6.2\) Hz, 6H). \(^{13}\)C NMR (126 MHz, MeOD) \(\delta\) 150.85, 150.16, 138.41, 136.09, 131.52, 126.42, 126.18,

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123.69, 120.11, 114.65, 72.54, 56.68, 49.70, 49.56, 27.65, 9.45. MS (ESI) calculated for C$_{19}$H$_{27}$N$_5$O, m/z 341.22, found 342.24 (M + H)$^+$. 

**Synthesis of Compound 4: N-((4-Amino-1-(2-hydroxy-2-methylpropyl)-1H-imidazo[4,5-c]quinolin-2-yl)methyl)-N-ethylacetamide**

To a solution of trifluoroacetate salt of 2 (40 mg, 0.074 mmol) in anhydrous THF, were added triethylamine (23 mg, 0.22 mmol) and acetyl chloride (6 mg, 0.08 mmol). After 30 minutes, the solvent was removed under vacuum. The residue was dissolved in EtOAc, washed with water and dried over Na$_2$SO$_4$. The EtOAc fraction was then concentrated under vacuum and the residue was purified using column chromatography (6% MeOH/dichloromethane) to obtain the compound 4 (22 mg, 85%). $^1$H NMR (400 MHz, MeOD) $\delta$ 8.34 (d, $J = 8.3$ Hz, 1H), 7.71 (d, $J = 8.3$ Hz, 1H), 7.53 (t, $J = 7.7$ Hz, 1H), 7.38 (t, $J = 7.7$ Hz, 1H), 5.23 (s, 2H), 4.81 (s, 2H), 3.63 (q, $J = 7.1$ Hz, 2H), 2.23 (s, 3H), 1.34 – 1.13 (m, 9H). $^{13}$C NMR (101 MHz, MeOD) $\delta$ 172.18, 154.62, 136.88, 134.07, 129.68, 129.48, 124.60, 122.64, 122.31, 118.21, 113.67, 70.85, 65.49, 55.12, 43.70, 41.79, 19.68, 14.03, 12.30. MS (ESI) calculated for C$_{19}$H$_{25}$N$_5$O$_2$, m/z 355.20, found 356.21 (M + H)$^+$. 

**Synthesis of Compound 5: N-((4-Amino-1-(2-hydroxy-2-methylpropyl)-1H-imidazo[4,5-c]quinolin-2-yl)methyl)-N-ethylpalmitamide**

To a solution of trifluoroacetate salt of 2 (20 mg, 0.05 mmol) in anhydrous THF, were added triethylamine (15 mg, 0.15 mmol) and palmitoyl chloride (16 mg, 0.06 mmol). After 30 minutes the solvent was removed under vacuum, residue was dissolved in EtOAc and washed with water. The EtOAc fraction was then dried under vacuum and the residue was purified using column chromatography (5% MeOH/dichloromethane) to obtain the compound 5 (10 mg, 30%). $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 8.12 (d, $J = 8.1$ Hz, 1H), 7.88 (d, $J = 8.3$ Hz, 1H), 7.54 (t, $J = 7.6$ Hz, 1H), 7.38 (t, $J = 7.8$ Hz, 1H), 4.75 (s, 2H), 3.56 (q, $J = 6.8$ Hz, 2H), 2.45 – 2.34 (m, 2H), 1.76 – 1.57 (m, 2H), 1.58 – 1.11 (m, 35H), 0.90 (t, $J = 6.8$ Hz, 3H). $^{13}$C NMR (101 MHz, CDCl$_3$) $\delta$ 192.82, 173.95, 152.03, 150.36, 135.45, 130.90, 128.29, 124.10, 123.24, 120.62, 114.81, 71.96, 55.58, 42.81, 42.30, 33.06, 31.92, 29.68, 29.65, 29.52, 29.47, 29.42, 29.35, 25.37, 22.69, 14.12, 13.70, 5.61. MS (ESI) calculated for C$_{33}$H$_{53}$N$_5$O$_2$, m/z 551.42, found 552.41 (M + H)$^+$. 

**Synthesis of Compound 6: 1-((4-Amino-1-(2-hydroxy-2-methylpropyl)-1H-imidazo[4,5-c]quinolin-2-yl)methyl)-3-(3-(dimethylamino)propyl)-1,2-diethylguanidine**

To a solution of trifluoroacetate salt of 2 (50 mg, 0.092 mmol) in anhydrous DMF, were added triethylamine (51 mg, 0.51 mmol), EDCI.HCl (53 mg, 0.28 mmol) and a catalytic amount of DMAP. The reaction was stirred for 24-30 hours. The solvent was then removed under vacuum, and the residue was purified using C$_{18}$ reverse-phase column chromatography to obtain the compound 6 (9 mg, 24%). $^1$H NMR (400 MHz, MeOD) $\delta$ 8.48 (d, $J = 8.4$ Hz, 1H), 7.84 (d, $J = 8.3$ Hz, 1H), 7.54 (t, $J = 7.6$ Hz, 1H), 7.38 (t, $J = 7.8$ Hz, 1H), 6.73 (t, $J = 7.2$ Hz, 1H), 5.18 (s, 2H), 4.85 – 4.71 (m, 2H), 3.58 (q, $J = 7.3$ Hz, 2H), 3.54 – 3.41 (m, 4H), 3.27 – 3.18 (m, 2H), 2.90 (d, $J = 3.7$ Hz, 6H), 2.27 – 2.00 (m, 2H), 1.30 (dt, $J = 14.4, 7.2$ Hz, 12H). $^{13}$C NMR (101 MHz, MeOD) $\delta$ 160.10, 153.79, 137.08, 134.45, 129.86, 124.93, 122.19, 118.63, 113.38, 71.07, 55.30, 54.69, 46.25, 45.16, 42.06, 41.41, 39.78, 24.50, 13.79, 11.77. MS (ESI) calculated for C$_{25}$H$_{40}$N$_8$O, m/z 468.33, found 469.33 (M + H)$^+$ and 235.17 (M + 2H)$^{2+}$. 

**Synthesis of Compound 9: tert-Butyl 2-(2-(2-hydroxyethoxy)ethoxy)ethylcarbamate**

To a solution of 7 (500 mg, 2.1 mmol) in anhydrous dichloromethane, cooled to 0 °C, were added triethylamine (420 mg, 4.2 mmol), tosyl chloride (609 mg, 3.54 mmol) and a catalytic amount of DMAP. The reaction was stirred for 16 hours and then the mixture was washed with water and dried over Na$_2$SO$_4$. The solvent was then removed under vacuum. This residue O-
tosylated compound was dissolved in anhydrous DMF, to which sodium azide (273 mg, 4.2 mmol) was added and the reaction was heated at 70 °C for 1 hour. The solvent was then removed under vacuum and the residue was dissolved in EtOAc, washed with water and dried over Na₂SO₄. The EtOAc fraction was concentrated under vacuum to obtain the compound 8 (600 mg). This 8 was then dissolved in MeOH and di-tert-butyl dicarbonate (687 mg, 3.5 mmol) was added to it. The solution was subjected to catalytic hydrogenolysis using Pd(OH)₂/C at 60 psi hydrogen pressure for 4 hours. The solution was then filtered through celite and filtrate was evaporated under reduced pressure to afford the residue, which was purified using column chromatography (3% MeOH/dichloromethane) to obtain the compound 9 (320 mg, 62%).

1H NMR (400 MHz, CDCl₃) δ 5.13 (s, 1H), 3.77 (d, J = 3.5 Hz, 2H), 3.70 – 3.61 (m, 6H), 3.58 (t, J = 5.2 Hz, 2H), 3.34 (d, J = 4.6 Hz, 2H), 2.50 (s, 1H), 1.46 (s, 9H).

13C NMR (101 MHz, CDCl₃) δ 159.62, 79.34, 72.55, 70.43, 70.29, 61.78, 40.34, 28.41. MS (ESI) calculated for C₁₁H₂₃NO₅, m/z 249.16, found 272.15 (M + Na⁺).

Synthesis of Compound 11: 1-(4-Amino-2-(((2-(2-(2-aminoethoxy)ethoxy)ethyl) (ethyl)amino)methyl)-1H-imidazo[4,5-c]quinolin-1-yl)-2-methylpropan-2-ol

To a solution of 9 (200 mg, 0.8 mmol) in 4 ml of 0.3M Dess Martin periodinane solution in dichloromethane, 10-12 drops of acetic acid were added and the reaction was stirred for 4-5 hours. The solvent was then removed under vacuum and the residue was purified using column chromatography (40-50% EtOAc/dichloromethane) to obtain the intermediate aldehyde derivative 10 (71 mg). To a solution of trifluoroacetate salt of 2 (150 mg, 0.29 mmol) in anhydrous MeOH, were added 10 (71 mg, 0.29 mmol), 6-8 drops of acetic acid, and sodium cyanoborohydride (22 mg, 0.35 mmol). The solution was stirred for 14-15 hours and the solvent was removed under vacuum to obtain the residue, which was purified using column chromatography (10% MeOH/dichloromethane) to obtain the intermediate N-Boc derivative. This was dissolved in 5 ml of trifluoroacetic acid and stirred for 30 minutes, followed by removal of the solvent by purging nitrogen and drying under vacuum to obtain the trifluoroacetate salt of the compound 11 (110 mg, 60%). 1H NMR (400 MHz, MeOD) δ 8.49 (d, J = 8.4 Hz, 1H), 7.84 (d, J = 7.4 Hz, 1H), 7.76 (t, J = 7.3 Hz, 1H), 7.63 (t, J = 7.2 Hz, 1H), 4.82 (s, 4H), 3.88 (t, J = 5.1 Hz, 2H), 3.70 (d, J = 6.5 Hz, 6H), 3.50 (s, 2H), 3.38 (s, 2H), 3.17 – 3.09 (m, 2H), 1.55 – 1.10 (m, 9H).

13C NMR (101 MHz, MeOD) δ 149.53, 148.21, 136.86, 134.61, 130.12, 122.23, 118.64, 113.15, 71.15, 69.99, 69.93, 66.50, 64.83, 55.36, 52.60, 49.74, 48.69, 46.46, 39.10, 26.24, 7.96. MS (ESI) calculated for C₂₃H₃₆N₆O₃, m/z 444.29, found 467.27 (M + Na⁺) and 223.15 (M + 2H)²⁺.

Synthesis of Compound 12: 3-(((4-Amino-1-(2-hydroxy-2-methylpropyl)-1H-imidazo[4,5-c]quinolin-2-yl)methyl)(ethyl)amino)propanenitrile

To a solution of 2 (250 mg, 0.46 mmol) in anhydrous MeOH, were added triethylamine (117 mg, 1.16 mmol) and acrylonitrile (61 mg, 1.16 mmol). The reaction was stirred for 30 hours, followed by removal of the solvent under vacuum. The residue was dissolved in EtOAc, washed with water and dried over Na₂SO₄. The EtOAc fraction was evaporated under reduced pressure and the residue was purified using column chromatography (11% MeOH/dichloromethane) to obtain the compound 12 (145 mg, 86%). 1H NMR (400 MHz, CDC₁₃) δ 8.06 (d, J = 8.4 Hz, 1H), 7.87 (d, J = 8.3 Hz, 1H), 7.55 (t, J = 7.6 Hz, 1H), 7.36 (t, J = 7.7 Hz, 1H), 6.04 (s, 2H), 4.88 (s, 2H), 4.12 (s, 2H), 2.94 (t, J = 6.7 Hz, 2H), 2.75 (q, J = 7.0 Hz, 2H), 2.54 (t, J = 6.7 Hz, 2H), 1.35 (s, 6H), 1.19 (t, J = 7.2 Hz, 3H). 13C NMR (101 MHz, CDC₁₃) δ 150.98, 150.41, 135.44, 127.81, 126.16, 122.48, 122.40, 122.18, 120.20, 118.64, 115.30, 71.49, 56.48, 50.85, 47.92, 47.64, 27.99, 15.33, 11.12. MS (ESI) calculated for C₂₀H₂₆N₆O, m/z 366.22, found 367.22 (M + H)⁺.
Synthesis of Compound 13: 1-(4-Amino-2-(((3-aminopropyl)(ethyl)amino)methyl)-1H-imidazo[4,5-c]quinolin-1-yl)-2-methylpropan-2-ol

To a solution of 12 (160 mg, 0.44 mmol) in glacial acetic acid, a catalytic amount of Pd(OH)_2/C was added and the reaction mixture was subjected to hydrogenation at 55 psi hydrogen pressure for 4 hours. The solution was then filtered. The filtrate was concentrated to obtain the acetate salt of the compound 13 (55 mg). This residue (20 mg, 0.05 mmol) was then dissolved in MeOH and di-tert-butyl dicarbonate (18 mg, 0.08 mmol) was added. After 30 minutes, the solvent was removed under vacuum and the residue was purified using column chromatography (15% MeOH/dichloromethane) to obtain the intermediate N-Boc derivative. This was then dissolved in 2 ml of HCl solution in dioxane and stirred for 2 hours. The solvent was then removed under vacuum to obtain the hydrochloride salt of the compound 13 (15 mg, 74%).

1H NMR (400 MHz, MeOD) δ 8.51 (d, J = 8.2 Hz, 1H), 7.85 (d, J = 8.3 Hz, 1H), 7.78 (t, J = 7.9 Hz, 1H), 7.65 (t, J = 7.7 Hz, 1H), 5.09 (s, 2H), 3.74 – 3.50 (m, 6H), 3.12 (d, J = 7.0 Hz, 2H), 2.32 (s, 2H), 1.51 (t, J = 6.9 Hz, 3H), 1.35 (s, 6H).

13C NMR (126 MHz, MeOD) δ 156.18, 151.90, 140.30, 137.56, 129.79, 126.47, 124.63, 123.13, 123.00, 72.46, 56.49, 53.02, 52.27, 48.77, 40.46, 27.84, 24.89, 11.74. MS (ESI) calculated for C_{20}H_{30}N_{6}O, m/z 370.25, found 371.26 (M + H)^+.

Synthesis of Compound 16: 1-(3-(((4-amino-1-(2-hydroxy-2-methylpropyl)-1H-imidazo[4,5-c]quinolin-2-yl)methyl)(ethyl)amino)propyl)-3-(3-(4-(3-aminopropylamino)butylamino)propyl)thiourea

To a solution of 13 (35 mg, 0.07 mmol) in anhydrous ethanol, were added triethylamine (22 mg, 0.21 mmol) and CS_2 (54 mg, 0.71 mmol). The reaction mixture was stirred for 30 minutes and then cooled to 0 °C. Di-tert-butyl dicarbonate (15 mg, 0.07 mmol) and a catalytic amount of DMAP dissolved in ethanol were added to the reaction mixture. After 30 minutes, the solvent was removed under vacuum and the residue was purified using column chromatography to obtain the compound 15. To a solution of 15 (10 mg, 0.02 mmol) in anhydrous dichloromethane, N^1,N^5,N^10-tris boc spermine (37 mg, 0.073 mmol) was added. The reaction mixture was stirred for 3 hours. The solvent was then removed under vacuum and the residue was purified using column chromatography (6% MeOH/dichloromethane) to obtain the intermediate N-Boc derivative. This was dissolved in 5 ml of trifluoroacetic acid and stirred for 30 minutes, followed by removal of the solvent by purging nitrogen and drying under vacuum to obtain the trifluoroacetate salt of the compound 16 (5 mg, 20%).

1H NMR (500 MHz, MeOD) δ 8.45 (d, J = 8.5 Hz, 1H), 7.83 (d, J = 8.4 Hz, 1H), 7.76 (t, J = 7.8 Hz, 1H), 7.67 – 7.56 (m, 1H), 4.96 (d, J = 6.9 Hz, 2H), 4.81 – 4.70 (m, 2H), 3.77 – 3.38 (m, 8H), 3.13 (dd, J = 18.0, 10.2 Hz, 2H), 3.10 – 2.96 (m, 8H), 2.16 – 2.04 (m, 4H), 1.97 – 1.88 (m, 2H), 1.84 – 1.74 (m, 4H), 1.44 (t, J = 7.2 Hz, 3H), 1.29 (s, 6H). 13C NMR (126 MHz, MeOD) δ 150.81, 138.54, 138.41, 135.99, 131.60, 126.50, 123.71, 120.06, 114.65, 72.55, 56.76, 52.24, 50.71, 49.86, 49.68, 48.26, 48.06, 46.25, 45.87, 37.80, 27.48, 25.39, 24.29, 9.47. MS (ESI) calculated for C_{31}H_{54}N_{10}O_{2}, m/z 614.42, found 615.43 (M + H)^+ and 308.22 (M + 2H)^{2+}.

Synthesis of Compound 19: 2-((tert-Butoxycarbonyl(undecyl)amino)acetic acid.

To a solution of 17 (200 mg, 1.6 mmol) in anhydrous MeOH, were added undecanal (218 mg, 1.28 mmol), 5-6 drops of acetic acid and sodium cyanoborohydride (111 mg, 1.76 mmol). The reaction was stirred for 3 hours and then the solvent was removed under vacuum. The residue was dissolved in dichloromethane and washed with water, brine and dried over Na_2SO_4. The dichloromethane fraction was then concentrated to obtain the residue 18, which was dissolved in MeOH, followed by the addition of triethylamine (202 mg, 2 mmol) and di-tert-butyl dicarbonate (436 mg, 2 mmol). The reaction was stirred for 4 hours and the solvent was removed under vacuum. The residue was then dissolved in a solvent mixture of THF:MeOH (3:1) and lithium hydroxide that was dissolved in water was added until the pH went above
12. The reaction was stirred for 24 hours, followed by removal of the solvent under vacuum. The residue was dissolved in water and the solution was acidified using 10% HCl until pH was less than 2. The water fraction was then extracted with EtOAc, followed by washing the EtOAc fraction with water and drying over Na$_2$SO$_4$. The EtOAc fraction was then concentrated under vacuum to obtain the compound 19 (320 mg, 61%). MS (ESI) calculated for C$_{18}$H$_{35}$NO$_4$, m/z 329.26, found 328.25 (M - H$^-$).

**Synthesis of Compound 22: 1-(4-Amino-2-((undecylamino)methyl)-1H-imidazo[4,5-c]quinolin-1-yl)-2-methylpropan-2-ol**

To a solution of 19 (171 mg, 0.52 mmol) in anhydrous DMF, were added HATU (217 mg, 0.57 mmol), triethylamine (58 mg, 0.57 mmol), a catalytic amount of DMAP and 20 (120 mg, 0.52 mmol). The reaction mixture was stirred for 3-4 hours. The solvent was evaporated under vacuum. The residue was dissolved in EtOAc, washed with water and dried over Na$_2$SO$_4$ to obtain the residue. This was dissolved in 10 mL of ethanol and a solution of NaOH (105 mg, 2.62 mmol) dissolved in 1 mL of water was added to it. The reaction mixture was refluxed for 5-6 hours, followed by evaporation under reduced pressure. The residue was purified by column chromatography (5% MeOH/dichloromethane) to obtain the compound 21 (165 mg).

To a solution of 21 in a solvent mixture of MeOH:dichloromethane:chloroform (0.1:1:1), 3-chloroperoxybenzoic acid (135 mg, 0.79 mmol) was added and the solution was refluxed at 45-50 °C for 40 minutes. The solvent was then removed and the residue was purified using column chromatography (7% MeOH/dichloromethane) to obtain the N-oxide derivative (75 mg). This was then dissolved in anhydrous dichloromethane, followed by the addition of benzoyl isocyanate (37 mg, 0.25 mmol) and heated at 45 °C for 15 minutes. The solvent was then removed under vacuum and the residue was dissolved in anhydrous MeOH, followed by the addition of excess sodium methoxide and heating at 80 °C for 1 hour. The solvent was then removed under vacuum and the residue was purified using column chromatography (8% MeOH/dichloromethane) to obtain the intermediate N-Boc derivative (27 mg). This was dissolved in 5 ml of trifluoroacetic acid and stirred for 30 minutes, followed by removal of the solvent by purging nitrogen and drying under vacuum to obtain the trifluoroacetate salt of the compound 22 (33 mg, 12%). $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 14.65 (s, 1H), 10.11 (s, 2H), 8.06 (s, 1H), 7.99 (d, $J = 8.3$ Hz, 1H), 7.44 (t, $J = 7.6$ Hz, 1H), 7.37 (t, $J = 7.8$ Hz, 1H), 7.20 (d, $J = 8.6$ Hz, 1H), 4.76 (s, 2H), 4.61 (s, 2H), 3.23 (s, 2H), 1.93 (s, 2H), 1.50 – 1.11 (m, 22H), 0.91 (t, $J = 6.7$ Hz, 3H). $^{13}$C NMR (101 MHz, CDCl$_3$) $\delta$ 149.04, 148.91, 136.14, 134.18, 129.98, 124.99, 124.25, 120.76, 119.24, 112.18, 72.07, 49.07, 44.01, 31.89, 29.55, 29.49, 29.34, 29.31, 28.98, 26.58, 25.46, 22.67, 14.10, 5.61. MS (ESI) calculated for C$_{26}$H$_{41}$N$_5$O, m/z 439.33, found 440.33 (M + H)$^+$ and 220.67 (M + 2H)$^{2+}$.

**Synthesis of Compound 24: 3-Nitroquinoline-2,4-diol**

Compound 23 (5 g, 31 mmol) was dissolved in 30 mL of nitric acid and stirred at room temperature for 10 minutes, followed by heating the reaction mixture at 75 °C for another 15 minutes. The reaction mixture was then allowed to cool down to room temperature and was added to ice-water mixture to precipitate the product. The solid, yellow precipitate was filtered and dried to obtain the compound 24 (5.9 g, 92%). $^1$H NMR (400 MHz, DMSO) $\delta$ 12.00 (s, 1H), 8.04 (d, $J = 7.9$ Hz, 1H), 7.66 (t, $J = 7.5$ Hz, 1H), 7.34 (d, $J = 8.2$ Hz, 1H), 7.28 (t, $J = 7.7$ Hz, 1H). $^{13}$C NMR (101 MHz, DMSO) $\delta$ 156.69, 156.23, 138.56, 133.61, 127.71, 124.92, 122.81, 116.32, 114.47. MS (ESI) calculated for C$_9$H$_8$N$_2$O$_4$, m/z 206.03, found 205.04 (M - H)$^+$. 

**Synthesis of Compound 25: 2,4-Dichloro-3-nitroquinoline**

Compound 24 (2 g, 9.7 mmol) was dissolved in 20 mL of phenylphosphonyl dichloride and heated at 135 °C for 3 hours. The reaction mixture was then allowed to cool down to room temperature and was added to ice-water mixture to precipitate the product. The solid, yellow precipitate was filtered and dried to obtain the compound 25 (2.1 g, 91%). $^1$H NMR (400 MHz, DMSO) $\delta$ 14.65 (s, 1H), 10.11 (s, 2H), 8.06 (s, 1H), 7.99 (d, $J = 8.3$ Hz, 1H), 7.44 (t, $J = 7.6$ Hz, 1H), 7.37 (t, $J = 7.8$ Hz, 1H), 7.20 (d, $J = 8.6$ Hz, 1H), 4.76 (s, 2H), 4.61 (s, 2H), 3.23 (s, 2H), 1.93 (s, 2H), 1.50 – 1.11 (m, 22H), 0.91 (t, $J = 6.7$ Hz, 3H). $^{13}$C NMR (101 MHz, DMSO) $\delta$ 156.69, 156.23, 138.56, 133.61, 127.71, 124.92, 122.81, 116.32, 114.47. MS (ESI) calculated for C$_9$H$_8$N$_2$O$_4$, m/z 206.03, found 205.04 (M - H)$^+$. 

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vigorously to obtain the precipitate, which was filtered and dried to afford the compound 25 (2.24 g, 74%). $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 8.30 (dd, $J = 8.4, 1.4, 0.6$ Hz, 1H), 8.14 (dd, $J = 8.5, 1.2, 0.6$ Hz, 1H), 7.97 (ddd, $J = 8.5, 7.0, 1.4$ Hz, 1H), 7.84 (ddd, $J = 8.3, 7.0, 1.2$ Hz, 1H). $^{13}$C NMR (101 MHz, CDCl$_3$) $\delta$ 146.64, 139.73, 135.62, 133.44, 129.76, 129.37, 125.22, 124.46.

**Synthesis of Compound 26: N-Benzyl-2-chloro-3-nitroquinolin-4-amine**

To a solution of 25 (2.24 g, 9.26 mmol) in 30 mL of anhydrous dichloromethane, were added triethylamine (1.4 g, 13.9 mmol) and benzylamine (1.19 g, 11.1 mmol). The reaction mixture was refluxed at 45 °C for 30 minutes. The solvent was then evaporated under vacuum and water was added to the residue to obtain the precipitate. This was filtered, washed several times with water and dried to obtain the compound 26 (2.1 g, 73%). $^1$H NMR (400 MHz, DMSO) $\delta$ 8.54 (dd, $J = 12.9, 7.3$ Hz, 2H), 7.89 – 7.81 (m, 2H), 7.69 (td, $J = 8.6, 5.0$ Hz, 1H), 7.39 – 7.21 (m, 5H), 4.46 (d, $J = 6.3$ Hz, 2H). $^{13}$C NMR (101 MHz, DMSO) $\delta$ 145.79, 144.82, 141.51, 138.27, 132.70, 129.08, 128.94, 127.89, 127.40, 127.15, 126.96, 126.52, 124.52, 120.12, 47.24. MS (ESI) calculated for C$_{16}$H$_{12}$ClN$_3$O$_2$, m/z 313.06, found 312.06 (M - H)$^+$. 

**Synthesis of Compound 27: N²-Benzyl-2-chloroquinoline-3,4-diamine**

To a solution of 26 (2.21 g, 7.06 mmol) in 20 mL of EtOAc, were added a catalytic amount of Pt/C and Na$_2$SO$_4$. The reaction mixture was subjected to hydrogenation at 55 psi hydrogen pressure for 3 hours. The reaction mixture was then filtered through celite and the filtrate was evaporated under vacuum to obtain the intermediate amide compound. This was dissolved in ethyl acetate and washed with water. The ethyl acetate fraction was dried using Na$_2$SO$_4$ and evaporated under vacuum to obtain the compound 27 (1.8 g, 90%). $^1$H NMR (400 MHz, MeOD) $\delta$ 7.90 (d, $J = 8.2$ Hz, 1H), 7.70 (d, $J = 8.4$ Hz, 1H), 7.43 (t, $J = 7.6$ Hz, 1H), 7.38 – 7.31 (m, 3H), 7.25 (t, $J = 7.5$ Hz, 2H), 7.20 (d, $J = 7.0$ Hz, 1H), 4.51 (s, 2H). $^{13}$C NMR (101 MHz, MeOD) $\delta$ 141.54, 141.28, 139.93, 137.86, 128.45, 128.04, 127.44, 126.96, 126.80, 126.22, 125.31, 123.22, 121.40, 49.56. MS (ESI) calculated for C$_{10}$H$_4$ClN$_3$, m/z 283.09, found 284.10 (M + H)$^+$. 


To a solution of 27 (50 mg, 0.18 mmol) in anhydrous THF, were added triethylamine (26 mg, 0.19 mmol) and hexanoyl chloride. The reaction mixture was stirred at room temperature for 6 hours. The solvent was then removed under vacuum, and the residue was dissolved in ethyl acetate and washed with water. The ethyl acetate fraction was dried using Na$_2$SO$_4$ and evaporated under vacuum to obtain the intermediate amide compound. This was dissolved in 0.1 ml of 2M solution of ammonia in MeOH. The sealed reaction vessel was heated in microwave at 150 °C for 1 hour. The solvent was then removed under vacuum and the residue was purified using column chromatography (7% MeOH/dichloromethane) to obtain the compound 32 (8 mg; unoptimized yields). $^1$H NMR (500 MHz, CDCl$_3$) $\delta$ 7.81 (d, $J = 8.3$ Hz, 1H), 7.72 (dd, $J = 8.3, 1.0$ Hz, 1H), 7.44 (qd, $J = 7.0, 3.5$ Hz, 1H), 7.38 – 7.29 (m, 3H), 7.18 – 7.10 (m, 1H), 7.06 (d, $J = 6.9$ Hz, 2H), 5.74 (s, 2H), 5.60 (s, 2H), 2.95 – 2.79 (m, 2H), 1.80 (dt, $J = 15.6, 7.6$ Hz, 2H), 1.42 – 1.27 (m, 4H), 0.87 (t, $J = 7.2$ Hz, 3H). $^{13}$C NMR (126 MHz, CDCl$_3$) $\delta$ 153.50, 150.28, 134.61, 133.41, 128.61, 127.40, 126.51, 125.94, 125.85, 124.85, 121.69, 119.07, 114.42, 48.20, 30.85, 26.97, 26.77, 21.65, 13.23. MS (ESI) calculated for C$_{22}$H$_{24}$N$_4$, m/z 344.20, found 345.21 (M + H)$^+$. 2D $^1$H COSY and NOESY experiments (included in Supporting Information) provided unambiguous assignment of $^1$H resonances.

**28: 1-Benzyl-2-methyl-1H-imidazo[4,5-c]quinolin-4-amine**

$^1$H NMR (500 MHz, CDCl$_3$) $\delta$ 7.81 (dd, $J = 8.4, 0.8$ Hz, 1H), 7.75 (dd, $J = 8.3, 1.0$ Hz, 1H), 7.45 (dd, $J = 8.4, 7.1, 1.4$ Hz, 1H), 7.38 – 7.28 (m, 3H), 7.15 (ddd, $J = 8.3, 7.1, 1.2$ Hz, 1H), 7.10 – 7.05 (m, 2H), 5.73 (s, 2H), 5.64 (s, 2H), 2.63 (s, 3H). $^{13}$C NMR (126 MHz, CDCl$_3$) $\delta$
29: 1-Benzyl-2-ethyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.80 (dd, J = 8.4, 0.8 Hz, 1H), 7.73 (dd, J = 8.3, 1.0 Hz, 1H), 7.44 (qd, J = 7.2, 3.7 Hz, 1H), 7.37 – 7.28 (m, 3H), 7.13 (ddd, J = 8.3, 7.0, 1.3 Hz, 1H), 7.09 – 7.01 (m, 2H), 5.73 (s, 2H), 5.59 (s, 2H), 2.97 – 2.86 (m, 2H), 1.41 (t, J = 7.5 Hz, 3H). 13C NMR (126 MHz, CDCl₃) δ 153.83, 149.91, 143.24, 134.14, 133.05, 128.19, 126.96, 126.04, 125.64, 125.52, 124.43, 121.20, 118.62, 114.05, 47.69, 19.72, 10.98. MS (ESI) calculated for C₁₈H₁₈N₄, m/z 308.15, found 303.16 (M + H)+.

30: 1-Benzyl-2-propyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.80 (dd, J = 8.4, 0.8 Hz, 1H), 7.71 (dd, J = 8.3, 1.0 Hz, 1H), 7.49 – 7.42 (m, 1H), 7.40 – 7.30 (m, 3H), 7.18 – 7.10 (m, 1H), 7.10 – 7.05 (m, 2H), 5.74 (s, 2H), 5.66 (s, 2H), 2.92 – 2.80 (m, 2H), 1.91 – 1.85 (m, 2H), 1.03 (t, J = 7.4 Hz, 3H). 13C NMR (126 MHz, CDCl₃) δ 153.50, 150.46, 143.54, 134.77, 133.58, 128.82, 127.60, 126.73, 126.12, 126.05, 125.01, 121.90, 119.30, 114.59, 48.36, 28.88, 20.92, 13.52. MS (ESI) calculated for C₁₉H₂₀N₄, m/z 316.17, found 317.17 (M + H)+.

31: 1-Benzyl-2-butyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.80 (dd, J = 8.4, 0.8 Hz, 1H), 7.72 (dd, J = 8.3, 1.0 Hz, 1H), 7.43 (ddd, J = 8.4, 7.0, 1.4 Hz, 1H), 7.36 – 7.28 (m, 3H), 7.12 (ddd, J = 8.3, 7.1, 1.2 Hz, 1H), 7.08 – 7.04 (m, 2H), 5.73 (s, 2H), 5.56 (s, 2H), 2.91 – 2.83 (m, 2H), 1.84 – 1.74 (m, 2H), 1.43 (dq, J = 14.8, 7.4 Hz, 2H), 0.92 (t, J = 7.4 Hz, 3H). 13C NMR (126 MHz, CDCl₃) δ 152.56, 149.54, 142.97, 133.87, 132.54, 127.81, 126.58, 125.61, 125.34, 125.21, 124.06, 120.78, 118.25, 113.71, 47.38, 28.54, 25.71, 21.04, 12.28. MS (ESI) calculated for C₂₀H₂₀N₄, m/z 330.18, found 331.19 (M + H)+.

33: 1-Benzyl-2-hexyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (400 MHz, CDCl₃) δ 7.82 (d, J = 8.4 Hz, 1H), 7.72 (d, J = 8.2 Hz, 1H), 7.45 (t, J = 7.7 Hz, 1H), 7.38 – 7.29 (m, 3H), 7.13 (t, J = 7.6 Hz, 1H), 7.07 (d, J = 7.4 Hz, 2H), 5.73 (s, 2H), 5.63 (s, 2H), 2.93 – 2.83 (m, 2H), 1.87 – 1.73 (m, 2H), 1.40 (dd, J = 14.3, 6.7 Hz, 2H), 1.34 – 1.24 (m, 4H), 0.88 (t, J = 6.5 Hz, 3H). 13C NMR (101 MHz, CDCl₃) δ 154.01, 151.10, 144.59, 135.38, 133.98, 129.25, 128.02, 127.02, 126.87, 126.69, 125.54, 122.17, 119.70, 115.20, 48.84, 31.44, 29.05, 27.90, 27.46, 22.46, 14.01. MS (ESI) calculated for C₂₃H₂₆N₄, m/z 358.22, found 359.22 (M + H)+.

34: 1-Benzyl-2-heptyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.81 (dd, J = 8.4, 0.8 Hz, 1H), 7.71 (dd, J = 8.3, 1.0 Hz, 1H), 7.44 (ddd, J = 8.4, 7.1, 1.3 Hz, 1H), 7.35 – 7.29 (m, 3H), 7.12 (ddd, J = 8.2, 7.1, 1.2 Hz, 1H), 7.05 (d, J = 6.9 Hz, 2H), 5.72 (s, 4H), 2.86 (dd, J = 18.2, 10.2 Hz, 2H), 1.79 (dt, J = 15.5, 7.7 Hz, 2H), 1.39 (dt, J = 14.9, 7.0 Hz, 2H), 1.34 – 1.18 (m, 6H), 0.86 (t, J = 7.0 Hz, 3H). 13C NMR (126 MHz, CDCl₃) δ 152.59, 149.22, 142.11, 133.55, 132.39, 127.57, 126.36, 125.49, 124.86, 124.69, 123.80, 120.69, 118.05, 113.31, 47.15, 29.89, 27.60, 27.19, 26.21, 25.75, 20.85, 12.34. MS (ESI) calculated for C₂₄H₂₈N₄, m/z 372.23, found 373.24 (M + H)+.

35: 1-Benzyl-2-nonyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.85 – 7.79 (m, 1H), 7.71 (dd, J = 8.3, 0.9 Hz, 1H), 7.44 (qd, J = 6.9, 3.4 Hz, 1H), 7.38 – 7.29 (m, 3H), 7.13 (ddd, J = 8.2, 7.1, 1.2 Hz, 1H), 7.05 (d, J = 6.9 Hz, 3H), 6.98 (m, 1H). 13C NMR (126 MHz, CDCl₃) δ 152.59, 149.22, 142.11, 133.55, 132.39, 127.57, 126.36, 125.49, 124.86, 124.69, 123.80, 120.69, 118.05, 113.31, 47.15, 29.89, 27.60, 27.19, 26.21, 25.75, 20.85, 12.34. MS (ESI) calculated for C₂₃H₂₆N₄, m/z 358.22, found 359.22 (M + H)+.

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36: 1-Benzyl-2-undecyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.81 (d, J = 8.2 Hz, 1H), 7.71 (d, J = 7.7 Hz, 1H), 7.47 – 7.42 (m, 1H), 7.36 – 7.28 (m, 3H), 7.15 – 7.11 (m, 1H), 7.06 (d, J = 7.0 Hz, 2H), 5.73 (s, 2H), 5.70 (s, 2H), 3.02 – 2.71 (m, 2H), 1.79 (dt, J = 15.5, 7.8 Hz, 2H), 1.34 – 1.18 (m, 14H), 0.88 (t, J = 7.0 Hz, 3H). 13C NMR (126 MHz, CDCl₃) δ 152.56, 149.23, 142.11, 133.58, 132.43, 127.60, 126.39, 125.53, 124.89, 124.73, 123.83, 120.73, 113.08, 113.34, 47.19, 30.20, 27.88, 27.73, 27.67, 27.62, 27.56, 26.24, 25.78, 20.98, 12.43. MS (ESI) calculated for C₂₈H₃₅N₄, m/z 428.29, found 429.30 (M + H)⁺.

37: 1-Benzyl-2-pentadecyl-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.81 (d, J = 8.4, 0.8 Hz, 1H), 7.72 (dd, J = 8.3, 1.0 Hz, 1H), 7.44 (ddd, J = 8.4, 7.1, 1.3 Hz, 1H), 7.36 – 7.27 (m, 3H), 7.13 (ddd, J = 8.3, 7.1, 1.2 Hz, 1H), 7.09 – 7.02 (m, 2H), 5.73 (s, 2H), 5.69 (s, 2H), 2.91 – 2.83 (m, 2H), 1.79 (dt, J = 15.5, 7.7 Hz, 2H), 1.59 – 1.33 (m, 2H), 1.33 – 1.16 (m, 22H), 0.88 (t, J = 7.0 Hz, 3H). 13C NMR (126 MHz, CDCl₃) δ 151.99, 148.66, 141.59, 133.01, 131.85, 127.03, 125.82, 124.96, 124.32, 124.17, 123.26, 120.16, 117.50, 112.78, 46.62, 29.66, 27.43, 27.40, 27.39, 27.37, 27.32, 27.17, 27.11, 27.10, 26.99, 25.68, 25.21, 20.43, 11.86. MS (ESI) calculated for C₂₈H₃₆N₄, m/z 484.36, found 485.36 (M + H)⁺.

38: 1-Benzyl-2-(but-3-enyl)-1H-imidazo[4,5-c]quinolin-4-amine

1H NMR (500 MHz, CDCl₃) δ 7.80 (d, J = 8.4, 0.8 Hz, 1H), 7.73 (dd, J = 8.3, 1.0 Hz, 1H), 7.46 – 7.42 (m, 1H), 7.36 – 7.29 (m, 3H), 7.13 (ddd, J = 8.2, 7.1, 1.2 Hz, 1H), 7.08 – 7.05 (m, 2H), 5.96 – 5.84 (m, 1H), 5.74 (s, 2H), 5.58 (s, 2H), 5.11 – 4.97 (m, 2H), 3.03 – 2.93 (m, 2H), 2.64 – 2.54 (m, 2H). 13C NMR (126 MHz, CDCl₃) δ 150.99, 148.87, 134.50, 133.06, 131.93, 127.16, 125.97, 125.02, 124.64, 124.58, 123.43, 120.17, 117.57, 113.92, 46.72, 29.59, 24.77. MS (ESI) calculated for C₂₁H₂₀N₄, m/z 328.17, found 329.18 (M + H)⁺.

Synthesis of Compound 40: N-Benzyl-3-nitroquinolin-4-amine

To a solution of 39 (2.03 mg, 9.76 mmol) in 20 mL of anhydrous dichloromethane, were added triethylamine (1.48 g, 14.64 mmol) and benzylamine (1.36 g, 12.69 mmol). The reaction mixture was refluxed at 45 °C for 30 minutes. The solvent was then evaporated under vacuum and water was added to the residue to obtain the precipitate. This was filtered, washed several times with water and dried to obtain the compound 40 (2.6 g, 95%). 1H NMR (400 MHz, CDCl₃) δ 8.98 (s, 1H), 9.42 (s, 1H), 8.31 (d, J = 8.6 Hz, 1H), 8.04 (d, J = 8.4 Hz, 1H), 7.79 (t, J = 7.6 Hz, 1H), 7.56 – 7.35 (m, 6H), 5.13 (d, J = 5.7 Hz, 2H). 13C NMR (101 MHz, CDCl₃) δ 151.01, 150.64, 147.37, 136.84, 132.73, 130.51, 129.36, 128.54, 127.16, 126.84, 126.33, 125.58, 119.14, 53.06. MS (ESI) calculated for C₁₆H₁₃N₃O₂, m/z 279.10, found 280.11 (M + H)⁺.

Synthesis of Compound 41: N⁴-Benzylquinoline-3,4-diamine

To a solution of 40 (2.6 g, 7.06 mmol) in 10 mL of EtOAc, were added a catalytic amount of Pt/C and Na₂SO₄. The reaction mixture was subjected to hydrogenation at 55 psi hydrogen pressure for 4 hours. The reaction mixture was then filtered through celite and the filtrate was evaporated under vacuum to obtain the compound 41 (2.3 g, 96%). 1H NMR (400 MHz, CDCl₃) δ 8.53 (s, 1H), 8.00 (d, J = 8.3 Hz, 1H), 7.75 (d, J = 8.2 Hz, 1H), 7.56 – 7.29 (m, 7H), 7.81 (d, J = 8.2 Hz, 1H), 7.71 (d, J = 7.7 Hz, 1H), 7.47 – 7.42 (m, 1H), 7.36 – 7.28 (m, 3H), 7.15 – 7.11 (m, 1H), 7.06 (d, J = 7.0 Hz, 2H), 5.73 (s, 2H), 5.70 (s, 2H), 3.02 – 2.71 (m, 2H), 1.79 (dt, J = 15.5, 7.8 Hz, 2H), 1.34 – 1.18 (m, 14H), 0.88 (t, J = 7.0 Hz, 3H). 13C NMR (126 MHz, CDCl₃) δ 152.56, 149.23, 142.11, 133.58, 132.43, 127.60, 126.39, 125.53, 124.89, 124.73, 123.83, 120.73, 113.08, 113.34, 47.19, 30.20, 27.88, 27.73, 27.67, 27.62, 27.56, 26.24, 25.78, 20.98, 12.43. MS (ESI) calculated for C₂₈H₃₅N₄, m/z 428.29, found 429.30 (M + H)⁺.
Synthesis of Compound 44: 1-Benzyl-2-pentyl-1H-imidazo[4,5-c]quinolin-4-amine

To a solution of pentynoic acid (37 mg, 0.38 mmol) in anhydrous DMF, were added HBTU (85 mg, 0.22 mmol), triethylamine (52 mg, 0.51 mmol), a catalytic amount of DMAP and 41 (30 mg). This was then dissolved in anhydrous dichloromethane, followed by the addition of benzoyl isocyanate (20 mg, 0.14 mmol) and heated at 45 °C for 15 minutes. The solvent was then removed under vacuum and the residue was dissolved in anhydrous MeOH, followed by the addition of excess sodium methoxide. The reaction mixture was then heated at 80 °C for 40 minutes. The solvent was then removed under vacuum. The residue was dissolved in EtOAc, washed with water, brine and then dried over Na₂SO₄. The reaction mixture was then heated in microwave at 110 °C for 1 hour. The solvent was then removed under vacuum. The residue was dissolved in EtOAc, washed with water, brine and then dried over Na₂SO₄ and then concentrated to obtain the residue. This was purified using column chromatography (7% MeOH/dichloromethane) to obtain the compound (38 mg). To a solution of 42 in a solvent mixture of MeOH:dichloromethane:chloroform (0.1:1:1), was added 3-chloroperoxybenzoic acid (83 mg, 0.48 mmol), and the solution was refluxed at 45-50 °C for 6-8 hours, followed by removal of the solvent under vacuum. The residue was dissolved in MeOH, followed by the addition of calcium oxide. The reaction mixture was then heated at 80 °C for 1 hour. The solvent was then removed under vacuum and the residue was purified using column chromatography (6% MeOH/dichloromethane) to obtain the compound (15 mg, 12%). 1H NMR (500 MHz, CDCl₃) δ 7.81 (dd, J = 8.4, 0.8 Hz, 1H), 7.74 (dd, J = 8.3, 1.0 Hz, 1H), 7.46 (ddd, J = 8.4, 5.7, 1.3 Hz, 1H), 7.37 – 7.30 (m, 3H), 7.15 (ddd, J = 8.3, 7.1, 1.2 Hz, 1H), 7.09 – 7.05 (m, 2H), 5.79 (s, 2H), 5.68 (s, 2H), 3.11 (dd, J = 8.8, 6.4 Hz, 2H), 2.87 – 2.71 (m, 2H), 1.99 (t, J = 2.6 Hz, 1H). 13C NMR (126 MHz, CDCl₃) δ 150.49, 149.42, 133.45, 132.82, 127.91, 126.75, 126.01, 125.19, 124.98, 124.08, 121.13, 118.33, 113.52, 81.16, 68.15, 47.46, 25.06, 15.63. MS (ESI) calculated for C₂₁H₁₈N₄, m/z 326.15, found 327.16 (M + H)⁺.


The compound 44 (50 mg, 0.14 mmol) was dissolved in 6M HCl and the solution was heated in microwave at 110 °C for 1 hour. The solvent was then removed under vacuum and the residue was purified using column chromatography (7% MeOH/dichloromethane) to obtain the compound (38 mg, 80%). 1H NMR (400 MHz, DMSO) δ 11.59 (s, 1H), 7.74 (d, J = 8.2 Hz, 1H), 7.43 (d, J = 8.2 Hz, 1H), 7.39 – 7.30 (m, J = 8.3 Hz, 3H), 7.30 – 7.23 (m, J = 7.2 Hz, 1H), 7.08 – 7.00 (m, J = 7.5 Hz, 3H), 5.83 (s, 2H), 2.83 (t, J = 7.5 Hz, 2H), 1.79 – 1.65 (m, 2H), 1.48 – 1.24 (m, 4H), 0.88 (t, J = 7.1 Hz, 3H). 13C NMR (101 MHz, CDCl₃) δ 156.40, 143.96, 143.60, 135.40, 134.74, 133.74, 129.93, 129.42, 128.30, 127.56, 126.58, 125.40, 119.91, 117.29, 49.19, 31.63, 27.91, 27.70, 22.29, 13.90. MS (ESI) calculated for C₂₂H₂₂ClN₃, m/z 363.15, found 364.15 (M + H)⁺.
1.40 – 1.16 (m, 4H), 0.82 (t, J = 7.0 Hz, 3H). $^{13}$C NMR (101 MHz, DMSO) δ 157.78, 154.36, 137.04, 136.73, 134.44, 131.09, 129.43, 128.08, 127.95, 125.94, 121.98, 121.27, 116.69, 112.04, 48.44, 31.22, 27.11, 26.69, 22.30, 14.26. MS (ESI) calculated for C$_{22}$H$_{23}$N$_{3}$, m/z 345.18, found 346.19 (M + H)$^+$. 

**Synthesis of Compound 47: 1-Benzyl-2-pentyl-4-phenyl-1H-imidazo[4,5-c]quinoline**

Suzuki coupling of the compound 44 with phenyl boronic acid was performed. To a solution of 44 (70 mg, 0.19 mmol) in THF, were added polystyrene-bound PPh$_3$-Pd (9.5 mg, 0.00095 mmol), K$_2$CO$_3$ (40 mg, 0.29 mmol) dissolved in water and phenyl boronic acid (28 mg, 0.23 mmol). The reaction was heated at 80 °C for 10-12 hours. The solution was filtered to remove the solid resin and the filtrate was evaporated under vacuum to obtain the residue. This was purified using column chromatography (30% EtOAc/hexane) to obtain the compound 47 (25 mg, 33%). $^1$H NMR (400 MHz, CDCl$_3$) δ 8.81 – 8.77 (m, 1H), 8.33 (dd, J = 8.5, 0.8 Hz, 1H), 7.93 (dd, J = 8.4, 0.8 Hz, 1H), 7.64 – 7.58 (m, 3H), 7.55 – 7.49 (m, 1H), 7.36 (dddd, J = 9.8, 6.9, 6.1, 2.2 Hz, 4H), 7.12 – 7.07 (m, 2H), 5.84 (s, 2H), 3.03 – 2.85 (m, 2H), 1.91 (dt, J = 15.5, 7.6 Hz, 2H), 1.42 (ddt, J = 36.4, 21.9, 7.4 Hz, 4H), 0.92 (t, J = 7.2 Hz, 3H). $^{13}$C NMR (101 MHz, CDCl$_3$) δ 155.17, 151.02, 144.50, 138.18, 135.43, 135.10, 135.01, 130.96, 130.01, 129.31, 129.27, 128.37, 128.05, 126.74, 125.76, 125.52, 119.61, 117.17, 48.92, 31.58, 27.56, 27.48, 22.37, 13.97. MS (ESI) calculated for C$_{28}$H$_{27}$N$_{3}$, m/z 405.22, found 406.23 (M + H)$^+$. 

**Synthesis of Compound 48: N-(1-Benzyl-2-butyl-1H-imidazo[4,5-c]quinolin-4-yl) hydroxylamine**

To a solution of 27 (100 mg, 0.35 mmol) in anhydrous THF, were added triethylamine (72 mg, 0.71 mmol) and valeryl chloride (51 mg, 0.42 mmol). The reaction mixture was then stirred for 6-8 hours, followed by removal of the solvent under vacuum. The residue was dissolved in EtOAc, washed with water, brine and then dried over Na$_2$SO$_4$ to obtain the residue. This was dissolved in MeOH, followed by the addition of calcium oxide. The reaction mixture was then heated in microwave at 110 °C for 1 hour. The solvent was removed under vacuum and the residue was purified using column chromatography (3% MeOH/dichloromethane) to obtain 45 (75 mg, 62%). To a solution of 45 (25 mg, 0.072 mmol) in anhydrous MeOH, were added triethylamine (72 mg, 0.72 mmol) and hydroxylamine hydrochloride (76 mg, 1.08 mmol). The reaction mixture was heated at 50 °C for 10-12 hours. The solvent was then removed under vacuum and the residue was dissolved in EtOAc, washed with water and dried over Na$_2$SO$_4$. The EtOAc fraction was then evaporated under vacuum and the residue was purified using basic-alumina (pH>7) column chromatography (30% MeOH/dichloromethane) to obtain the compound 48 (10 mg, 40%). $^1$H NMR (500 MHz, CDCl$_3$) δ 7.41 (d, J = 8.1 Hz, 1H), 7.32 (dq, J = 25.0, 7.2 Hz, 4H), 7.21 (s, 1H), 7.06 (d, J = 7.1 Hz, 2H), 6.88 (dd, J = 10.6, 5.7 Hz, 1H), 5.61 (s, 2H), 2.84 – 2.77 (m, 2H), 1.78 (dt, J = 15.5, 7.7 Hz, 2H), 1.40 (dt, J = 15.0, 7.4 Hz, 2H), 0.90 – 0.86 (m, 3H). $^{13}$C NMR (126 MHz, CDCl$_3$) δ 151.78, 133.18, 127.13, 125.87, 125.13, 123.30, 119.10, 117.94, 46.50, 27.68, 24.81, 20.34, 11.58. MS (ESI) calculated for C$_{21}$H$_{22}$N$_{4}$O, m/z 346.18, found 347.19 (M + H)$^+$. 

**Synthesis of Compound 49: 1-Benzyl-2-butyl-4-hydrazinyl-1H-imidazo[4,5-c]quinoline**

To a solution of 45 (60 mg, 0.17 mmol) in anhydrous MeOH, was added tert-butyl carbazate (57 mg, 0.43 mmol). After 15 hours, the solvent was removed under vacuum and the residue was purified using column chromatography (30% MeOH/dichloromethane) to obtain the intermediate N-Boc derivative. This was then dissolved in 5 ml of trifluoroacetic acid and stirred for 30 minutes, followed by removal of the solvent by purging nitrogen and drying under vacuum to obtain the trifluoroacetate salt of the compound 49 (55 mg, 57%). $^1$H NMR (500 MHz, CD$_3$OD) δ 7.82 (d, J = 8.4 Hz, 1H), 7.79 (d, J = 8.3 Hz, 1H), 7.45 – 7.40 (m, 1H), 7.28 (dt, J = 25.5, 7.2 Hz, 3H), 7.15 – 7.10 (m, 1H), 7.03 (d, J = 7.4 Hz, 2H), 5.82 (s, 2H), 2.94 –
2.90 (m, 2H), 1.79 – 1.69 (m, 2H), 1.40 (dq, $J = 14.8, 7.4$ Hz, 2H), 0.90 (t, $J = 7.4$ Hz, 3H). $^{13}$C NMR (126 MHz, MeOD) $\delta$ 154.65, 151.25, 143.19, 135.64, 133.54, 128.81, 128.68, 127.41, 126.96, 125.31, 125.10, 124.68, 120.02, 114.47, 48.23, 29.28, 26.26, 21.84, 12.51. MS (ESI) calculated for C$_{21}$H$_{23}$N$_5$, m/z 345.20, found 346.20 (M + H)$^+$. 

**Synthesis of Compound 50: 1-Benzyl-2-butyl-1H-imidazo[4,5-c]quinoline**

To a solution of 41 (300 mg, 1.221 mmol) in anhydrous THF, were added triethylamine (183 mg, 1.82 mmol) and valeryl chloride (174 mg, 1.45 mmol). The reaction mixture was then stirred for 6-8 hours, followed by removal of the solvent under vacuum. The residue was dissolved in EtOAc, washed with water, brine and then dried over Na$_2$SO$_4$ to obtain the intermediate amide compound. This was dissolved in MeOH, followed by the addition of calcium oxide. The reaction mixture was then heated in microwave at 110 °C for an hour. The solvent was removed under vacuum and the residue was purified using column chromatography (9% MeOH/dichloromethane) to obtain the compound 50 (360 mg, 94%). $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 9.38 (s, 1H), 8.25 (d, $J = 8.1$ Hz, 1H), 7.93 (d, $J = 8.5$ Hz, 1H), 7.66 – 7.57 (m, 1H), 7.42 (dd, $J = 11.2, 4.1$ Hz, 1H), 7.40 – 7.31 (m, 3H), 7.06 (d, $J = 6.6$ Hz, 2H), 5.80 (s, 2H), 3.02 – 2.85 (m, 2H), 1.90 (dt, $J = 15.4, 7.7$ Hz, 2H), 1.57 – 1.39 (m, 2H), 0.96 (t, $J = 7.4$ Hz, 3H). $^{13}$C NMR (101 MHz, CDCl$_3$) $\delta$ 155.81, 144.98, 144.75, 136.53, 135.14, 134.10, 130.83, 129.35, 128.15, 126.72, 126.29, 125.47, 119.88, 117.58, 48.94, 29.57, 27.18, 22.53, 13.77. MS (ESI) calculated for C$_{21}$H$_{21}$N$_3$, m/z 315.17, found 316.18 (M + H)$^+$. 

**Synthesis of Compound 52: 1-Benzyl-2-((ethylamino)methyl)-1H-imidazo[4,5-c]quinolin-4-amine**

To a solution of 2-(tert-butoxycarbonyl(ethyl)amino)acetic acid (90 mg, 0.44 mmol) in anhydrous DMF, were added HBTU (167 mg, 0.44 mmol), triethylamine (53 mg, 0.52 mmol), a catalytic amount of DMAP and 41 (100 mg, 0.4 mmol). The reaction mixture was then heated to 70 °C for 18 hours. The solvent was then removed under vacuum. The residue was dissolved in EtOAc, washed with water, dried using Na$_2$SO$_4$ and then concentrated to obtain the residue, which was purified using column chromatography (6% MeOH/dichloromethane) to obtain the compound 51 (133 mg). To a solution of 51 in a solvent mixture of MeOH:dichloromethane:chloroform (0.1:1:1), 3-chloroperoxybenzoic acid (138 mg, 0.8 mmol) was added and the solution was refluxed at 45-50 °C for 40 minutes. The solvent was then removed under vacuum and the residue was purified using column chromatography (10% MeOH/dichloromethane) to obtain the N-oxide derivative (31 mg). This was then dissolved in anhydrous dichloromethane, followed by the addition of benzoyl isocyanate (16 mg, 0.11 mmol) and heated at 45 °C for 15 minutes. The solvent was then removed under vacuum and the residue was dissolved in anhydrous MeOH, followed by the addition of excess sodium methoxide and heated at 80 °C for 1 hour. The solvent was then removed under vacuum and the residue was purified using column chromatography (8% MeOH/dichloromethane) to obtain the intermediate N-Boc derivative. This was dissolved in 5 ml of trifluoroacetic acid and stirred for 30 minutes, followed by removal of the solvent by purging nitrogen and drying under vacuum to obtain the trifluoroacetate salt of the compound 52 (26 mg, 20%). $^1$H NMR (400 MHz, MeOD) $\delta$ 8.02 (d, $J = 8.4$ Hz, 1H), 7.82 (d, $J = 8.4$ Hz, 1H), 7.69 (t, $J = 7.8$ Hz, 1H), 7.49 – 7.32 (m, 4H), 7.15 (d, $J = 7.6$ Hz, 2H), 6.00 (s, 2H), 4.71 (s, 2H), 3.38 (q, $J = 7.3$ Hz, 2H), 1.45 (t, $J = 7.3$ Hz, 3H). $^{13}$C NMR (101 MHz, MeOD) $\delta$ 149.52, 148.45, 136.86, 134.46, 134.03, 130.18, 129.15, 128.12, 125.29, 125.15, 124.97, 121.76, 118.34, 112.47, 48.94, 43.17, 42.36, 10.06. MS (ESI) calculated for C$_{20}$H$_{21}$N$_5$, m/z 331.18, found 332.19 (M + H)$^+$. 

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Synthesis of Compound 54: 1-(4-Amino-2-butyl-1H-imidazo[4,5-c]quinolin-1-yl)-2-methylpropan-2-ol

To a solution of 20 (100 mg, 0.43 mmol) in anhydrous THF, were added triethylamine (66 mg, 0.65 mmol) and valeryl chloride (62 mg, 0.52 mmol). The reaction mixture was then stirred for 6-8 hours, followed by removal of the solvent under vacuum. The residue was dissolved in EtOAc, washed with water, brine and then dried over Na$_2$SO$_4$ to obtain the intermediate amide compound. This was dissolved in MeOH, followed by the addition of calcium oxide and was heated in microwave at 110 °C for 1 hour. The solvent was then removed and the residue was purified using column chromatography (9% MeOH/dichloromethane) to obtain the compound 53 (58 mg). To a solution of 53 in a solvent mixture of MeOH:dichloromethane:chloroform (0.1:1:1), was added 3-chloroperoxybenzoic acid (84 mg, 0.49 mmol) and the solution was refluxed at 45-50 °C for 40 minutes. The solvent was then removed and the residue was purified using column chromatography (20% MeOH/dichloromethane) to obtain the compound 55 (58 mg). This was then dissolved in anhydrous dichloromethane, followed by the addition of benzoyl isocyanate (39 mg, 0.26 mmol) and heated at 45 °C for 15 minutes. The solvent was then removed under vacuum and the residue was dissolved in anhydrous MeOH, followed by the addition of excess sodium methoxide. The reaction mixture was then heated at 80 °C for an hour. The solvent was removed under vacuum and the residue was purified using column chromatography (11% MeOH/dichloromethane) to obtain the compound 54 (40 mg, 30%).

$^1$H NMR (400 MHz, MeOD) δ 8.26 (d, $J = 8.3$ Hz, 1H), 7.69 (d, $J = 8.4$ Hz, 1H), 7.47 (t, $J = 7.7$ Hz, 1H), 7.31 (t, $J = 7.7$ Hz, 1H), 4.63 (s, 2H), 3.20 – 3.02 (m, 2H), 1.98 – 1.80 (m, 2H), 1.62 – 1.45 (m, 2H), 1.27 (s, 6H), 1.04 (t, $J = 7.4$ Hz, 3H).

$^{13}$C NMR (101 MHz, MeOD) δ 156.33, 151.22, 143.80, 134.53, 126.75, 125.41, 125.12, 121.49, 120.93, 115.31, 71.16, 54.80, 29.72, 27.12, 26.52, 22.24, 12.86. MS (ESI) calculated for C$_{18}$H$_{24}$N$_4$O, m/z 312.20, found 313.20 (M + H)$^+$.  

Synthesis of Compound 55: 1-Benzyl-4-chloro-1H-[1,2,3]triazolo[4,5-c]quinoline

To a solution of 27 (60 mg, 0.21 mmol) in 1.5 ml of 1:1 acetic acid/water at 0 °C, was added 0.5N aqueous solution of sodium nitrite (0.56 ml, 0.28 mmol) and the reaction was stirred for 2 hours. The solvent was then removed under vacuum. The residue was dissolved in EtOAc, washed with water and dried over Na$_2$SO$_4$. The EtOAc fraction was then concentrated under vacuum to obtain the compound 55 (58 mg, 94%). $^1$H NMR (500 MHz, CDCl$_3$) δ 8.12 (dd, $J = 8.4$, 0.7 Hz, 1H), 7.96 (d, $J = 8.4$, 1H), 7.47 (t, $J = 7.7$ Hz, 1H), 7.31 (t, $J = 7.7$ Hz, 1H), 4.63 (s, 2H), 3.20 – 3.02 (m, 2H), 1.98 – 1.80 (m, 2H), 1.62 – 1.45 (m, 2H), 1.27 (s, 6H), 1.04 (t, $J = 7.4$ Hz, 3H). $^{13}$C NMR (101 MHz, CDCl$_3$) δ 156.33, 151.22, 143.80, 134.53, 126.75, 125.41, 125.12, 121.49, 120.93, 115.31, 71.16, 54.80, 29.72, 27.12, 26.52, 22.24, 12.86. MS (ESI) calculated for C$_{16}$H$_{11}$ClN$_4$, m/z 294.07, found 317.06 (M + Na$^+$) and 612.11 (M + M + Na$^+$).

Synthesis of Compound 56: 1-Benzyl-1H-[1,2,3]triazolo[4,5-c]quinolin-4-amine

Compound 55 (46 mg, 0.16 mmol) was dissolved in 0.1 ml of 2M ammonia solution in MeOH and the reaction was heated in microwave at 150 °C for 1 hour. The solvent was then removed under vacuum and the residue was purified using column chromatography (65% EtOAc/hexane) to obtain the compound 56 (35 mg, 82%). $^1$H NMR (500 MHz, TFA) δ 8.25 – 8.13 (m, 1H), 7.87 (d, $J = 7.2$ Hz, 1H), 7.79 (d, $J = 8.0$ Hz, 1H), 7.63 (t, $J = 6.1$ Hz, 1H), 7.39 (s, 3H), 7.24 (s, 2H), 6.38 (d, $J = 23.6$ Hz, 2H). $^{13}$C NMR (126 MHz, TFA) δ 134.33, 133.27, 131.52, 129.18, 129.06, 127.08, 125.83, 123.58, 118.48, 117.54, 115.29, 113.04, 110.78, 54.87. MS (ESI) calculated for C$_{16}$H$_{11}$N$_5$, m/z 275.12, found 276.12 (M + H)$^+$. 

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To a solution of 41 (30 mg, 0.12 mmol) in anhydrous THF, were added triethylamine (15 mg, 0.14 mmol) and n-propyl isocyanate (33 mg, 0.39 mmol); the n-propylamine released as a consequence of cyclization en route to the cyclic urea was found to consume n-propyl isocyanate, necessitating the use of additional equivalents of the reagent. The reaction was heated in microwave at 100 °C for 1 hour. The solvent was then removed under vacuum and the residue was purified using column chromatography (6% MeOH/dichloromethane) to obtain the compound 57 (26 mg, 61%).

1H NMR (500 MHz, CDCl3) δ 9.90 (s, 1H), 8.79 (s, J = 5.4 Hz, 1H), 8.15 (d, J = 8.5, 0.6 Hz, 1H), 7.91 (d, J = 8.6, 0.6 Hz, 1H), 7.60 (t, J = 8.4, 6.9, 1.3 Hz, 1H), 7.40 (t, J = 8.3, 6.9, 1.2 Hz, 1H), 7.37 – 7.32 (m, 2H), 7.31 – 7.28 (m, 1H), 7.26 (d, J = 6.7, 5.4 Hz, 2H), 5.60 (s, 2H), 3.49 – 3.45 (m, J = 7.1, 5.8 Hz, 2H), 1.78 – 1.68 (m, 2H), 1.04 (t, J = 7.4 Hz, 3H).

13C NMR (126 MHz, CDCl3) δ 151.08, 148.67, 143.44, 136.25, 132.69, 128.54, 127.08, 127.02, 125.87, 125.74, 124.61, 118.43, 117.87, 112.78, 44.45, 39.68, 20.57, 9.22. MS (ESI) calculated for C21H20N4O2, m/z 360.16, found 361.17 (M + H)+.

NF-κB induction

The induction of NF-κB was quantified using HEK-Blue-7 cells and HEK-Blue-8 cells as previously described by us.8,24 HEK293 cells were stably transfected with human TLR7 (or human TLR8), MD2, and secreted alkaline phosphatase (sAP), and were maintained in HEK-Blue™ Selection medium containing zeocin and nornocin. Stable expression of secreted alkaline phosphatase (sAP) under control of NF-κB/AP-1 promoters is inducible by the TLR7 (or TLR8) agonists, and extracellular sAP in the supernatant is proportional to NF-κB induction. HEK-Blue cells were incubated at a density of ~10^5 cells/ml in a volume of 80 μl/well, in 384-well, flat-bottomed, cell culture-treated microtiter plates until confluency was achieved, and subsequently graded concentrations of stimuli. sAP was assayed spectrophotometrically using an alkaline phosphatase-specific chromogen (present in HEK-detection medium as supplied by the vendor) at 620 nm.

IFN-α induction in whole human blood

Aliquots (3 mL) of heparin-anticoagulated blood obtained from healthy human donors after informed consent and stimulated for 12 h with graded concentrations of test compounds. The plasma was isolated by centrifugation, diluted 1:20, and IFN-α was assayed in triplicate using a high-sensitivity human IFN-α-specific ELISA kit (PBL Interferon Source, Piscataway, NJ).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This work was supported by NIH/NIAID contract HHSN272200900033C.

References


Abbreviations

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<td>AP-1</td>
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<td>CD</td>
<td>cluster of differentiation</td>
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<td>DC</td>
<td>dendritic cells</td>
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<td>DMAP</td>
<td>4-dimethylaminopyridine</td>
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<td>DMF</td>
<td>N,N-dimethylformamide</td>
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<tr>
<td>EC₅₀</td>
<td>half-maximal effective concentration</td>
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<td>EDCI.HCl</td>
<td>1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride</td>
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<td>ELISA</td>
<td>enzyme linked immunosorbent assay</td>
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<td>ESI-TOF</td>
<td>electrospray ionization-time of flight</td>
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<table>
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<td>O-benzotriazole-N,N,N',N'-tetramethyl uronium hexafluorophosphate</td>
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<td>HEK</td>
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<td>interferon</td>
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<td>TLR</td>
<td>Toll-like receptor</td>
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Figure 1.
Structures of Imiquimod and 2.
Figure 2.
Alkyl chain length dependence of TLR7-agonistic activity profiles of the N^1-benzyl-C2-alkyl compounds (right panel). The activity of 2 (reference compound) is shown on the left. Error bars represent standard deviations obtained on quadruplicate samples.
Figure 3.
Dose-dependent induction of IFN-α in whole human blood by 2, 31, and 34. Plasma IFN-α was assayed in triplicate by ELISA.
Figure 4.
TLR7-agonistic activities of active compounds in a human TLR7-specific reporter gene assay. Error bars represent standard deviations obtained on quadruplicate samples.
Figure 5.
Correlation of TLR7-agonistic potency and hydrophobicity (measured as retention time on C18 reverse-phase chromatography).
Scheme 1.
Syntheses of 2-imidazoyl sidechain-modified analogues of 2.

Reagents: i. CH₂CHO, NaCNBH₃, MeOH; ii. CH₃COCl, Et₃N, THF; iii. C₁₅H₂₉COCl, Et₃N, THF; iv. EDCl, Et₃N, DMF.

Reagents: i. (a) TsCl, Et₃N, DMAP, CH₂Cl₂, 0°C (b) NaN₃, DMF, 70°C; ii. (Boc)₂O, Pd(OH)$_₂$/C, H₂, MeOH; iii. Dess-martin periodinane, CH₂Cl₂; iv. (a) NaCNBH₃, MeOH (b) CF₃CO₂H.
Scheme 2.
Syntheses of 2-imidazolyl sidechain-modified analogues of 2.
Scheme 3.
Syntheses of 1-benzyl-2-(alkyl)-1H-imidazo[4,5-c]quinolin-4-amines.

Reagents: i. HNO₃, 75°C; ii. PhPOCl₂, 135°C; iii. benzylamine, Et₃N, CH₂Cl₂; iv. Pt/C, H₂, Na₂SO₄, EtOAc; v. (a) RCOCl, Et₃N, THF (b) NH₃/MeOH, 150°C.
Scheme 4.
Syntheses of 4-substituted-1-benzyl-2-(alkyl)-1H-imidazo[4,5-c]quinolines.

Reagents: i. (a) RCOCl, Et$_3$N, THF (b) CaO, MeOH, 110° C; ii. 6N aq. HCl, 110° C; iii. polystyrene-bound PPh$_3$-Pd, phenyl boronic acid, K$_2$CO$_3$, THF/H$_2$O; iv. NH$_2$OH.HCl, Et$_3$N, MeOH, 50° C; v. (a) t-butyl carbazate, 80° C, MeOH (b) CF$_3$CO$_2$H, vi. (a) RCOCl, Et$_3$N, THF (b) CaO, MeOH, 110° C.
Scheme 5.
Syntheses of C-2/N-1 substituent-swapped 31/2 hybrids.

Reagents: i. HBTU, 2-(tert-butoxycarbonyl(ethyl)amino)acetic acid, Et3N, DMAP, DMF, 70° C; ii. (a) 3-chloroperoxybenzoic acid, CH2Cl2, CHCl3, MeOH (b) benzoyl isocyanate, CH2Cl2 (c) NaOCH3, MeOH (d) CF3CO2H; iii. (a) C4H9COCl, Et3N, THF (b) CaO, MeOH, 110° C; iv. (a) 3-chloroperoxybenzoic acid, CH2Cl2, CHCl3, MeOH (b) benzoyl isocyanate, CH2Cl2 (c) NaOCH3, MeOH.
Scheme 6.
Syntheses of imidazole-modified analogues.

Reagents: i. NaNO₂, CH₃CO₂H, H₂O, 0°C; ii. NH₃/MeOH, 150°C; iii. n-Propyl isocyanate, Et₃N, 100°C, THF.
Table 1
EC₅₀ values (μM) of compounds in human TLR7-specific reporter gene assay.

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**TLR7-Agonistic Activity (μM)**

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