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## Interactions of *Bordetella pertussis* adenylyl cyclase toxin CyaA with calmodulin mutants and calmodulin antagonists: Comparison with membranous adenylyl cyclase I

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### Abstract

The adenylyl cyclase (AC) toxin CyaA from *Bordetella pertussis* constitutes an important virulence factor for the pathogenesis of whooping cough. CyaA is activated by calmodulin (CaM) and compromises host defense by excessive cAMP production. Hence, pharmacological modulation of the CyaA/CaM interaction could constitute a promising approach to treat whooping cough, provided that interactions of endogenous effector proteins with CaM are not affected. As a first step toward this ambitious goal we examined the interactions of CyaA with wild-type CaM and four CaM mutants in which most methionine residues were replaced by leucine residues and studied the effects of the CaM antagonists calmidazolium, trifluoperazine and N-(6-aminohexyl)-5-chloro-1-naphthalenesulfonamide (W-7). CyaA/CaM interaction was monitored by CaM-dependent fluorescence resonance energy transfer (FRET) between tryptophan residues in CyaA and 2'-(N-methylantraniloyl)-3'-deoxy-adenosine 5'-triphosphate and catalytic activity. Comparison of the concentration/response curves of CaM and CaM mutants for FRET and catalysis revealed differences, suggesting a two-step activation mechanism of CyaA by CaM. Even in the absence of CaM, calmidazolium inhibited catalysis, and it did so according to a biphasic function. Trifluoperazine and W-7 did not inhibit FRET or catalysis. In contrast to CyaA, some CaM mutants were more efficacious than CaM at activating membranous AC isoform I. The slope of CyaA activation by CaM was much steeper than of AC1 activation. Collectively, the two-step activation mechanism of CyaA by CaM offers opportunities for pharmacological intervention. The failure of classic CaM inhibitors to interfere with CyaA/CaM interactions and the different interactions of CaM mutants with CyaA and AC1 point to unique CyaA/CaM interactions.

## Keywords

*Bordetella pertussis*; Adenylyl cyclase; Calmodulin; Fluorescence spectroscopy; Calmodulin antagonists

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## 1. Introduction

CyaA is a 188 kDa exotoxin secreted from *Bordetella pertussis* with a N-terminal AC domain of a ~400 amino acids and a C-terminal hemolysin domain of ~1300 amino acids which is also responsible for toxin insertion into host cells [1–4]. CyaA is an important virulence factor for the pathogenesis of whooping cough. Following insertion into the plasma membrane of host cells, CyaA binds CaM with high affinity. CaM induces a conformational change in CyaA, resulting in very high cAMP formation rates [1–4]. As a result, the function of immune cells is compromised, and the infection with *Bordetella pertussis* becomes more severe and prolonged.

Recently, despite wide-spread vaccination programs, increasing numbers of whooping cough outbreaks have been noted in geographical regions ranging from Ireland to New Zealand and in diverse sociocultural settings including military schools and university laboratories [5–10]. These findings show that whooping cough is still an important medical problem and accordingly, novel therapeutic approaches are required. One strategy to treat whooping cough is to block the function of CyaA by inhibitors. Three sites of intervention can be envisaged. First, one could target the catalytic site [4,11,12]. Indeed, recently, we have developed a novel class of Bis-MANT-nucleotides that inhibit CyaA with high selectivity relative to mammalian ACs [13]. Second, one could target CyaA/CaM interaction [14], and third, one could target other allosteric sites in CyaA that are important for the conformational changes during activation [15]. When targeting CyaA/CaM interaction, one should keep in mind that CaM regulates numerous mammalian cell effector proteins including myosin light-chain kinase, sarcoplasmic Ca<sup>2+</sup> ATPase, phosphodiesterases, nitric oxide synthases and mammalian AC isoforms 1 and 8 [16–24].

The specific aims of our present study were first to better understand the CyaA/CaM interaction and second to examine the effects of CaM inhibitors on CyaA/CaM interactions. To achieve the first aim, we compared the interactions of wild-type CaM with several CaM mutants in which most of the methionine residues were replaced by leucine residues (Table 1). The methionine–leucine exchange results only in minimal conformational changes in CaM [25]. To achieve the second aim, we examined the effects of calmidazolium, W-7 and trifluoperazine, three classic CaM inhibitors (Fig. 1) [16–18,26]. As first methodological approach, we studied conformational changes in CyaA by monitoring the fluorescence of 2'-MANT-3'-d-ATP. Upon binding of CaM to CyaA, the fluorescence of 2'-MANT-3'-d-ATP increases substantially as a result of an interaction of the MANT group with phenylalanine 306 [12]. Moreover, there is a strong CaM-dependent FRET from tyrosine and tryptophan residues of CyaA to the MANT group [12]. As second methodological approach, we examined the catalytic activity of CyaA in a radiometric assay [12]. In order to paradigmatically assess the specificity of the interaction of CyaA with CaM, we also examined the effects of CaM and CaM mutants on the catalytic activity of recombinant AC1.

## 2. Materials and methods

### 2.1. Materials

Lyophilized CaM-WT from bovine brain (purity > 99%), calmidazolium chloride and W-7 were from Merck (Darmstadt, Germany). The N-terminal catalytic domain of CyaA (CyaA-N), containing the amino acids 1–373 was expressed in *E. coli* with plasmid pEx-CyaA-N. CyaA-N was then purified as described [17]. Expression and purification of the four CaM mutants (CaM-206, -213, -214 and -215) were performed as described [25]. 2'-MANT-3'-d-ATP was obtained from Jena Bioscience (Jena, Germany). [ $\alpha$ - $^{32}$ P]ATP (3000 Ci/mmol) was purchased from Hartmann (Braunschweig, Germany). 3-Isobutyl-1-methylxanthine was from Sigma-Aldrich (Taufkirchen, Germany). Sources of all other materials were as described before [12,13,27,28]. AC1 was expressed in Sf9 insect cell membranes as described [27,28].

### 2.2. Direct fluorescence and fluorescence resonance energy transfer (FRET) experiments

All fluorescence measurements were recorded on a Varian Cary Eclipse fluorescence spectrometer (Varian, Inc., Palo Alto, CA, USA) essentially as described [12]. Briefly, experiments were performed at 25 °C. Reaction buffer consisted of 100 mM KCl, 100  $\mu$ M CaCl<sub>2</sub>, 5 mM MnCl<sub>2</sub> and 75 mM HEPES/NaOH, pH 7.4. Steady-state measurements were recorded in the scan mode from  $\lambda_{em} = 300$  to 500 nm with  $\lambda_{ex} = 280$  nm and from  $\lambda_{em} = 380$  to 540 nm with  $\lambda_{ex} = 350$  nm. In the case of CaM-mutants experiments the cuvette contained first 49 ml of reaction buffer. 2'-MANT-3'-d-ATP, CyaA-N and CaM were added stepwise each in a volume of 7  $\mu$ l (final concentration were 300 nM each). The cuvette content was mixed after each addition, and final volume was 70  $\mu$ l.

Steady-state measurements with CaM inhibitors were recorded as described for the CaM mutants. First, the cuvette contained 2'-MANT-3'-d-ATP (final concentration 300 nM), CyaA-N (final concentration 300 nM) and CaM (final concentration 100 nM). The CaM concentration was chosen as low as possible, to ensure that all CaM binding sites were occupied with inhibitor. Finally, the CaM inhibitors (calmidazolium, trifluoperazine and W-7) were added at final concentrations ranging from 100 nM to 10  $\mu$ M. Measurements were first carried out in the kinetic mode to ensure steady-state conditions. In these experiments the fluorescence emission spectra were corrected by subtracting baseline fluorescence (buffer alone) and the baseline-corrected CaM inhibitor-dependent emission of each concentration of the inhibitor (buffer + inhibitor). The final DMSO concentration was kept constant at 1% (v/v).

Time-dependent experiments were conducted in the kinetic modus. For the determination of the EC<sub>50</sub> and E<sub>max</sub> values the cuvette initially contained a volume of 70  $\mu$ l with following content: 56  $\mu$ l reaction buffer, 7  $\mu$ l 2'-MANT-3'-d-ATP (final concentration 300 nM) and 7  $\mu$ l CyaA-N (final concentration 300 nM). After 3 min one  $\mu$ l of either CaM-WT or one of the mutants CaM-206, CaM-213, CaM-214 or CaM-215 (final concentration 10 nM) were added. Then, consecutively another 1  $\mu$ l of the different CaM concentrations were added to the cuvette. In each case seven CaM concentration steps (10 nM, 20 nM, 45 nM, 70 nM, 120 nM, 220 nM and 320 nM) were analyzed. Equilibration time was always 3 min. For CaM-WT, it would have been desirable to use longer equilibration times than 3 min because equilibration was slower than with CaM mutants. However, since prolonged UV irradiation of samples beyond 24 min resulted in progressive decline of fluorescence, reflecting denaturation of CyaA and CaM, we needed to find a technically feasible compromise.

To explore the binding of CaM to CyaA-N over the time, experiments were carried out by sequential addition of reaction buffer, 300 nM 2'-MANT-3'-d-ATP, 300 nM CyaA-N and

final 300 nM CaM-WT, CaM-206, CaM-213, CaM-214 or CaM-215. Binding kinetics were analyzed by the one-phase exponential association equation in GraphPad prism 5.04 software. Baseline fluorescence (reaction buffer + 2'-MANT-3'-dATP + CyaA-N) and initial time before the addition of CaM was subtracted.

In competition experiments the binding of CaM to CyaA-N was disrupted by successive addition of increasing calmidazolium concentrations (1–10  $\mu\text{M}$ ). In this case the system was additionally excited at  $\lambda_{\text{ex}} = 350 \text{ nm}$  in order to analyze the changes in the direct fluorescence emission of 2'-MANT-3'-d-ATP at  $\lambda_{\text{em}} = 350 \text{ nm}$ .

### 2.3. AC activity assay

AC activity was determined essentially as described [12]. Briefly, assay tubes contained 10  $\mu\text{l}$  of CaM-WT, CaM-206, CaM-213, CaM-214 or CaM-215 at final concentrations ranging from 10 pM to 1  $\mu\text{M}$  to obtain saturated curves. Tubes were preincubated for 2 min at 25 °C after the addition of 20  $\mu\text{l}$  reaction mixture. This reaction mixture consisted of (final) 100 mM KCl, 5 mM free  $\text{Mn}^{2+}$ , 10  $\mu\text{M}$  free  $\text{Ca}^{2+}$ , 100  $\mu\text{M}$  EGTA, 100  $\mu\text{M}$  cAMP, 40  $\mu\text{M}$  ATP and [ $\alpha$ - $^{32}\text{P}$ ]ATP (0.2  $\mu\text{Ci}/\text{tube}$ ). Free concentrations of divalent cations were calculated with WebMax C standard ([http:// www.stanford.edu/cpatton/webmaxcE.htm](http://www.stanford.edu/cpatton/webmaxcE.htm)). The preincubation was carried out to ensure the formation of a steady-state equilibrium of complexing agents and divalent cations. Reactions were started by the addition of 20  $\mu\text{l}$  of CyaA-N. Final toxin concentration was 10 pM CyaA-N. Tubes were incubated for 15 min at 25 °C.

For the analysis of the CaM inhibitor potency, the CaM concentration was chosen as low as possible, to ensure that all CaM binding sites were occupied with inhibitor but guarantee that the catalytic activity was not too low. CaM concentration was kept at 1 nM, whereas the concentrations of calmidazolium, trifluoperazine and W-7 ranged from 1 nM to 10  $\mu\text{M}$ . After the addition of 20  $\mu\text{l}$  of CyaA-N (final concentration 10 pM) the tubes were preincubated for 2 min at 25 °C. Reactions were initiated by the addition of 20  $\mu\text{l}$  reaction mixture. Reaction time was 15 min at 25 °C. The lipophilic compounds calmidazolium and W-7 were dissolved in dimethyl sulfoxide (DMSO) and in all experiments with inhibitors the final DMSO concentration was kept constant at 1% (v/v).

For the evaluation of direct effects of the CaM inhibitors on CyaA-N, experiments were performed in the absence of CaM and  $\text{Ca}^{2+}$ , respectively. Without the activation of CaM the catalytic activity of CyaA-N is about 10,000 times lower [29]. Therefore, the final toxin concentration was increased to 100 pM and thus, the [ $\alpha$ - $^{32}\text{P}$ ]ATP concentration was increased as well (0.8  $\mu\text{Ci}/\text{tube}$ ). In these experiments, reaction time was 30 min at 37 °C. Concentrations of calmidazolium, trifluoperazine and W-7 ranged from 1 nM to 100  $\mu\text{M}$ .

The effect of CaM and CaM mutants on the activity of AC1 in Sf9 insect cell membranes was determined essentially as described [27,28]. Briefly, Sf9 membranes expressing AC1 (20  $\mu\text{g}/\text{tube}$ ) were pre-incubated for 2 min in the absence and the presence of CaM-WT or CaM mutants (10 nM–10  $\mu\text{M}$ ) at 30 °C. Reactions were initiated by the addition of reaction mixture containing 100  $\mu\text{M}$  3-isobutyl-1-methylxanthine, 400  $\mu\text{M}$  EDTA, 5 mM  $\text{MgCl}_2$ , 10  $\mu\text{M}$  free  $\text{Ca}^{2+}$ , 40  $\mu\text{M}$  ATP, 0.25  $\mu\text{Ci}$  [ $\alpha$ - $^{32}\text{P}$ ]ATP, 100  $\mu\text{M}$  cAMP, 0.4 mg/ml creatine kinase and 9 mM phosphocreatine. Reactions were conducted for 20 min at 30 °C.

### 2.4. Molecular modeling

Relationships between the positions of mutated CaM methio-nine residues and the CaM/CyaA interaction were probed in silico by analyzing the structure of the *Bordetella pertussis* AC toxin (PDB: 1YRT) [30] via the SYBYL software (version 8.1, Tripos, Inc., St. Louis MO, USA, 2009) to which protons had been added according to physiological pH (i.e.,

cationic arginine and lysine residues; anionic aspartates and glutamates). Using SYBYL's distance computation tool, each relevant CaM methionine was analyzed according to whether or not it coupled closely with CyaA (i.e., if it had any atoms within 4.0Å of any CyaA atom). Secondary structure assignments specified within the crystal structure were used to identify whether any CaM methionine was located within a CaM secondary structure element (i.e., was further than one residue away from the start or end of a specific a helix,  $\beta$  strand or turn).

## 2.5. Statistical analysis

Time courses of CyaA activation by CaM and CaM mutants, concentration–response curves for the stimulatory effects of CaM and CaM mutants on FRET and catalytic activity of CyaA and AC1 were analyzed by non-linear regression using the software Prism 5.04 (GraphPad Prism, San Diego, CA, USA). The effects of CaM mutants versus CaM at CyaA were compared using one-way analysis of variance with a Dunnett's multiple comparison post test. In case of the analysis of AC1, the effects of CaM and CaM mutants were compared with each other using one-way analysis of variance with Bonferroni multiple comparison post test.

## 3. Results

### 3.1. Analysis of the interaction of CyaA with CaM-WT and CaM mutants in steady-state FRET and direct fluorescence experiments

At an excitation wavelength of 280 nm, 2'-MANT-3'-d-ATP showed only minimal fluorescence (Fig. S1). In contrast, CyaA, bearing numerous tyrosine and tryptophan residues [12,13,30], showed substantial fluorescence with an emission maximum of 350 nm. Upon addition to CyaA, both CaM-WT and CaM mutants caused a significant FRET from tyrosine and tryptophan residues to the MANT group of 2'-MANT-3'-d-ATP as reflected by new emission peaks with a maximum at 430 nm. The FRET signals were of very similar magnitude for all CaM proteins. However, we noted that for the CaM mutants, but not for CaM-WT, the endogenous tyrosine and tryptophan fluorescence of CyaA was increased, indicating that CaM-WT and CaM mutants stabilize different CyaA conformations.

At a wavelength of 350 nm, the MANT group of 2'-MANT-3'-d-ATP is excited and gives rise to a substantial fluorescence signal with an emission peak at 450 nm (Fig. S2). The addition of CyaA did not increase the fluorescence signal, but upon addition of CaM-WT or CaM mutants, a substantial fluorescence increase was noted, in addition to a blue-shift of the emission maximum to 430 nm, reflecting effective interaction of the MANT group with phenylalanine 306 [12]. Again, no large differences between CaM-WT and CaM mutants were apparent.

### 3.2. Kinetic analysis of the interaction of CyaA with CaM and CaM mutants in FRET experiments

Fig. S3 shows representative time courses for the stimulatory effects of CaM-WT and CaM mutants on FRET, Fig. 2 shows merged time courses, and Table 2 summarizes the results of the non-linear regression analysis of kinetics. While CaM-WT and CaM mutants reached a similar maximum FRET, association proceeded in the order CaM-215 > CaM-214 > CaM-206 > CaM-213 > CaM.

### 3.3. Concentration/response curves for the interactions of CyaA with CaM-WT and CaM mutants in FRET experiments

Fig. S4 shows representative cumulative concentration/response curves for the stimulatory effects of CaM-WT and CaM mutants on FRET, Fig. 3 shows the results of merged

concentration/response curves and Table 3 summarizes the results of the nonlinear regression analysis of the curves. CaM-WT and CaM mutants increased FRET according to sigmoidal concentration/response curves with very similar maximum effects and EC<sub>50</sub> values. No statistically significant differences between CaM-WT and CaM mutants were apparent.

#### 3.4. Concentration/response curves for the interactions of CyaA with CaM-WT and CaM mutants in AC assays

Fig. 4 shows the results of merged concentration/response curves for the stimulatory effects of CaM-WT and CaM mutants on AC activity, and Table 4 summarizes the results of the non-linear regression analysis of the curves. CaM-WT and CaM mutants increased catalysis according to sigmoidal concentration/response curves. All CaM mutants were significantly less potent than CaMWT at activating catalysis. Moreover, CaM-214 and CaM-215 were significantly less efficacious than the other CaM proteins at stimulating cAMP formation.

#### 3.5. Effects of CaM antagonists on FRET, direct fluorescence and AC activity of CyaA in the presence and the absence of CaM

Calmidazolium, trifluoperazine and W-7 are classic CaM antagonists [16–18,26]. However, trifluoperazine and W-7 failed to inhibit CaM-stimulated FRET (Fig. S5). Calmidazolium up to 1  $\mu$ M also failed to reduce FRET, and only at the exceedingly high concentration of 10  $\mu$ M, about four orders of magnitudes higher than the affinity for CaM [26], did calmidazolium partially reduce FRET. The three CaM antagonists revealed only partial inhibitory effects on direct fluorescence (Fig. S5).

Trifluoperazine and W-7 failed to inhibit CaM-stimulated catalysis of CyaA, and calmidazolium inhibited cAMP formation with an IC<sub>50</sub> of 0.66  $\mu$ M (Fig. 5A). In the absence of CaM, the catalytic activity of CyaA is very low [29], but when the experimental conditions are adjusted accordingly, nonetheless, catalytic activity can be readily measured (see Section 2). In the absence of CaM, trifluoperazine and W-7 up to 10  $\mu$ M had no effect on catalysis. Only at the highest concentration (100  $\mu$ M) did the two inhibitors a slight inhibitory effect. In contrast, calmidazolium inhibited the catalytic activity of CyaA in the absence of CaM according to a biphasic function with a high-affinity IC<sub>50</sub> of 340 nM and a low-affinity IC<sub>50</sub> of 34  $\mu$ M.

#### 3.6. Concentration/response curves for the interactions of AC1 with CaM-WT and CaM mutants in AC assays

Fig. S6 shows the absolute activities of AC1 with increasing concentrations of CaM-WT and CaM mutants. For all CaM proteins, shallow concentration/response curves with Hill slopes <1.0 were obtained (CaM-WT, 0.34; CaM-206, 0.61; CaM-213, 0.51; CaM-214, 0.32; CaM-215, 0.43). For none of the CaM proteins, saturation was reached at a concentration of up to 10  $\mu$ M. Fig. 6A shows that there were no significant differences in potency between CaM-WT and CaM mutants. However, we noticed differences in the magnitude of the stimulatory effects of CaM and CaM mutants at a concentration of 10  $\mu$ M each (Fig. 6B and C). Specifically, CaM-214 and CaM-215 were significantly more efficacious AC1 activators than CaM-WT.

#### 3.7. Molecular modeling of the interactions of CyaA with CaM

Based on the crystal structure [30] it is possible to rationalize the observed effects that specific CaM mutations may have on the stability of the CyaA/CaM complex. Mutations to residues that directly impinge on the PPI interface and alter its steric or electrostatic character are among those likely to affect stability of the PPI complex. Mutations within

secondary structure elements that significantly alter the steric or electrostatic profile may also affect the PPI stability by disrupting the stability of those secondary structure elements. Finally, mutations that are likely to affect intramolecular stabilization interactions (e.g., salt bridges and disulfide bonds) are also potentially important in that they may indirectly influence the stability of the PPI complex by favoring a different conformations within one of more of the participating proteins.

From Fig. 7, one sees that the four CaM methionine residues under investigation in this work are all situated at the periphery of the PPI interface, and thus their mutation may be expected to have some effect on stability of the complex. However; the methionine to leucine mutation (i.e., exchanging residues of similar size, shape and electrostatic profile) does not produce a substantially different protein surfaces that would significantly compromise PPI affinity. Furthermore, methionine and leucine have similarly favorable helix-forming propensities (0.24 and 0.21, respectively) [31], so the fact that all four mutated methionine residues occur in  $\alpha$ -helices also does not suggest likely destabilization of the PPI via secondary-structure disruption. Finally, methionines and leucines do not participate in salt bridges or disulfide bonds, and have similar propensity for engaging in weaker lipophilic stabilization mechanisms, thus once again arguing for at most a modest impact on the PPI stability.

## 4. Discussion

### 4.1. Interactions of CyaA with CaM mutants: evidence for a two-step mechanism of activation

CaM is an important  $\text{Ca}^{2+}$  sensor in mammalian cells and regulates the activity of numerous effector proteins including various kinases,  $\text{Ca}^{2+}$  ATPase, ion channels and AC1 [32,33]. The amino acid sequences recognized by CaM are quite diverse [34], and several mechanisms by which CaM activates its targets, have been described [35]. The mechanism of CaM-CyaA interaction is unique. Strikingly, CyaA and another bacterial AC toxin, edema factor from *Bacillus anthracis*, interact in distinct manners with CaM [30,36]. CaM interacts with four different regions in CyaA to stabilize a catalytically active conformation [30]. Hydrophobic interactions of methionine residues 109, 124, 144 and 145 in CaM with CyaA are of particular importance for PPI as revealed by crystallographic studies (Fig. 7) [30]. This notion is supported by the fact that oxidation of methionine residues 109, 124 and 145 reduces the affinity of CyaA for CaM mutants [37].

The long-term goal of our group is the development of specific inhibitors of the catalytic activity of CyaA since CyaA contributes substantially to the pathogenesis of whooping cough, a major health problem [5–10]. One possible strategy is to target the CyaA/CaM interaction [14,37]. Accordingly, it is very important to understand in detail this PPI. Originally, already in this study, we intended to take advantage of the methionine oxidation approach, assuming that the exchange of methionine against leucine residues does not substantially affect PPIs. We focused on methionine residues 144 and 145 in CaM because these amino acids were previously shown to play an important role in CyaA/CaM interaction [37]. Our molecular modeling studies predicted that the exchange of methionine against leucine at these positions would not exhibit a major impact on PPIs (Fig. 7). However, to our surprise, the “control” experiments with unoxidized CaM mutants revealed subtle differences between individual mutants that shed light onto the molecular mechanism of CyaA/CaM interactions.

Specifically, substitution of all nine methionines in CaM-215 resulted in an around three-fold increase in the  $\text{EC}_{50}$  for CaM to activate catalysis and an approximately 15% decrease in the maximal catalytic activity of CyaA (Fig. 4 and Table 4). A loss of the first eight

methionines (CaM-206 and CaM-213) increased the  $EC_{50}$  two-fold. However, methionine side chains are not essential for the CaM-dependent activation of CyaA as shown by the largely preserved function of CaM-215 which is devoid of methionine residues (Figs. 3 and 4). Methionine 145 slowed down CaM/CyaA interaction substantially, whereas the impact of methionine 144 was moderate (Fig. 2 and Table 2). Thus, a leucine at position 145 facilitates the initial hydrophobic interactions between CaM and CyaA, probably because of the larger hydrophobic surface of leucine compared to methionine [30]. However, when the FRET signals under state-state conditions are considered, no differences in  $EC_{50}$  and efficacy between CaM and CaM mutants are apparent (Fig. 3 and Table 3). Nonetheless, when the endogenous tryptophan/tyrosine fluorescence signals of CyaA are considered it becomes evident that CaM mutants stabilize different CyaA conformations than CaM-WT (Fig. S1).

Mechanistically, dissociations in the effects of CaM-WT and CaM mutants between binding and catalysis indicate a two-step mechanism of CyaA activation. First, a complex between CaM and CyaA forms, followed by the activation of catalysis by CaM. The binding of CaM to CyaA can be readily monitored in kinetic and steady-state fluorescence experiments, whereas the nature of the second conformational step leading to catalysis cannot be monitored individually with the methods currently available to us and is still elusive. The AC assay monitors the sum of steps 1 and 2. A two-step mechanism for activation of catalysis is also supported by the CaM inhibitor experiments in which we observed dissociations in the effects on FRET, direct fluorescence and catalysis (Fig. S5 and Fig. 5).

Two-step activation of catalysis has also been proposed for mammalian membranous ACs that are activated by the diterpene forskolin. Specifically, all diterpenes studied so far increase FRET from tyrosine and tryptophan residues in membranous AC to MANT-nucleotides bound to the catalytic site, but only diterpenes substituted with a hydroxyl group at position 1 of the diterpene ring activate catalysis [27]. Thus, although CyaA and mammalian membranous ACs are structurally very different proteins as are the activation mechanisms [1–4,20,38], fundamental regulatory principles are conserved in these proteins.

#### 4.2. Effects of CaM inhibitors on CyaA-CaM interaction

Our data show that calmidazolium is the only one out of three classic CaM inhibitors studied (Fig. 1) capable of robustly inhibiting the catalytic activity of CyaA (Fig. 5). Previous studies revealed that these CaM inhibitors can exert their inhibitory effects by interacting with CaM or by modulating the CaM target structures [16–19,26,39]. In the case of CyaA, our results indicate that the inhibitory effect of calmidazolium is independent of CaM. This notion is supported by the finding that even in the absence of CaM, calmidazolium inhibits catalysis (Fig. 5B). The biphasic inhibition isotherm indicates that calmidazolium possesses two functionally or structurally distinct binding sites on CyaA.

The affinity of CaM for calmidazolium is in the nM-range, whereas for trifluoperazine and W-7 affinities in the  $\mu$ M-range are reported [26]. If one compares the structures of the three CaM inhibitors it becomes clear that calmidazolium possesses the most complex and bulky structure with its four aromatic rings (Fig. 1). It has been suggested that there is a correlation between the hydrophobicity and size of these compounds and their ability for CaM inhibition [18]. Thus, our data suggest that calmidazolium binds to one or two large hydrophobic pockets in CyaA and prevents conformational changes required for activation of catalysis. An interaction of calmidazolium with CaM does not appear to be important in this respect. A previous study showed that in intact CHO cells, calmidazolium inhibits CyaA-induced cAMP accumulation with an  $IC_{50}$  of 2.5  $\mu$ M, but it remained unclear whether these effects were CaM-dependent or not [40].

Complex CaM-independent effects of calmidazolium were also reported for mammalian membranous ACs [41]. The structurally related nitric oxide-stimulated soluble guanylyl cyclase is inhibited by calmidazolium in a CaM-independent manner, too [42]. Moreover, apparently CaM-independent effects of CaM antagonists on the superoxide-generating NADPH oxidase in intact human neutrophils were reported [43]. Thus, CaM antagonists in general and even the most potent CaM antagonist currently available, calmidazolium, are certainly not specific for CaM. The lack of selectivity of the currently available CaM antagonists for CaM severely limits their use for intact cell studies. In contrast, in studies with purified or recombinant proteins, selectivity of CaM antagonists, or lack thereof, can be readily assessed in control experiments without CaM. Our present study highlights the importance for such control experiments.

#### 4.3. Interactions of AC1 with CaM: comparison with CyaA

An important goal in pharmacologically targeting the CyaA/CaM interaction is to leave the interaction of CaM with endogenous target proteins unaffected. While it is beyond the scope of this study to comprehensively examine selectivity of, or lack thereof, CaM/target PPIs, we examined an important paradigmatic PPI, the AC1/CaM interaction. AC1 is an important AC isoform exclusively expressed in neuronal cells that play a critical role in memory, learning, neuronal damage, and pain responses [20–23,38,44,45]. Thus, in case of whooping cough treatment, AC1/CaM interaction should be left unaffected while in case of chronic pain and neuronal excitotoxicity, selective inhibition of the AC1/CaM interaction may be desirable.

Although CaM activation of AC1 has been known for more than two decades [21], relatively little efforts have been undertaken so far to understand the molecular details of AC1/CaM interaction [22,23,46] and accordingly, no targeted inhibitor studies have been performed. In marked contrast to the data obtained for CyaA, CaM mutants 214 and 215 were more effective than CaM-WT at activating catalysis. Moreover, the slope of CyaA activation by CaM was much steeper than the slope of AC1 activation (Figs. 4 and 6). Furthermore, the binding affinity of CaM for CyaA compared to AC1 was much higher. Lastly, CaM mutants showed decreases in potency relative to CaM with respect to CyaA activation but not with respect to AC1 activation. Collectively, these data demonstrate that the interactions of CyaA and AC1 with CaM are profoundly different.

Our data also suggest that the interaction of AC1 with CaM could be improved pharmacologically. CaM-215, devoid of all methionine residues, was the most effective AC1 activator. The higher efficacy of CaM mutants at AC1 compared to CaM-WT indicates that leucine substitutions facilitate hydrophobic AC1/CaM interactions, thereby enhancing catalysis. These data also render it likely that oxidation of individual methionine residues will exhibit a larger impact on AC1/CaM interactions than on CyaA/CaM interactions. Thus, even these limited “control” experiments that were originally assumed to reveal no mechanistically relevant data, unmasked important differences in the interactions of CyaA and AC1 with CaM. Based on these studies, the design of selective compounds targeting a defined AC isoform/CaM interaction is principally feasible.

#### 4.4. Concluding remarks and future directions of research

Using CaM mutants and different functional assays, we obtained evidence for a two-step mechanism of CyaA activation which is conserved among ACs. The kinetics of CyaA/CaM interactions are sensitive to change in hydrophobicity, and classic CaM inhibitors do not effectively block CyaA/CaM interactions. Rather, the most potent CaM inhibitor known so far, calmidazolium, exhibits CaM-independent direct effects on the catalytic activity of CyaA. The lack of potent inhibition of CyaA/CaM interaction by classic CaM inhibitors

suggests that the PPI in this couple are different from the interactions of other effector/CaM interactions [24,32–35]. In principle, such differences offer the opportunity to develop inhibitors that specifically target the interaction between CyaA and CaM but not between endogenous proteins and CaM, reducing potential side effects. Considering the fact that interactions of CyaA and AC1, an important mammalian target protein of CaM, with CaM and CaM mutants are different, specific inhibition of CyaA/CaM interactions is feasible.

Our present study also highlights the importance to carefully examine the properties of unoxidized CaM mutants since such proteins may differ more substantially from CaM-WT than is generally appreciated. Moreover, the detailed analysis of such mutants can be used to delineate the mechanisms of PPI interaction. Now that we have conducted a basic characterization of CaM mutants on CyaA and AC1, we are in the position to examine the impact of oxidation of individual methionine residues, using the properties of each unoxidized CaM mutant as control. In conjunction with molecular modeling studies, such studies will provide the basis for the development of specific small molecule PPI inhibitors. An advantage of targeting the AC/CaM interaction site as compared to the catalytic site of AC is the fact that tentative inhibitors of AC/CaM interactions will have to exhibit a moderate degree of lipophilicity, implying membrane-permeability and potential usefulness for intact cell studies. In contrast, while selective CyaA inhibitors targeting the catalytic site are already available [13,47], such compounds are not feasible for intact cell studies because of their lack of membrane-permeability. For the development of inhibitors of mammalian membranous ACs, we have come to the conclusion that targeting the allosteric “stimulatory” diterpene-binding site is more promising to achieve isoform-selective inhibitors than targeting the catalytic site [47]. By analogy, this may also apply to “stimulatory” AC/CaM interaction sites.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

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## Abbreviations

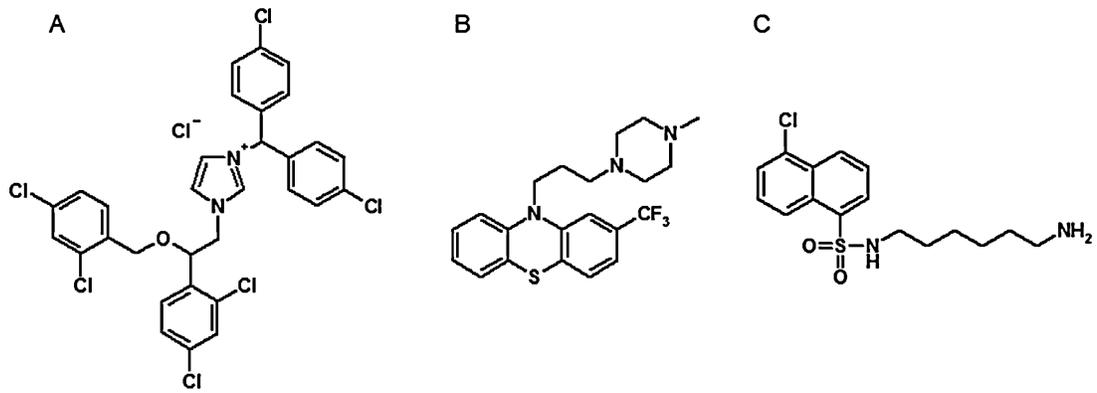
<b>AC</b>	adenylyl cyclase
<b>AC1</b>	membranous adenylyl cyclase isoform 1
<b>CyaA</b>	Bordetella pertussis adenylyl cyclase toxin
<b>CaM</b>	calmodulin
<b>CaM-WT</b>	wild-type calmodulin
<b>FRET</b>	fluorescence resonance energy transfer
<b>2'-MANT-3'-d-ATP</b>	2'-(N-methylanthraniloyl)-3'-deoxy-adenosine 5'-triphosphate
<b>PPI</b>	protein-protein interaction
<b>W-7</b>	N-(6-aminoethyl)-5-chloro-1-naphthalenesulfonamide

## References

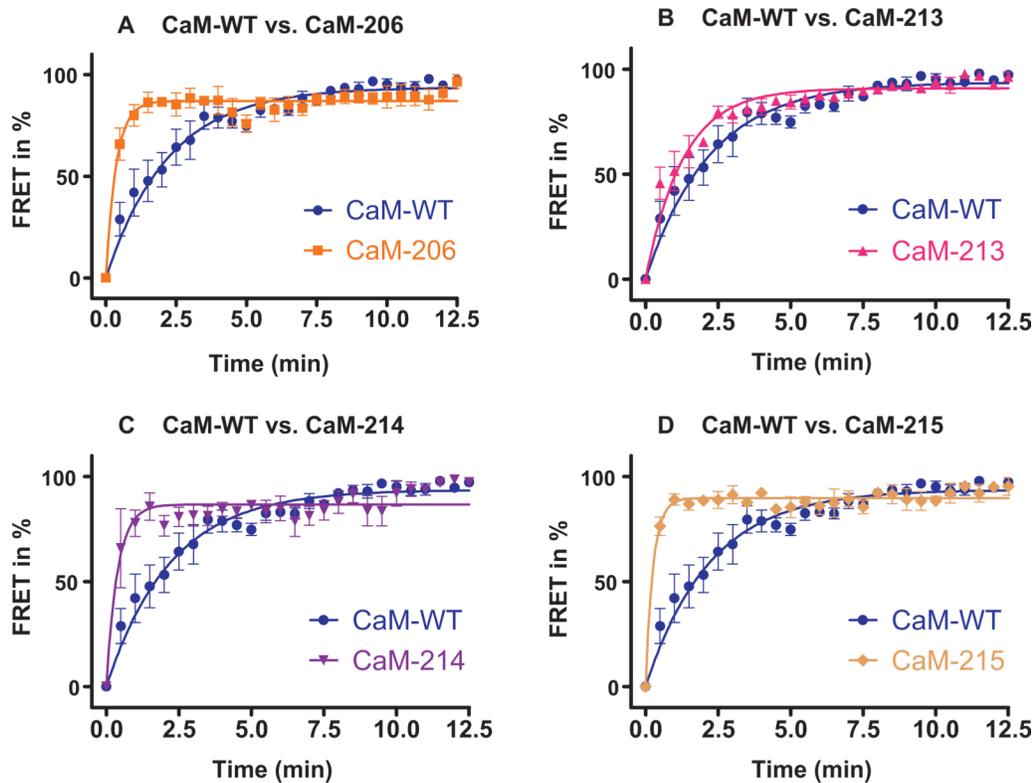
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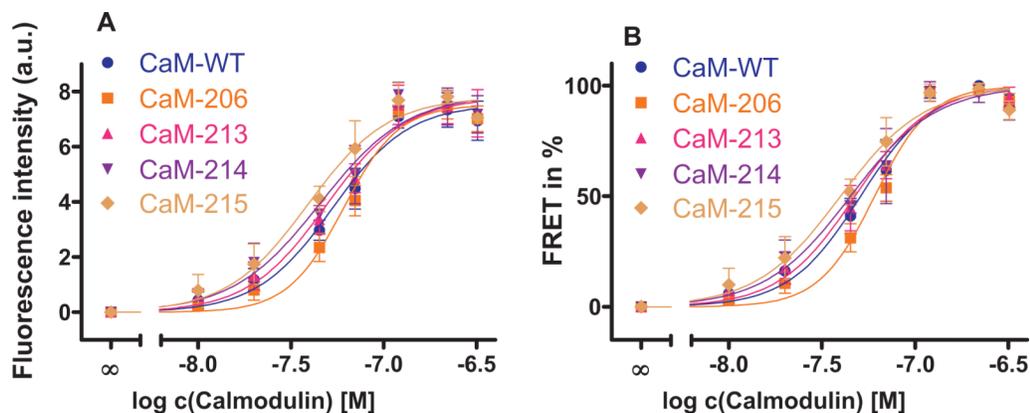
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**Fig. 1.** Structures of CaM antagonists. (A) Calmidazolium chloride; (B) trifluoperazine; and (C) W-7.

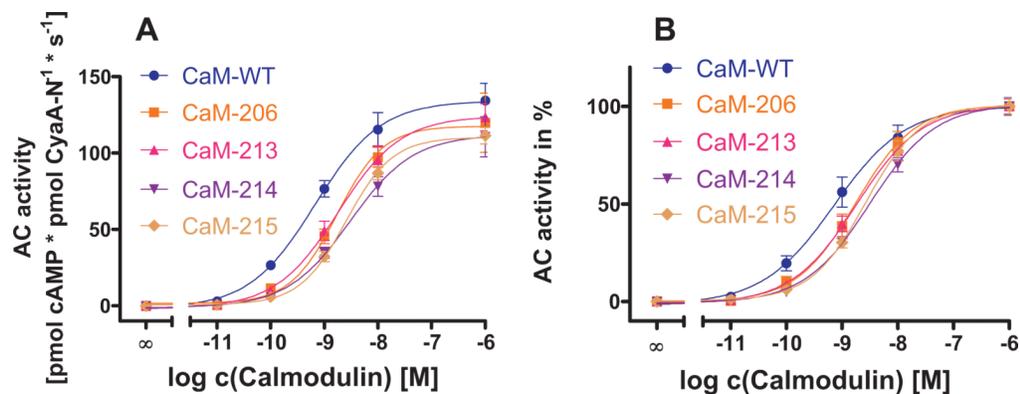


**Fig. 2.** Kinetic analysis of the binding CaM-WT and CaM mutants to CyaA-N in FRET assays. Rate of binding CaM to CyaA-N over time was determined in FRET experiments as described in Section 2. The kinetic curves of binding CaM to CyaA-N were analyzed by the one-phase exponential association equation in Prism 5.04. Data shown are the means  $\pm$  SD of three independent experiments.



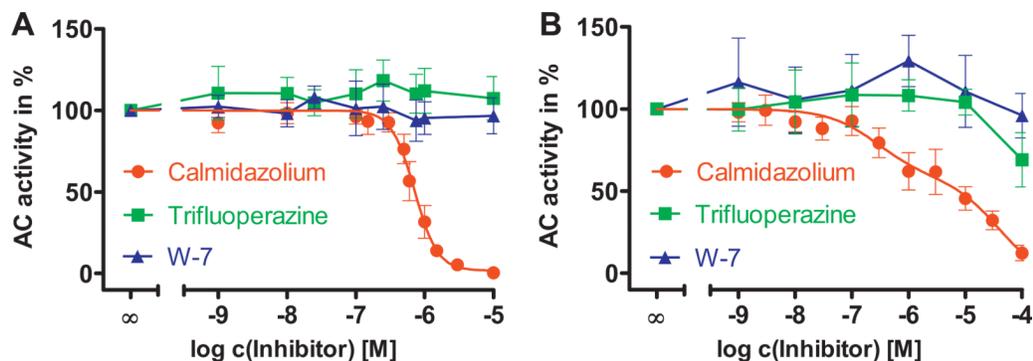
**Fig. 3.**

Concentration/response curves for the interaction of CyaA with CaM and CaM mutants in FRET assays. FRET was determined as described in Section 2. The cuvette contained 300 nM of 2'-MANT-3'-d-ATP and 300 nM of CyaA-N. For assigning a FRET emission to each CaM concentration the arithmetic average of the last ten measuring points were used. The basal fluorescence (in this case reaction buffer + 2'-MANT-3'-d-ATP + CyaA-N) was subtracted from each data point. Concentration–response curves were analyzed by nonlinear regression using Prism 5.04 software.  $r^2$  values were  $>0.96$ , except for that of CaM-214 which equaled 0.95. The plots show the means  $\pm$  SD of three independent experiments. A shows the fluorescence intensity in arbitrary units, while in B the fluorescence was normalized, whereby 100% was defined as largest value in each data set.



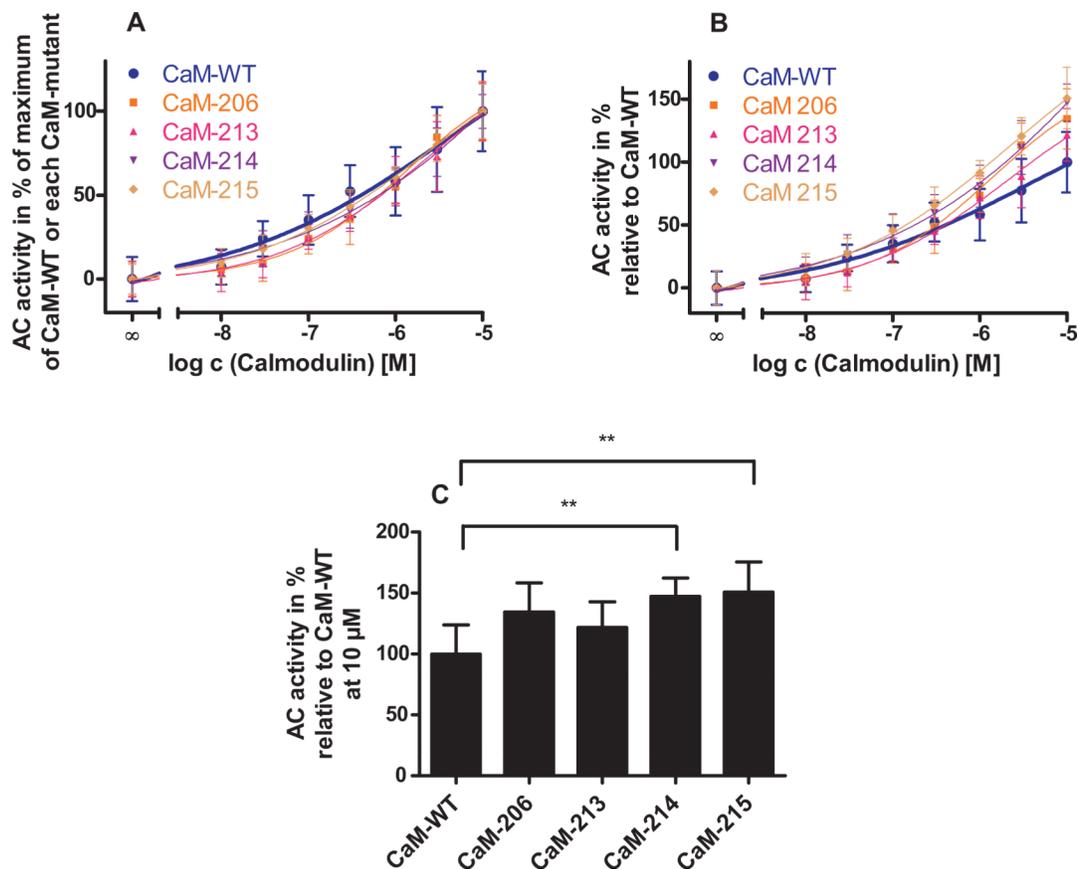
**Fig. 4.**

Concentration/response curves for the interaction of CyaA with CaM-WT and CaM mutants in AC assay. Reactions were carried out as described in Section 2. Final CyaA-N concentration was 10 pM and CaM-WT, CaM-206, CaM-213, CaM-214 or CaM-215 were added at concentrations from 10 pM to 1  $\mu$ M. Reactions were started by the addition of the toxin. Concentration–response curves were analyzed by nonlinear regression using Prism 5.04.  $r^2$  values were  $>0.97$ . Data show the means  $\pm$  SD of three independent experiments. A shows the AC activity in pmoles of cAMP formed per pmol CyaA-N per second, while in B the AC activity was normalized, whereby 100% was defined as largest value in each data set.

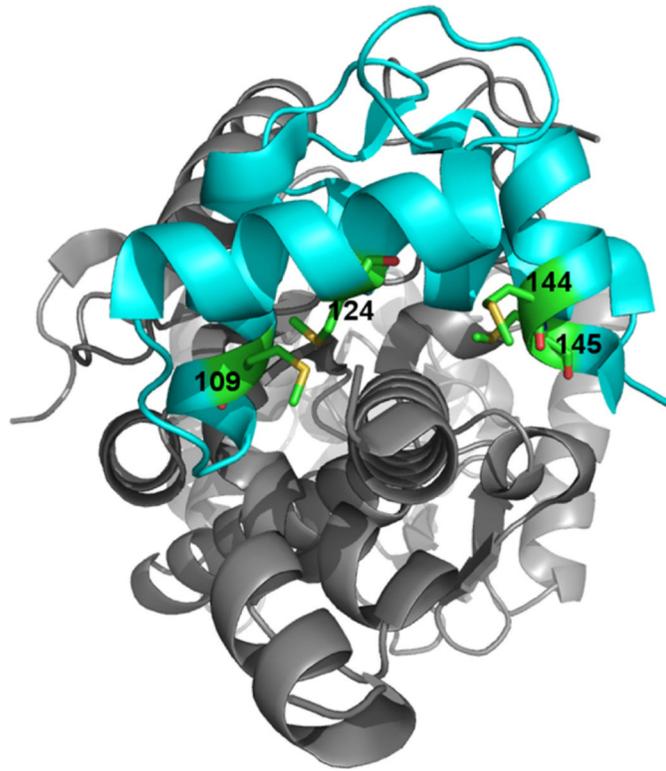


**Fig. 5.**

Effects of CaM inhibitors on the AC activity of CyaA in the presence and the absence of CaM-WT. (A) AC activity in the presence of CaM. Assays were performed as described in Section 2. Reaction mixtures contained 1 nM CaM-WT, 10 pM CyaA-N and CaM inhibitors (calmidazolium, trifluoperazine or W-7) in various concentrations (1 nM to 10  $\mu$ M). The calmidazolium inhibition curve was analyzed by nonlinear regression using Prism 5.04. Data show the means  $\pm$  SD of three independent experiments. Assays were performed as duplicate or triplicate. AC activity was normalized with respect to the initial activity in the absence of inhibitor. (B) AC activity in the absence of CaM. Assays were performed as described in Section 2. Reaction mixtures contained 100 pM CyaA, and concentrations of CaM-inhibitors (calmidazolium, trifluoperazine or W-7) ranged from 1 nM to 100  $\mu$ M. The calmidazolium inhibition curve was analyzed by nonlinear regression using Prism 5.04. Data show the means  $\pm$  SD of three independent experiments. Assays were performed as duplicate or triplicate. AC activity was normalized with respect to the initial activity in the absence of inhibitor.



**Fig. 6.** Concentration/response curves for the interaction of AC1 with CaM-WT and CaM mutants in AC assay. Reactions were carried out as described in Section 2. The concentrations of CaM and CaM mutants were varied between 10 nM–10  $\mu$ M. Concentration–response curves were analyzed by nonlinear regression using Prism 5.04. Specifically, data were compared for best fit to sigmoidal concentration/response curves or variable slope concentration/response curves, the latter providing a better fit in all cases. (A) comparison of the potencies of CaM-WT and CaM mutants with the maximum effect of each CaM protein at 10  $\mu$ M being set to 100%. B, comparison of the effects of CaM-WT and CaM mutants with the maximum effect of CaM-WT being set to 100%. C, comparison of the effects of CaM mutants at a concentration of 10  $\mu$ M each relative to CaM-WT. \*\* $p < 0.01$ . Data show the means  $\pm$  SD of three independent experiments performed in duplicates.



**Fig. 7.** Molecular modeling of the interaction of CyaA with CaM. Interaction complex of CyaA (grey ribbons) and CaM (cyan ribbons) showing the location of four crystallographically resolved methionine residues (CPK-colored sticks with green carbon atoms) that have been subjected to leucine mutational analysis. The basis of the model is the crystal structure PDB 1YRT. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of the article.)

**Table 1**

Nomenclature of analyzed CaM mutants.

CaM	Methionines	Leucine substitutions
Wild-type	36, 51, 71, 72, 76, 109, 124, 144,145	None
206	144,145	36, 51, 71, 72, 76, 109, 124
213	145	36, 51, 71, 72, 76, 109, 124, 144
214	144	36, 51, 71, 72, 76, 109, 124, 145
215	None	36, 51, 71, 72, 76, 109, 124, 144, 145

Positions of methionines and their associated site-directed leucine substitutions for wild-type CaM and indicated CaM mutants.

**Table 2**

Analysis of the binding kinetics of CaM-WT and CaM mutants to CyaA.

Calmodulin	$K$ ( $\text{min}^{-1}$ )	Half-life (min)
CaM-WT	$0.568 \pm 0.24$	$1.625 \pm 0.48$
CaM-206	$2.977 \pm 0.67$	$0.254 \pm 0.05^{**}$
CaM-213	$0.873 \pm 0.23$	$0.902 \pm 0.21$
CaM-214	$3.902 \pm 1.33^*$	$0.248 \pm 0.11^{**}$
CaM-215	$3.978 \pm 0.51^*$	$0.180 \pm 0.02^{**}$

Listed are the means and standard errors of three independent experiments. The basis for the analysis are the experiments shown in Fig. 2.  $K$  is the association rate constant expressed in  $\text{min}^{-1}$ , and half-life expressed in min is the time required for CaM to bind 50% of the available CyaA-N molecules. A one-way analysis of variance with a Dunnett's multiple comparison post test with CaM-WT as control were performed to find out significant differences in  $K$  and half-life values of the calmodulin mutants with reference to CaM-WT.

No \*:  $p$ -value  $> 0.05$ .

\*  $0.01 < p$ -value  $< 0.05$ .

\*\*  $0.001 < p$ -value  $< 0.01$ .

**Table 3**

Analysis of the concentration/response curves for the interaction of CyaA with CaM-WT and CaM mutants in FRET assays.

Calmodulin	Fluorescence resonance energy transfer	
	EC <sub>50</sub> (nM)	E <sub>max</sub> (%)
CaM-WT	52 ± 9	100
CaM-206	60 ± 6	101 ± 7
CaM-213	48 ± 9	103 ± 18
CaM-214	46 ± 3	101 ± 1
CaM-215	40 ± 8	104 ± 11

The experiments shown in Fig. 3 are the basis for the analysis. Concentration–response curves were analyzed by nonlinear regression using Prism 5.04. EC<sub>50</sub> and E<sub>max</sub> represent the means ± SD of three independent experiments. A one-way analysis of variance with a Dunnett's multiple comparison post test with CaM-WT as control were done to find out significant differences in EC<sub>50</sub> and E<sub>max</sub> values of the calmodulin mutants with reference to CaM-WT.

**Table 4**

Analysis of the concentration/response curves for the interaction of CyaA with CaM-WT and CaM mutants in the AC assay.

Calmodulin	AC activity	
	EC <sub>50</sub> (nM)	E <sub>max</sub> (%)
CaM-WT	0.8 ± 0.3	100
CaM-206	1.8 ± 0.7 *	90 ± 4
CaM-213	1.8 ± 0.2 *	93 ± 2
CaM-214	3.2 ± 0.4 ***	85 ± 8 *
CaM-215	2.7 ± 0.1 ***	84 ± 5 **

The experiments shown in Fig. 4 are the basis for the analysis. Concentration–response curves were analyzed by nonlinear regression using Prism 5.04. EC<sub>50</sub> and E<sub>max</sub> represent the means ± SD of three independent experiments. A one-way analysis of variance with a Dunnett's multiple comparison post test with CaM-WT as control were computed to find out significant differences in EC<sub>50</sub> and E<sub>max</sub> values of the calmodulin mutants with reference to CaM-WT.

No\*: *p*-value > 0.05.

\* 0.01 < *p*-value < 0.05.

\*\* 0.001 < *p*-value < 0.01.

\*\*\* *p*-value < 0.001.