

Antiproliferative withanolides from the Solanaceae: A structure–activity study*

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Abstract: As part of our search for bioactive compounds from plant biodiversity, 29 withanolides were recently isolated from three members of the Solanaceae: *Physalis longifolia*, *Vassobia breviflora*, and *Withania somnifera*. Six derivatives were prepared from these naturally occurring withanolides. All compounds were evaluated for in vitro antiproliferative activity against an array of cell lines [melanoma cell lines (B16F10, SKMEL28); human head and neck squamous cell carcinomas (HNSCC) cell lines (JMAR, MDA1986, DR081-1); breast cancer cell line (Hs578T), and non-malignant human cell line (MRC5)]. This led to the discovery of 15 withanolides, with IC₅₀ values in the range of 0.067–17.4 μM, including withaferin A, withaferin A 4,27-diacetate, 27-*O*-glucopyranosylwithaferin A, withalongolide H, withalongolide C, withalongolide A, withalongolide A 4,27-diacetate, withalongolide A 4,19,27-triacetate, withalongolide B, withalongolide B 4-acetate, withalongolide B 4,19-diacetate, withalongolide D, withalongolide E, withalongolide G, and 2,3-dihydrowithaferin A 3-*O*-sulfate. In order to update the growing literature on withanolides and their activities, we summarized the distribution, structural types, and antiproliferative activities for all published withanolides to date. The structure–activity relationship analysis (SARA) confirmed the importance of the presence of a Δ²-1-oxo-functionality in ring A, a 5β,6β-epoxy or 5α-chloro-6β-hydroxy grouping in ring B, and nine-carbon side chain with a lactone moiety for cytotoxic activity. Conversely, the SARA indicated that the –OH or –OR groups at C-4, 7, 11, 12, 14, 15, 16, 17, 18, 19, 20, 23, 24, 27, and 28 were not contributors to the observed antiproliferative activity within the systems analyzed.

Keywords: antiproliferative; *Physalis longifolia*; Solanaceae; structure classification; structure–activity relationship; *Vassobia breviflora*; *Withania somnifera*; withanolide.

INTRODUCTION

The name “withanolides” was originally given to a family of C₂₈ ergostane-type steroidal δ-lactones (derived from a parent 22-hydroxy-26-oic acid) isolated from the genus *Withania* (Solanaceae). The term “withan” refers to *Withania* and “-olide” to the presence of lactone functionality in this type of compound. With the structural elucidation of withaferin A **1** (Fig. 1) in 1965, a new area of withanolide chemistry was initiated [1]. Further investigations in this field led to the discovery of many hun-

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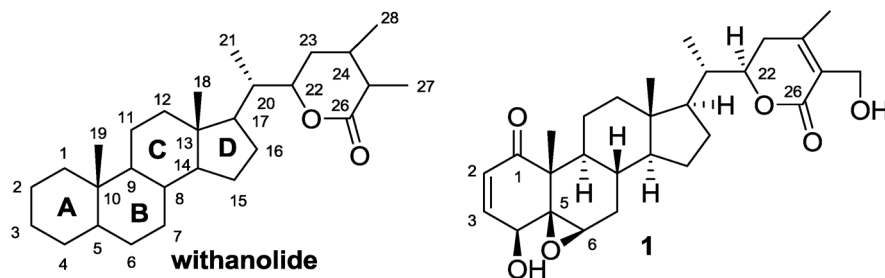


Fig. 1 Withanolide carbon-frame and structure of withaferin A 1.

dreds of additional compounds, which include 22 related but different structural types such as: (1) new withanolides exhibiting γ -lactone functionality (from 23-hydroxy-26-oic or 28-hydroxy-26-oic acids), or γ -lactol functionality (from 28-hydroxy-26-als), or δ -lactol functionality (from 22-hydroxy-26-als) in the side chain (Fig. 2); (2) new withanolides isolated from sources other than the genus *Withania* (Table 1); and (3) a number of structural varieties with novel skeletons derived or modified from, or biogenetically related to, the highly oxygenated withanolides by ring fission, cyclization, and skeleton rearrangements in either or both of the steroid nucleus and in the nine-carbon side chain [2–8]. In view of these findings, the original term “withanolide” is no longer limited to its primary definition, and it has expanded to encompass the structurally diverse variants bearing different carbon scaffolds, such as those present in acnistin, joborol, neophysalin, nicandrenone, physalin, taccalonolide, and withajardin (Fig. 2).

In recent years, withanolides have held the interest of the scientific community owing to their structural features and display of significant pharmacological activities—antimicrobial, antitumor, anti-inflammatory, immunomodulatory, and insect-antifeedant [2–4,9]. Over the last 46 years, approximately 750 withanolides, exhibiting more than 22 carbon skeletons (Fig. 2), have been reported from various plant sources. As listed in Table 1, withanolides are predominant in 25 genera (including *Acnistus*, *Datura*, *Deprea*, *Discopodium*, *Dunalia*, *Iochroma*, *Jaborosa*, *Lycium*, *Nicandra*, *Physalis*, *Solanum*, *Trechonaetes*, *Tubocapsicum*, *Vassobia*, *Withania*, and *Witheringia*) of the Solanaceae family [2–7,10–12]. Not limited to the Solanaceae, withanolides have also been isolated from a smaller number of species in the Dioscoreaceae [13], Fabaceae [7], Lamiaceae [2], Myrtaceae [14], Taccaceae [2], and from two genera in the marine Alcyoniidae [15,16]. Although the large family Solanaceae is composed of six subfamilies, with the exception of *Browallia viscosa*, all species containing withanolides belong to the subfamily Solanoideae [3,4]. Within the Solanoideae *Jaborosa*, *Physalis*, and *Withania* have been the genera most extensively studied to date. For example, more than 130 withanolides with diversified functional groups were isolated from different parts (whole plant, aerial parts, flowers, fruits, leaves, roots, and stem barks) of *Withania somnifera*, a traditional plant of wide use in Ayurvedic medicine. This plant contains the largest number of withanolides reported from any one species so far. All the withanolides from *Withania* possess only a type Ia skeleton (Fig. 2), while those reported from *Physalis* show greater variety with more than 10 structural types (Fig. 2 and Table 1).

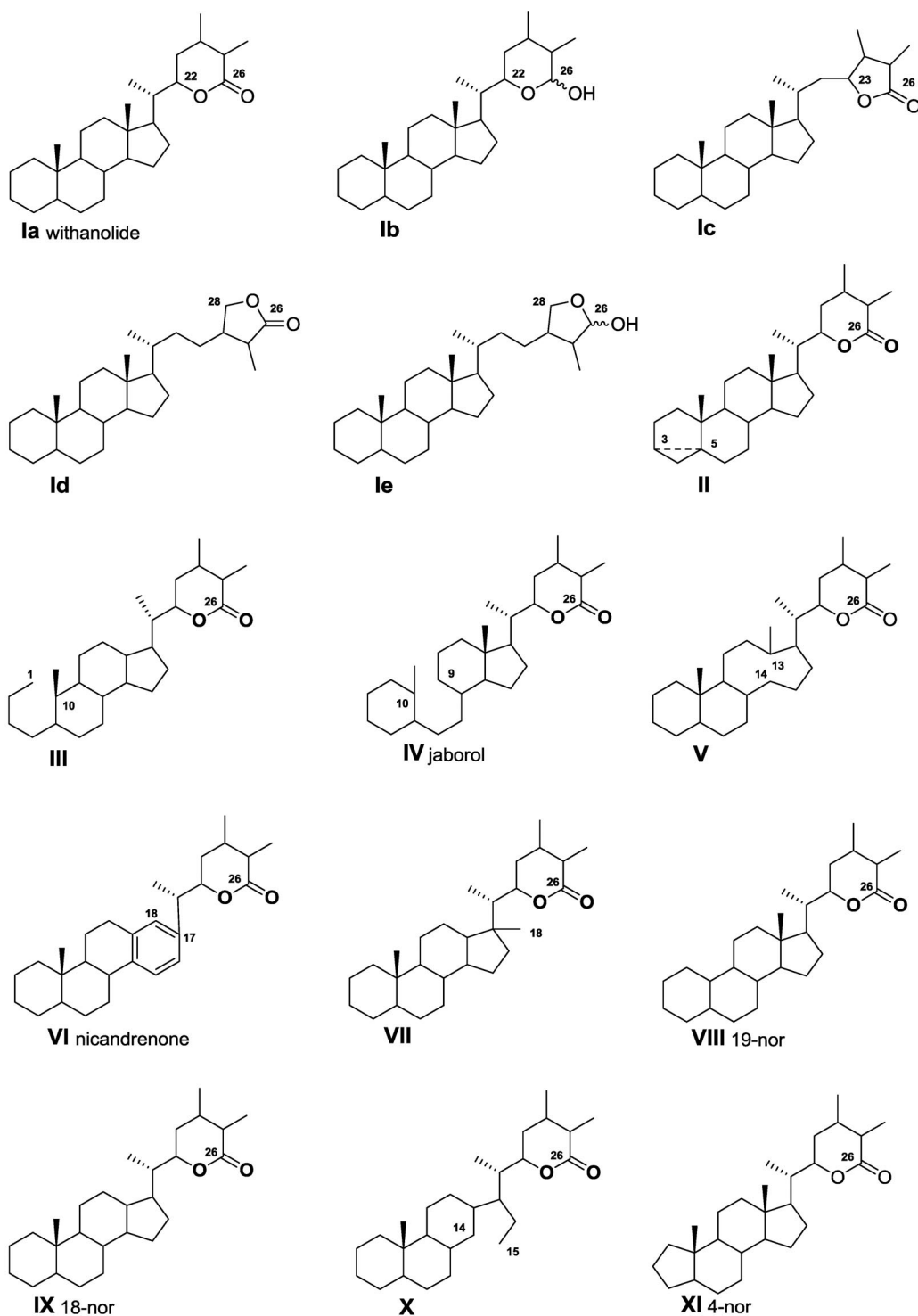


Fig. 2 Twenty-two structure types of withanolides based on their carbon skeletons (1965–2011).

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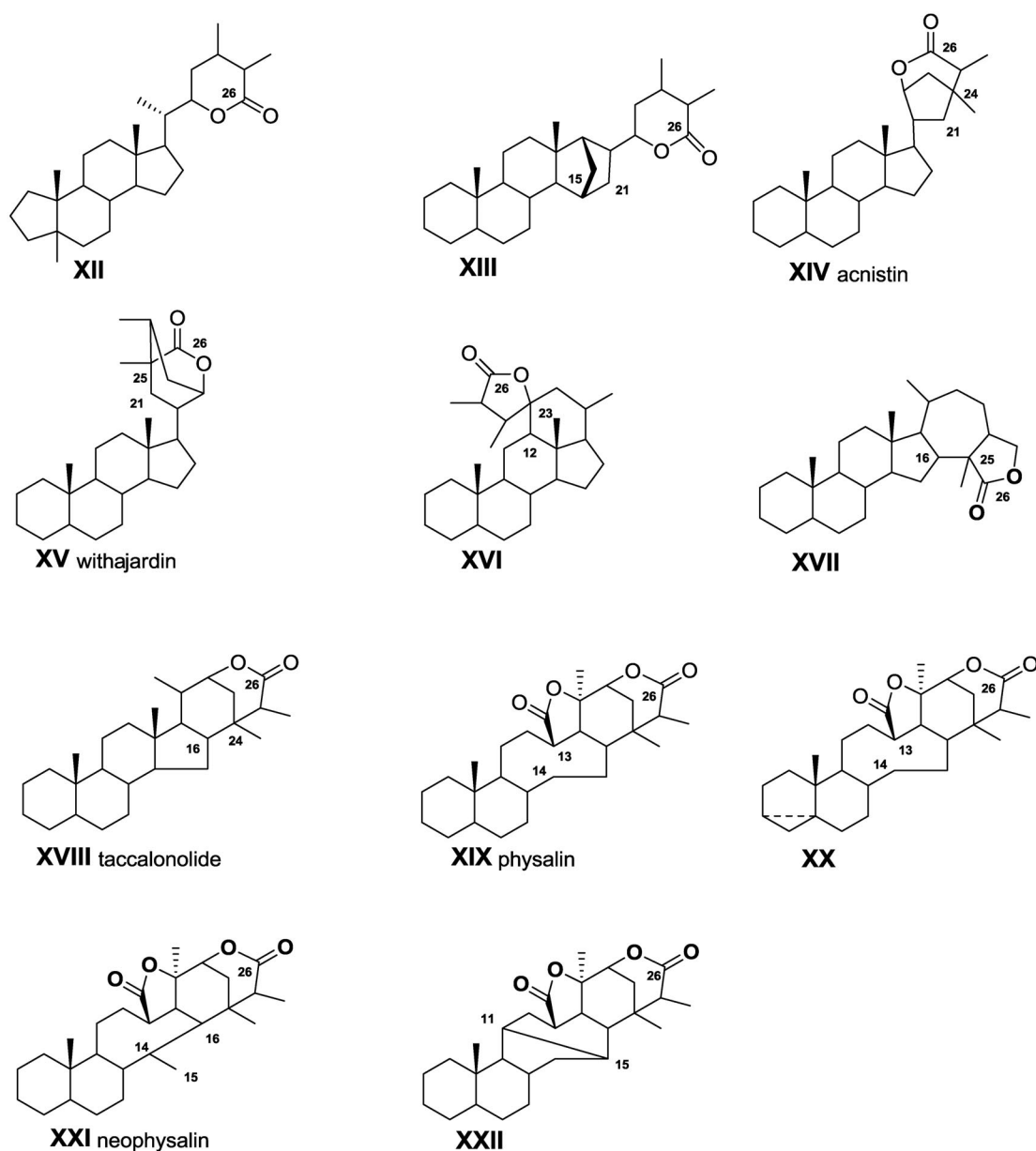


Fig. 2 (Continued).

Type I withanolides (Fig. 2), also called unmodified withanolides, are the most abundant form found in nature. With approximately 540 compounds in this category reported to date, they are distributed in the majority of the genera listed in Table 1. From the biogenetic origin, it is likely that type I compounds are precursors to the more advanced structural types II–XXII. Type I withanolides that display the most promising antiproliferative characteristics also contain Δ^2 -1-oxo-functionality in ring A, a 5 β ,6 β -epoxy-group in ring B, and a nine-carbon side chain incorporating the δ -lactone functionality found in withaferin A **1** [17], withacnistin **59** [18,19], withanolide D **48** [20–23], withanolide E **36** [8,24–27], and 4 β -hydroxywithanolide E **37** (Fig. 4) [8,24–27]. Withaferin A **1** (Fig. 1) was shown to induce apoptosis in vitro and in vivo by suppressing the growth of an array of tumor cells, including

Table 1 Sources of withanolides and their corresponding structure types.

Family	Species	Structure type
Solanaceae	1 <i>Acnistus arborescens</i> , <i>A. australis</i> , <i>A. breviflorus</i> , <i>A. lorentzii</i> , <i>A. ramiflorus</i>	I, XIV
	2 <i>Aureliana fasciculata</i> [10]	I
	3 <i>Brachistus stramonifolius</i>	I
	4 <i>Browallia viscosa</i>	I, VI
	5 <i>Datura fastuosa</i> , <i>D. ferox</i> , <i>D. innoxia</i> , <i>D. metel</i> , <i>D. quercifolia</i> , <i>D. stramonium</i> , <i>D. tatula</i>	I, III
	6 <i>Deprea orinocensis</i> , <i>D. subtriflora</i>	I, IX, XV
	7 <i>Discopodium pernninervivum</i>	I, XIV
	8 <i>Dunalia arborescens</i> , <i>D. australis</i> , <i>D. brachyacantha</i> , <i>D. solanacea</i> , <i>D. tubulosa</i>	I, XIV
	9 <i>Exodeconus maritimus</i>	I
	10 <i>Hyoscyamus niger</i> , <i>H. muticus</i>	I
	11 <i>Iochroma australe</i> , <i>I. coccineum</i> , <i>I. fuchsiodies</i> , <i>I. gesnerioides</i>	I
	12 <i>Jaborosa araucana</i> , <i>J. bergii</i> , <i>J. caulescens</i> , <i>J. integrifolia</i> , <i>J. kurtzii</i> , <i>J. laciniata</i> , <i>J. lanigera</i> , <i>J. leucotricha</i> , <i>J. magellanica</i> , <i>J. odonelliana</i> , <i>J. parviflora</i> , <i>J. rotacea</i> , <i>J. runcinata</i> , <i>J. sativa</i>	I, IV, VIII, XIII, XVI
	13 <i>Larnax glabra</i> [11]	I
	14 <i>Lycium chinense</i> , <i>L. halimifolium</i>	I
	15 <i>Mandragora officinarum</i> [12]	I, XIV
	16 <i>Nicandra physaloides</i>	I, VI
	17 <i>Physalis alkekengi</i> , <i>P. angulata</i> , <i>P. chenopodifolia</i> , <i>P. cinerascens</i> , <i>P. coztomatl</i> , <i>P. curasavica</i> , <i>P. divaricata</i> , <i>P. franchetii</i> , <i>P. ixocarpa</i> , <i>P. lanceifolia</i> , <i>P. longifolia</i> , <i>P. minima</i> , <i>P. peruviana</i> , <i>P. philadelphica</i> , <i>P. pubescens</i> , <i>P. solanaceus</i> , <i>P. virginiana</i> , <i>P. viscosa</i>	I, III, V, VIII, XI, XVII, XIX, XX, XXI, XXII
	18 <i>Salpichroa origanifolia</i>	I, VI
	19 <i>Saracha viscosa</i>	XIX
	20 <i>Solanum ciliatum</i> , <i>S. sisymbriifolium</i>	I, II
	21 <i>Trechonaetes laciniata</i>	I
	22 <i>Tubocapsicum anomalum</i>	I, VII, XIV, XV
	23 <i>Vassobia lorentzii</i> , <i>V. breviflora</i>	I
	24 <i>Withania adpressa</i> , <i>W. aristata</i> , <i>W. coagulans</i> , <i>W. frutescens</i> , <i>W. obtusifolia</i> , <i>W. somnifera</i>	I
	25 <i>Witheringia coccoloboides</i> , <i>W. hunzikeri</i> , <i>W. solanacea</i>	XIX
Dioscoreaceae	<i>Dioscorea japonica</i> [13]	I
Fabaceae	<i>Cassia siamea</i>	I
Lamiaceae	<i>Ajuga parviflora</i> , <i>A. bracteosa</i>	I
Myrtaceae	<i>Eucalyptus globulus</i> [14]	I
Taccaceae	<i>Tacca chantrieri</i> , <i>T. integrifolia</i> , <i>T. paxiana</i> , <i>T. plantaginea</i> , <i>T. subflabellata</i>	I, XVIII
Alcyoniidae	<i>Paraminabea acronocephala</i> , <i>Minabea</i> sp. (soft coral) [15,16]	I

breast, pancreatic, prostate, lung, leukemia, as well as various head and neck squamous cell carcinomas (HNSCCs) [17]. This compound has the potential for development as an antiproliferative agent.

As part of our continuing investigations to discover drug leads from plant biodiversity, we have explored the antiproliferative potential of compounds present in several members of the large family Solanaceae. An approach of in vitro MTS cytotoxicity bioassay of *Physalis longifolia* Nutt., *Vassobia breviflora* (Sendtn.) Hunz, and *Withania somnifera* (L.) Dunal led to the identification of bioactive withanolides. A panel of HNSCC cells (JMAR, MDA1986, and DR081-1), melanoma cells (B16F10 and

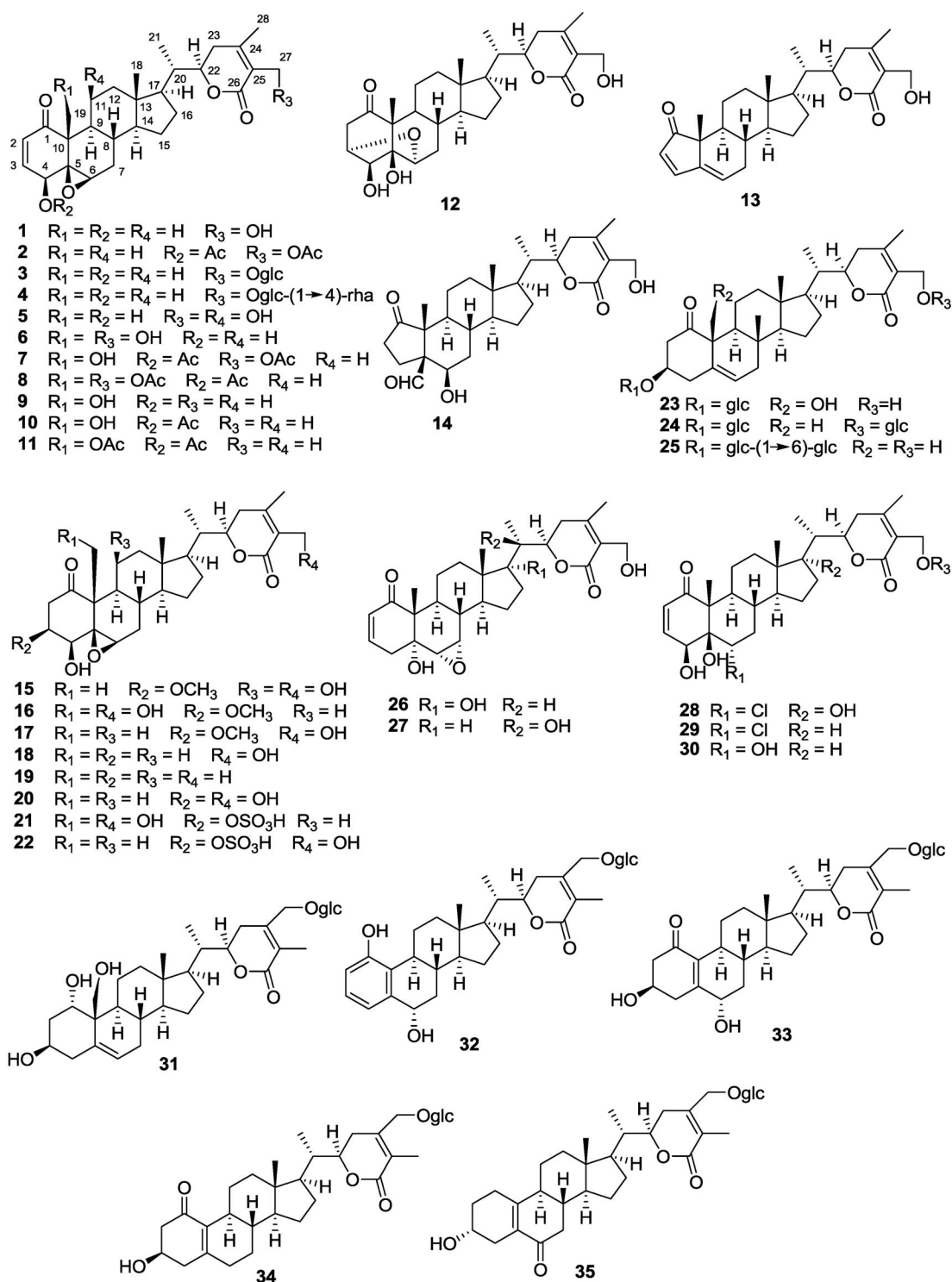


Fig. 3 Withanolides (**1**, **3–6**, **9**, **12–18**, and **20–35**) from *Physalis longifolia*, *Vassobia breviflora*, and *Withania somnifera* and their derivatives (**2**, **7**, **8**, **10**, **11**, and **19**) [17,28,29].

SKMEL28), and/or, human breast cancer cell lines (Hs578T) and non-malignant human renal epithelial cell line (MRC5) were used to evaluate the extracts/fractions/compounds. This work resulted in the isolation and characterization of 35 withanolides (**1–35**, Fig. 3). Of these, 29 withanolides (**1, 3–6, 9, 12–18**, and **20–35**) were isolated from the three taxa investigated whereas the remaining six analogues (**2, 7, 8, 10, 11**, and **19**) were prepared from the naturally occurring compounds (**1, 6**, and **9**). In this report, we provide a summary of the withanolide chemistry and anti proliferative results obtained during our studies [17,28,29] and attempt to provide a structure–activity relationship analysis (SARA) in comparison with published data in the literature for other withanolides with antiproliferative activity.

RESULTS AND DISCUSSION

Our own investigation of selected members of the Solanaceae afforded withanolides **1, 20**, and **22** from the aerial parts of *Vassobia breviflora* [17], withanolides **1, 14, 17, 18**, and **24–30** from the leaves of *Withania somnifera* [28], as well as withanolides **1, 3–6, 9, 12, 13, 15–18, 20–23**, and **31–35** from the aerial part of *Physalis longifolia* [29]. The acetate derivatives **7** and **8** were prepared from withanolide A **6**; compounds **10** and **11** were obtained from withanolide B **9** while compounds **2** and **19** were derived from withaferin A **1**. Withanolides **13–17** are likely to be artifacts formed during the isolation process. The structures of **1–35** were determined by electrospray ionization-mass spectrometry (ESI-MS) and 2D NMR experiments while those of **5, 6, 9, 13, 19**, and **28** [28,29] were confirmed by X-ray crystallographic analysis.

Based on their carbon skeletons, the withanolides isolated by our group can be assigned to four structural types with various degrees of oxygenation: type I (**1–12**, and **15–30**), type VIII (**31–35**), type XI (**13**), and type XII (**14**). All the compounds show the commonly oxygenated carbons at C-1, 22, and 26 except compound **35** which lacks an oxygenated C-1. Further oxygenated positions can occur at C-3 (**12, 15–17, 20–25, 31**, and **33–35**), C-4 (**1–12, 15–22**, and **28–30**), C-5 and C-6 in the form of an epoxy-group (**1–11** and **15–22**), C-6 and C-7 in the form of an epoxy-group (**26** and **27**), C-11 (**5** and **15**), C-17 (**26** and **28**), C-19 (**6, 9, 16, 21, 23**, and **31**), C-20 (**27**), C-27 (**1–8, 12–18**, and **20–30**), and C-28 (**31–35**). In addition, withanolides **21** and **22** occur as 3-*O*-sulfates, while withanolides **3, 4, 23–25**, and **31–35** occur as withanolide saponins with one or two sugar residues and withanolides **28** and **29** are chlorinated at C-6.

All 35 withanolides isolated during our studies were tested against the HNSCC cells (JMAR, MDA1986, and DR081-1), melanoma cells (B16F10 and SKMEL28), and/or breast cancer cells (Hs578T) as well as non-malignant human renal epithelial cells (MRC5) for their in vitro antiproliferative activities. Cisplatin was used as a positive control. The results, expressed as IC₅₀ (50 % inhibition of cell growth), are summarized in Table 2. Overall, 15 withanolides (**1–11, 16–17**, and **21–22**) including the five acetate derivatives (**2, 7, 8, 10**, and **11**) showed antiproliferative effects against the cells tested with IC₅₀ values in the range 0.067–17.4 μM, while the other withanolides were inactive when tested at the higher concentration of 20 μM. A preliminary structure–activity relationship (SAR) could be inferred based on the assay data obtained.

Table 2 IC₅₀ values (μM) of withanolides (natural and derivatives) against six cell lines [17,29].

Withanolide	Description	B16F10	SKMEL28	JMAR	MDA1986	DR081-1	MRC5	Hs578T
1	Withaferin A	0.29	3.96	2.0	0.80	1.09	0.20	1.19
2	Withaferin A 4,27-diacetate	0.15	0.11	0.19	0.32	0.76	–	–
3	27- <i>O</i> -Glucopyranosylwithaferin A	3.65	8.3	4.20	NA	NA	5.24	8.79
4	4'- <i>O</i> -Rhamnosyl 3	NA	17.4	8.24	8.1	NA	8.72	11.08
5	11β-Hydroxywithaferin A	0.49	2.99	0.77	2.64	5.46	3.60	2.47
6	19-Hydroxywithaferin A	11.1	5.06	5.32	3.34	2.37	12.7	2.22
7	6 4,27-Diacetate	0.098	0.81	0.14	2.24	1.28	0.41	0.46
8	6 4,19,27-Triacetate	0.067	0.54	0.16	0.91	0.29	0.58	0.32
9	27-Deoxy- 6	0.2	3.91	0.17	1.3	0.90	0.40	0.47
10	9 4-Acetate	0.19	0.64	0.12	0.49	0.96	0.16	0.43
11	9 4,19-Diacetate	0.13	0.27	0.24	0.11	0.54	0.51	0.26
16	2,3-Dihydro-3-methoxy- 6	3.21	9.27	4.69	NA	5.84	6.5	8.43
17	2,3-Dihydro-3-methoxy- 5	5.57	11.6	12.0	8.34	NA	7.33	–
21	2,3-Dihydro- 6 3- <i>O</i> -sulfate	1.33	4.8	2.25	2.02	NA	3.33	3.4
22	2,3-Dihydrowithaferin A 3- <i>O</i> -sulfate	0.18	5.12	0.48	0.27	2.46	1.35	1.16
Cisplatin	Positive control	1.0	1.07	1.12	1.62	–	8.90	–

Note: Cell lines include melanoma cell lines (murine B16F10 and human SKMEL28), HNSCC cell lines (JMAR, MDA1986, and DR081-1), non-malignant human renal epithelial cell line (MRC5), and breast cancer cell line (Hs578T); – not tested; NA: no activity, meaning IC₅₀ > 20 μM.

Direct comparison revealed that opening of the 5β,6β-epoxy-group of withaferin A **1** to furnish the corresponding 5β,6α-diol **29** or 6α-chloro-5β-ol **30** resulted in loss of activity. These findings confirm that the 5β,6β-epoxy-group in ring B is essential for the observed cytotoxic activity, in agreement with results previously reported in the literature for compounds with similar structural features [25–27].

Withanolides **15–22** contain a 5β,6β-epoxy-group in ring B but lack a Δ²-1-oxo-system in ring A. A loss of activity was observed when this latter feature is absent, by comparing activity of withaferin A **1** with 2,3-dihydrowithaferin A **18** or 2,3-dihydro-27-deoxywithaferin A **19**, or 2,3-dihydro-3β-hydroxywithaferin A **20**. A decrease of activity was also observed when Δ²-1-oxo-functionality was replaced with a 2,3-dihydro-3β-methoxy moiety, by comparing withaferin A **1** with 2,3-dihydro-3β-methoxywithaferin A **17**, and withalongolide **B 5** with 2,3-dihydro-3β-methoxywithalongolide **C 15**. On the other hand, the activity observed for 2,3-dihydrowithalongolide A 3β-*O*-sulfate **21** and 2,3-dihydrowithaferin A 3β-*O*-sulfate **22** was likely due to their spontaneous conversion in the cell culture media to the corresponding withalongolide A **6** and withaferin A **1**, both compounds having Δ²-1-oxo-functionality [30]. Our data thus confirm the significance of the presence of this ring A feature for the observed cytotoxic activity and are in agreement with results available in the literature [24,26].

As in the case of withaferin A **1**, withanolides **2–11** with identical functionality in rings A and B were also active, further supporting the significance of this functional array. In contrast, withanolides **12–14** and **23–35**, in which it is absent, were shown to be inactive.

Furthermore, withaferin A **1**, 27-*O*-glucopyranosylwithaferin A **3**, and 27-*O*-[α-L-rhamnopyranosyl(1 → 4)]-glucopyranosylwithaferin A **4**, in which the combination of 5β,6β-epoxy- and Δ²-1-oxo-functionality is retained, showed different activities against the cells tested. Withanolide glycoside **3** with one sugar residue was less active relative to the aglycone **1**, while withanolide saponin **4** with two sugar residues displayed less cytotoxicity when compared to **3**. This observation suggested that glycosylation of withanolides resulted in a decrease of cytotoxic activity. On the other hand, esterification of the hydroxyl groups at C-4, C-19, and C-27 increased cytotoxicity when comparing withaferin A **1** with withaferin A 4,27-diacetate **2**; withalongolide A **6** with withalongolide A 4,27-diacetate **7** and withalongolide A 4,19,27-triacetate **8**; withalongolide B **9** with withalongolide B 4-acetate **10** and withalongolide B 4,19,27-triacetate **11**.

golide B 4,19-diacetate **11** (Table 2). These results are in agreement with previously reported SARs [24–27,31]. It should be noted that the cytotoxic activity of withalongolide B **9** is comparable to that of withaferin A **1**, while the triacetate derivative **8** and diacetate derivative **11** are more potent than withaferin A **1** by exhibiting submicromolar activity with IC_{50} values in the range of 0.067–0.91 μ M against all the cell lines tested.

All withanolides so far discovered can be generally defined as poly-oxygenated steroids with all their carbon atoms bearing oxygen functions except for C-8 and C-10. Scattered throughout the literature, it is apparent by qualitative measures, that the –OH or –OR groups do not contribute to the antiproliferative ability of the active withanolides. This characteristic has been observed with different cell lines used in the evaluation of cytotoxicity for this type of compound. Various withanolides containing hydroxyl groups at different positions were assembled from the literature (Fig. 4) and listed in Table 3 for a comparison of their respective cytotoxic activities. It is clear from these data that hydroxy- or acetoxy-groups at C-4, 7, 11, 12, 14, 15, 16, 17, 18, 19, 20, 23, 24, and 27 have no significant effect on their cytotoxic activity. In most cases, withanolides possessing hydroxyl groups at such positions are slightly less potent than those without this functional group. The results of the analysis based on our own data as well as those obtained from the literature (Table 3) are summarized below, taking into consideration different functional groups in the withanolide molecules:

4-OH group: The presence of a 4 β -hydroxy-group is not required for the cytotoxic activity when comparing the activity of withanolide E **36** and 4 β -hydroxywithanolide E **37** [24,26,27]; 4-deoxy-7,16-diacetoxywithanolide D **39** and 7,16-diacetoxywithanolide D **40** [32]; 4-deoxyphysagulin C **42** and physagulin C **43** [33]; 4-deoxywithangulatin A **44** and withangulatin A **45** [33]; acnistin A **46** and 4-hydroxyacnistin E **47** [34]. In most cases, the 4-hydroxywithanolides are slightly less potent than the corresponding 4-deoxywithanolides.

7-OAc group: The presence of 7 β -acetoxy-group had essentially no effect on the cytotoxic activity when comparing the activity of withanolides D **41** and **40** [32].

11-OH group: 11 β -Hydroxywithaferin A **5** is slightly less potent than withaferin A **1** (Table 2).

12-OH group: 12 β -Hydroxywithanolide D **49** is slightly less potent than withanolide D **48** [22].

14,15-epoxy-group: 14 α ,15 α -Epoxywithaferin A **50** is less potent than withaferin A **1** [35].

16-OH group: 16 α -Hydroxyanomanolide A **52** is slightly less potent than anomanolide A **51** [36].

16-OAc group: 7 β ,16 α -Diacetoxywithanolide D **40** is slightly less potent than 7 β -acetoxywithanolide D **41** [32,37].

16,17-epoxy-group: 16 α ,17 α -Epoxywithanolide D **55** is slightly less potent than withanolide D **48** [36].

17-OH group: 17 α -Hydroxywithanolide D **56** is slightly less potent than withanolide D **48** [36].

18-OAc group: 18-Acetoxy-27-deoxywithaferin A **58** is comparable to withaferin A **1** [18].

19-OH group: 19-Hydroxywithaferin A **6** is less potent than withaferin A **1**. 2,3-dihydro-19-hydroxywithaferin A 3 β -O-sulfate **22** is less potent than 2,3-dihydroxywithaferin A 3 β -O-sulfate **21**. 4,27-Diacetoxy-19-hydroxywithaferin A **7** is comparable to 4,27-diacetoxywithaferin A **2** (Table 2). In most cases, the 19-hydroxywithanolides are slightly less potent than the corresponding 19-deoxywithanolides.

20-OH group: 20-Hydroxytubocapsanolide F **56** is comparable to tubocapsanolide F **57** while 20-hydroxytubocapsanolide A **55** is slightly less potent than tubocapsanolide A **53** [36].

23-OH group: 23-Hydroxytubocapsanolide A **54** is slightly less potent than tubocapsanolide A **53** [36].

24-OH group: 24-Hydroxywithaphysacarpin **60** is less potent than withaphysacarpin **59** [38].

27-OH group: 27-Hydroxy withalongolide B **6** is less potent than withalongolide B **9** (Table 2).

It should be noted that although there are several 21-hydroxywithanolides (mainly isolated from *Datura* [39] and *Jaborosa* [5]) and 28-hydroxywithanolides reported in the literature, their antiprolifer-

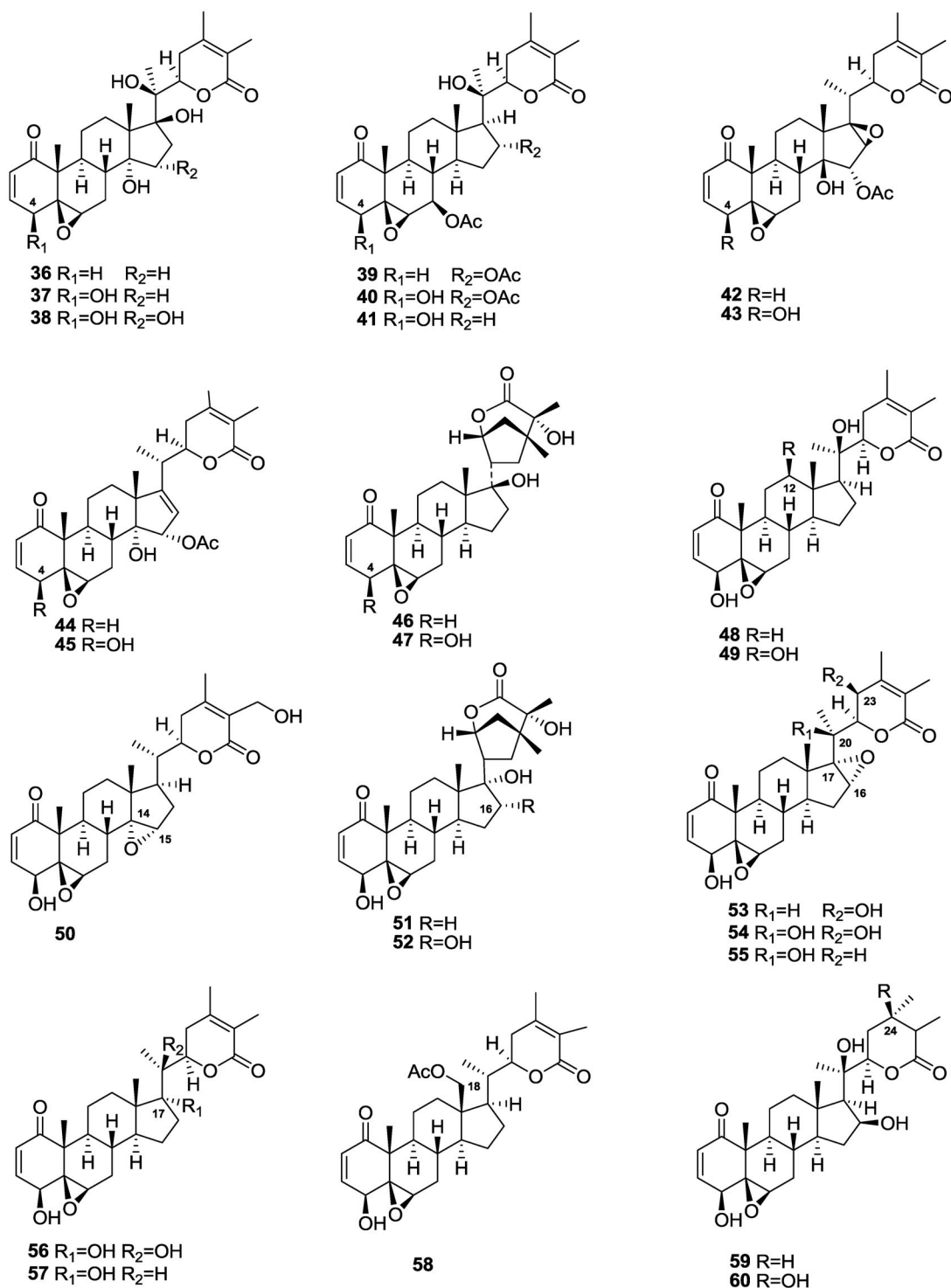


Fig. 4 Withanolides used for SARA in Table 3 (analysis from literature sources).

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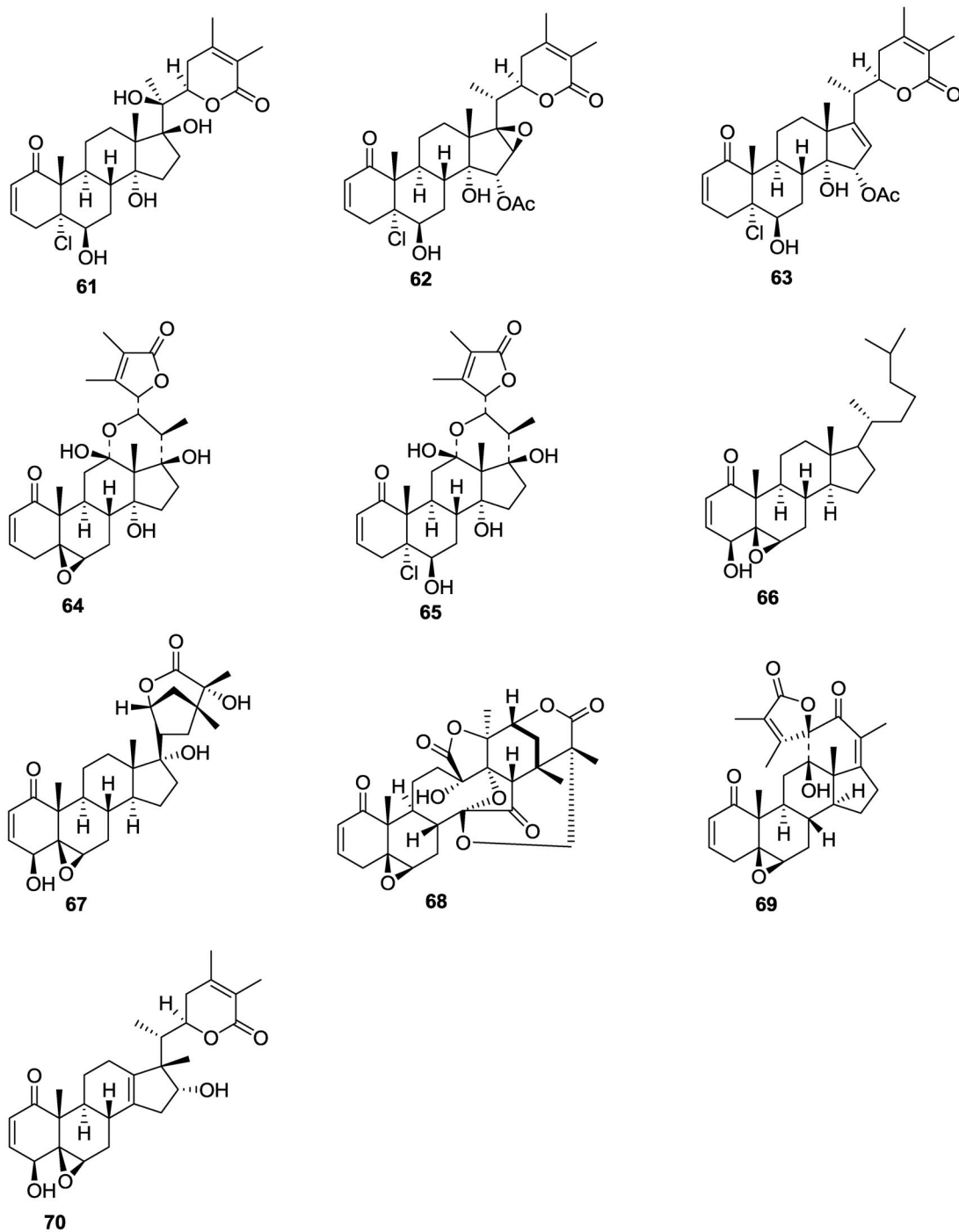


Fig. 4 (Continued).

Table 3 IC₅₀ values (μM) of cytotoxic withanolides in Fig. 4 against cancer cell lines (analysis from literature sources).

Withanolides	1	36	37	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	59	60	
L5178Y	0.64	0.62	0.99																					
T-47D		0.16	2.2																					
MCF7		0.31	1.0			10.2	>120					0.79			4.75	16.0	3.14	4.24	3.04	2.98	4.21			
MCF7/BUS	0.11	0.42	1.1																			0.59	7.37	
MDA-MB-231	0.04	0.24	2.1												2.82	3.15	0.45	2.46	0.45	1.46	2.11			
SK-BR-3		0.71	1.7																					
Hep G2	0.13	0.64	1.06												6.4	8.78	1.84	0.91	1.78	1.01	1.36			
Hep 3B	0.13	3.64	0.20												7.47	3.68	0.90	1.01	0.87	1.75	1.70			
A549	0.04	6.58	2.95												2.08	5.58	1.00	1.63	0.97	1.48	1.87			
BC-1				0.38	0.68																			
Lu-1				2.46	3.58																			
Col-2				0.06	0.14																			
KB				0.76	0.85																			
LNCap				0.38	0.34																			
HeLa						11	12	28	11															
A375										8.3	12.4													
K562										21.7	68.2													
K562R												1.23	1.54											
DAI-3b/M2												0.52	0.93											
BM cells												7.24	9.71											
EML-C1												1.37	1.37											
NCL-H460	0.38													3.1										
MRC-5	0.15											1.70					1.56	1.86	1.51	3.89	1.72			
U251																						0.45	10.36	
PC-3																						0.95	14.11	
K-562																						6.17	8.26	
HCT-15																						0.38	12.54	
SKLU-1																						0.61	5.4	
References	24,26,27,35,36	32,37				33				34		22,36	35					36				38		

ative screening data are not available in order to evaluate the significance of the presence of the –OH or –OR groups at such positions.

In addition, it has been reported that when the 5 β ,6 β -epoxy moiety in ring B is replaced by 5 α -chloro-6 β -hydroxy-functionality, the cytotoxicity of the two withanolides was comparable and in most cases the activity increased in the presence of the latter functional array. Several pairs of withanolides were compared for their activities, and similar results could be found in the literature: withanolide C **61** and withanolide E **36** [26]; physagulin I **62** and physagulin H **42** [38]; physagulin B **63** and 4-deoxy-withangulatin A **44** [38]; physagulin B **63** and withangulatin 45 [40]; and trechonolide A **64** and jaborosalactone 42 **65** [27].

Side chain: Withaferin A **1**, withanolide D **48**, withanolide E **36**, and 4 β -hydroxywithanolide E **37** bearing a δ -lactone in the nine-carbon side chain are more cytotoxic than the synthetic analogue, 5,6 β -epoxy-4 β -hydroxy-5 β -cholest-2-en-1-one **66** [24]. Tubocapsanolide F **57** with a δ -lactone in the side chain was shown to be more potent than anomanolide A **67** with a type XIV skeleton [36], suggesting the importance of a δ -lactone in order to retain cytotoxic potency.

Structural types: 4 β ,14 α -Dihydroxywithanolide E **38** with an unmodified skeleton was more potent than physalin F **68** with a type XIX skeleton and physanolide A with a type XVII framework [25]. Withanolide E **36** was more potent than jaborosalactone 1 **69** with a type XVI skeleton. Tubocapsenolide A **70** with a δ -lactone in the side chain showed strong cytotoxicity comparable to that of withanolide D **48** [36]. From these observations, it seems that type I withanolides with a lactone present in the nine-carbon side chain showed the most potent cytotoxic activity among the diverse variants discussed here.

In summary, the SARA derived from our own study and from data obtained from the literature confirmed that the 5 β ,6 β -epoxy-group or 5 α -chloro-6 β -hydroxy-functionality in ring B, and conjugated Δ^2 -1-oxo-functionality in ring A are the important requirements for the enhancement of antiproliferative activities in various cell systems. The unmodified type I withanolides showed more potent cytotoxicity than the other structural types analyzed to date. Based on these observations, it is suggested that the –OH or –OR groups at C-4, 7, 11, 12, 14, 15, 16, 17, 18, 19, 20, 23, 24, and 27 are non-contributors to the antiproliferative activity. In most cases, withanolides possessing –OH or –OR groups at such positions are slightly less potent than those lacking the –OH or –OR groups. Furthermore, glycosylation of withanolides decreased the cytotoxic effect while esterification of the hydroxyl groups reversed the effect by increasing cytotoxicity (Fig. 5).

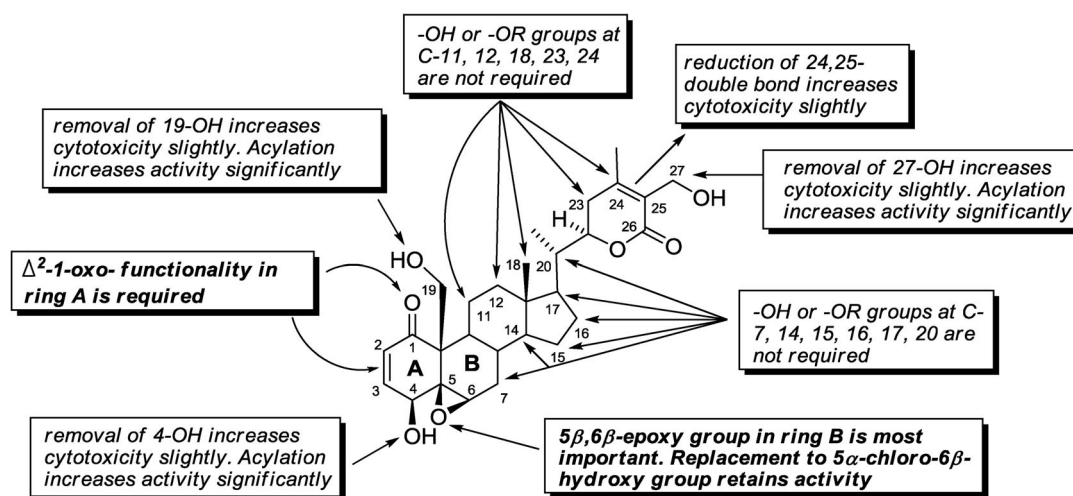


Fig. 5 SARA of withanolides.

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