

Ortenburger (1927) reported that all (number unspecified) *G. agassizii* in a cage completely exposed to the sunlight died within a day, and a tethered animal that got tangled and was unable to reach shade died before noon.

Kicking sand appears never to have been reported before in fighting male *Gopherus*. Our observations of this act show it is not merely a consequence of rapid locomotion away from the opponent but is also done by stationary gopher tortoises. One of us (JNL) has observed sand kicking as a regular feature of burrow maintenance by the entering gopher tortoise. This behavior was associated with tortoises being chased into their burrows when he (JNL) attempted to capture them for marking. He has the impression that the forefeet are always used for this purpose. Thus, one action pattern (sand kicking) is used in three contexts: burrow maintenance, predator repulsion, and intraspecific aggression.

#### LITERATURE CITED

- Camp, C. L. 1916. Notes on the local distribution and habits of amphibians and reptiles of southeastern California in the vicinity of the Turtle Mountains. Univ. California Publ. Zool. 12:503-544.
- Carr, A. 1952. Handbook of Turtles. Cornell Univ. Press, Ithaca, New York. 542 pp.
- Cassell, R. L. 1945. The ways of the desert tortoise. Desert Mag. 9(2):25.
- Douglass, J. F. 1976. The mating system of the gopher tortoise, *Gopherus polyphemus*, in southern Florida. Unpubl. M.A. thesis, Univ. of South Florida, 79 pp.
- \_\_\_\_\_. 1986. Patterns of mate-seeking and aggression in a southern Florida population of the gopher tortoise, *Gopherus polyphemus*. Proc. Symp. Desert Tortoise Council 1986:155-199.
- \_\_\_\_\_, and J. N. Layne. 1978. Activity and thermoregulation of the gopher tortoise (*Gopherus polyphemus*) in southern Florida. Herpetologica 34:358-374.
- Ernst, C. H., and R. W. Barbour. 1972. Turtles of the United States. Univ. Press of Kentucky, Lexington.
- Fletcher, W. B. 1899. The Florida gopher. Proc. Indiana Acad. Sci. 1899:46-52.
- Grant, C. 1936. The southwestern desert tortoise, *Gopherus agassizii*. Zoologica 21:225-229.
- \_\_\_\_\_. 1960. Differentiation of the southwestern tortoises (genus *Gopherus*), with notes on their habits. Trans. San Diego Soc. Nat. Hist. 12:441-448.
- Legler, J. M., and R. G. Webb. 1961. Remarks on a collection of Bolson tortoises, *Gopherus flavomarginatus*. Herpetologica 17:26-37.
- Miller, L. M. 1932. Notes on the desert tortoise (*Testudo agassizii*). Trans. San Diego Soc. Nat. Hist. 7:187-208.
- Ortenburger, A. L., and R. D. Ortenburger. 1927. Field observations on some amphibians and reptiles of Pima County, Arizona. Proc. Oklahoma Acad. Sci. 6:101-121.
- Pepper, C. 1963. The truth about the tortoise. Desert Mag. 26(10):10-11.
- Rose, F. L. 1970. Tortoise chin gland fatty acid composition: behavioral significance. Comp. Biochem. Physiol. 32:577-580.
- Weaver, W. G., Jr. 1970. Courtship and combat behavior in *Gopherus berlandieri*. Bull. Florida State Mus. 15:1-43.

#### JACK P. HAILMAN

Department of Zoology  
University of Wisconsin  
Madison, Wisconsin 53706, USA

#### JAMES N. LAYNE

Archbold Biological Station  
Post Office Box 2057  
Lake Placid, Florida 33852, USA  
and

#### ROSEMARY KNAPP<sup>1</sup>

Department of Biological Sciences  
Barnard College  
New York, New York 10027-6598, USA

<sup>1</sup>Present address: Department of Zoology, Arizona State University, Tempe, Arizona 85287, USA

### NOTES ON EARLY SPRING PARASITES AND PATHOLOGIES OF OKLAHOMA *Crotalus atrox*

A study of western diamondback rattlesnakes, *Crotalus atrox*, collected in the spring of 1987 and 1988 during Oklahoma's rattlesnake round-ups presented an opportunity to examine parasites and pathologies of this species from three localities: Waurika (Jefferson Co.), Waynoka (Major Co.), and Okeene (Blaine Co.).

#### METHODS

Snakes sampled were collected within a 30-mile radius of each locality by area residents two weeks prior to the public event (at which time snakes may be brought from various localities). In 1988, viscera from 296 snakes (128 male:168

female) were collected immediately as snakes were killed by decapitation, and were preserved at once by submerging in 10% formalin. After formalin-fixation for several days, viscera were soaked for 24 h in water, then transferred to 70% ethanol for storage. Viscera were referenced to previously weighed and measured (several parameters) snakes by recovery of a numbered plastic tag inserted deeply into the esophagus of each live snake examined. In 1987, viscera from 60 (22:38) representative Waynoka snakes were similarly collected, but were not tag-referenced to individual living snakes.

Viscera were examined during the summers of 1987 and 1988. G-I tracts were slit lengthwise from the esophagus to the rectum, and macroscopically examined for parasites, pathologies, food, and fangs. Samples of tissue and gut contents for microscopic examination were removed and transferred to vials of ethanol. Gut contents samples were examined and photographed at 160x or 1000x magnification using Kodacolor-200 film and a Zeiss "Standard-17" microscope with Contax RTS camera. Photos were printed to 86 X 125 mm size. Representative tissue samples were sectioned, stained, and examined at the Research Animal Diagnostic and Investigative Laboratory, College of Veterinary Medicine, Univ. of Missouri (Columbia) (Accession number 003155-88). Parasite indications were observed in 41 snakes (11.5%), though as will be elaborated below not all of these can be unequivocally designated as parasites of *C. atrox*. All material is presently maintained in the University of Kansas Biology Teaching Collection.

#### RESULTS

**Waurika.** Results are summarized in Table 1. Pulmonary aspergillosis was evident as severe, diffuse interstitial and alveolar granuloma with numerous hyphae and terminal conidia. The multiple intraserosal trematode cysts recorded from a single male were clustered around the small intestine. A male (1220 mm SVL, 1520 g) contained numerous pentastomid eggs, and 14 adults (*Porocephalus crotali*) were observed in the lung (see Self and Kuntz 1967).

**Okeene.** Results are summarized in Table 2. The intraserosal trematode cysts recorded were clustered primarily around the animals' small intestines and kidneys, but occasionally were found in the adjacent mesentery and the serosal surface of a fat body (one animal). The two males containing pentastomids [*Porocephalus*

*crotali* (see Self and Kuntz 1967)] were not equivalently infected. One (1168 mm SVL) contained 36 adult and subadult pentastomids in the lung; additional subadults (ca. 20 mm) were also seen in the coelom and under the mesentery adjacent to the large intestine. Total weight of all pentastomids in this animal was 8.3 g. The other male (SVL not recorded) contained one adult pentastomid (wt. 0.2 g) in its lung.

**Waynoka.** Results are summarized in Table 3. Pentastomids [*Porocephalus crotali* (see Self and Kuntz 1967)] were recorded from two females. One of these (921 mm SVL) contained 26 pentastomids (adults and subadults—21 within the lung, 4 attached to its serosal surface, and 1 attached to a kidney). Weight of the 21 from within the lung was 4.6 g. The other female (930 mm SVL) contained 25 pentastomids (wt. 5.0 g) in the lung.

Again, the multiple intraserosal trematode cysts were found clustered mainly around the small intestine and adjacent coelom wall. Additionally (in one male) cysts were adherent to the gall bladder. The female containing trematode cysts was non-reproductive. An unidentified tapeworm was found in one snake. No pentastomid eggs were noted.

**Table 1.** Waurika Summary.

Viscera examined (m:f)	137 (78:59)
Coccidians found in	12
Pulmonary aspergillosis in	7
Trematode	
ova (m:f)	2 (1:1)
cysts (m:f)	1 (1:0)
Pentastomid	
ova and/or	
adults/subadults (m:f)	1 (1:0)
Pinworm eggs	1

**Table 2.** Okeene Summary.

Viscera examined (m:f)	105 (63:42)
Coccidians found in	11
Trematode	
ova (m:f)	2 (2:0)
cysts (m:f)	10 (9:1)
Pentastomid	
ova and/or	
adults/subadults (m:f)	2 (2:0)

**Table 3.** Waynoka Summary.

Viscera examined (m:f)	60[1987] (38:22)	92[1988] (52:40)
Coccidians found in	9 (1988)	
Pulmonary aspergillosis in (m:f)	2 (2:0) (1987)	
Trematode		
ova (m:f)	2 (1:1)	
cysts (m:f)	5 (4:1)	
Pentastomid		
ova and/or		
adults/subadults (m:f)	2 (0:2)	
Unidentified tapeworm segments	1	

## DISCUSSION

The Order Coccidia contains 25 genera (Hammond and Long 1973; Olsen 1986), two of which are *Isospora* (oocysts with two sporocysts, each containing four sporozoites) and *Eimeria* (oocysts with four sporocysts, each containing two sporozoites). *Isospora* is common in reptiles and *Eimeria* is quite common in mammals, and is not uncommon in snakes. Unfortunately, formalin fixation of the viscera made it impossible to distinguish between oocysts of these coccidial genera. Therefore it is not possible to accurately determine the percent of snakes that were infected with *Isospora*. Several viperid-murid life cycles have been demonstrated for coccidians (see Wacha and Christiansen 1982; Haefner and Frank 1984; Upton and Barnard 1988), so this is not unexpected.

Pentastomid ova were found in Okeene and Waurika snakes, but none was found in Waynoka snakes. Waurika is located in south-central Oklahoma, whereas the other two localities are north-central. It is therefore unlikely that this difference is meaningful. Adult pentastomids were observed in one Waurika snake (male), two Okeene snakes (male), and two Waynoka snakes (female). Pentastomids are not uncommon parasites of snakes, and fairly large infestations seem not to cause host debilitation (Self and Kuntz 1967; Self 1969; Cooper 1984).

Intraserosal trematode cysts (Hoff et al. 1984) were identified almost exclusively from males (14 of 16 instances); of the two females, one was non-reproductive. The most common site for these cysts was the serosal surface of the small intestine. Trematode ova were found in seven animals (five of them male; one female was non-reproductive). Lyon (1986) documented helminths from six Idaho lizard species, two genera of which (*Crotaphytus* and *Cnemidophorus*) are

potential prey of Oklahoma *C. atrox*. If lizards are part of the life cycle of these parasites in Oklahoma (a likely assumption), our data would indicate differential predation by the sexes of *C. atrox* in late summer and fall.

Keenlyne (1972) reported differences in the summer feeding behavior of gravid female (versus male and non-reproductive female) *Crotalus horridus* in Wisconsin. Gravid females did not feed during the later stages of embryogenesis. As reported separately (Fitch and Pisani, in press), most of the female *C. atrox* in the populations reported here seem to follow an annual reproductive cycle. If reproductive females of this species follow the observed pattern of its congener (a pattern also reported for gravid *Agkistrodon contortrix*; Fitch and Shirer 1971), this could explain the observed differential trematode parasitism.

We additionally offer another possible, though conjectural, hypothesis for this observation. It is probable that the two lizard genera mentioned above commence fall dormancy earlier than does *C. atrox*, and that some individuals may select hibernation sites around *C. atrox* den sites. If male rattlesnakes return to den sites earlier than females, they may consume these lizards as their final fall meals, rendering them more susceptible to trematode infection. These two hypotheses are not mutually exclusive.

Finally, it is also conceivable that our observed ratio is a statistical artifact of the small percent of snakes overall that were recorded with pathologies. While the total number of snakes with pathologies is small, the strong bias observed is nonetheless interesting in light of the foregoing.

Pulmonary aspergillosis was seen in nine snakes from Waurika and Waynoka, but in none from Okeene. As remarked above for pentastomids, we feel this locality difference is not meaningful.

These findings are presented with the intent of providing a spring reference point for further and more detailed work. The life history of *Crotalus atrox* is not well known, and it would be instructive to follow the parasite cycle of this species through several years (a project we are presently not in a position to pursue).

## ACKNOWLEDGMENTS

This study was funded by a grant from the State of Oklahoma Department of Wildlife Conservation Non-game Program (grant 205-320-6188) to Pisani and Henry S. Fitch, who together collected the samples reported upon here. Hank

*Prepublication Announcement*

Herpetological Circular No. 20

**AGE DETERMINATION IN TURTLES**

George R. Zug

A complete and detailed look at techniques used to determine the age of turtles, as well as a comparison of their effectiveness. The first section deals with known-aged samples via mark-release-recapture, and captive rearing; the second section looks at measures of size as a substitute for age, using body length or mass, and lens mass; a third area of this booklet addresses incremental growth markers for age determination with respect to scute growth zones, claws and rhamphothecae, and skeletochronology; and a fourth part covers age estimation through structural modifications as revealed by scute polishing, skeletal changes, and coloration changes. Bibliography. August 1991. 28 pages. ISBN 0-916984-24-9.

*Prepublication price* \$4.<sup>00</sup>

*After October 15, 1991* \$5.<sup>00</sup>

*Orders may be sent to the Society Treasurer, Dr. Douglas H. Taylor, Department of Zoology, Miami University, Oxford, Ohio 45046 USA.*

Guarisco assisted at Waurika. We thank John Skeene for advice and logistic support through 1988. Members of the Oklahoma Herpetological Society helped in various capacities at the hunts. Andrew Grady, DVM (Univ. of Missouri, College of Veterinary Medicine, Columbia) provided identifications of aspergillosis, and trematode cysts. James Bresnahan, DVM (Animal Care Unit, University of Kansas, Lawrence) generously provided work space and access to photo equipment. James L. Christiansen, Richard Montanucci, and anonymous reviewers provided helpful discussion of parts of the data.

Many persons at the hunts generously contributed time, animals, and information to us. While it is not possible to list all these persons, we would like to acknowledge the following. Waurika: Tommy Martindale, and members of the Waurika JCs; Okeene: Lee Laubach, Gary Cypert, Ken Westfahl, Paul Johannesmeyer, Peppy Wenglarz, Bob Jenni, and the Okeene JCs; Waynoka: Bill Strond, Mildred Sampson, Irene ("Sani") Danner, Bobby Sutherland, and members of the Waynoka Saddle Club. A special note of thanks is reserved for Leo Sutherland, recently deceased, without whose acceptance and help during our 1987 pilot study and thereafter this work would have been most impractical.

**LITERATURE CITED**

Cooper, J. E. 1984. Diseases of Amphib-

ians and Reptiles. Vols I & II, Academic Press Inc., London.  
 Fitch, H. S., and G. R. Pisani. *in press*. Life history parameters of *Crotalus atrox* in Oklahoma—a baseline study. Univ. Kansas. Publ. Mus. Nat. Hist.  
 \_\_\_\_\_, and H. W. Shirer. 1971. A radiotelemetric study of spatial relationships in some common snakes. *Copeia* 1971(1):118-128.  
 Haefner, U., and W. Frank. 1984. Host specificity and host range of the genus *Sarcocystis* in three snake-rodent life cycles. *Zentralbl. Bakteriologie, Mikrobiologie, Hygiene, Ser. A* 256(3):296-299.  
 Hammond, D. M., and P. L. Long. 1973. The Coccidia. University Park Press, Baltimore, Maryland.  
 Hoff, G. L., F. L. Frye, and E. R. Jacobson. 1984. Diseases of Amphibians and Reptiles. Plenum Press, New York.  
 Keenlyne, K. D. 1972. Sexual differences in feeding habits of *Crotalus horridus horridus*. *J. Herpetol.* 6(3-4):234-237.  
 Lyon, R. E. 1986. Helminth parasites of six lizard species from southern Idaho. *Proc. Helminthol. Soc. Washington* 53(2):291-293.  
 Olsen, O. W. 1986. Animal Parasites: Their Life Cycles and Ecology. University Park Press, Dover Publications, Inc. Mineola, New York.  
 Self, J. T. 1969. Biological relationships of the Pentastomida: a bibliography on the Pentastomida. *Exp. Parasitol.* 24: 63-119.  
 Self, T. J., and R. E. Kuntz. 1967. Host-

parasite relations in some Pentastomida. *J. Parasitol.* 53:202-206.  
 Upton, S. J., and S. M. Barnard. 1988. Development of *Caryospora bigenetica* (Apicomplexa: Eimeriorina) in experimentally infected mice. *Int. J. Parasitol.* 18(1):15-20.  
 Wacha, R. S., and J. L. Christiansen. 1982. Life cycle pattern of *Caryospora* sp. (Coccidia). *J. Protozool.* 29(2):289.

**BARBARA R. STEPHENSON**  
 Animal Care Unit  
 University of Kansas  
 Lawrence, Kansas 66045, USA  
 and  
**GEORGE R. PISANI**  
 Division of Biological Sciences  
 University of Kansas  
 Lawrence, Kansas 66045, USA

**THE FIRST  
 REINTRODUCTION OF  
 BLACK CAIMAN,  
*Melanosuchus niger*, INTO  
 THE WILD**

The black caiman (*Melanosuchus niger*) is a large crocodylian found throughout the Amazon River basin, the upper Essequibo and Berbice drainages in Guyana, and in coastal French Guiana. Owing to extensive commercial hide hunting which began in the 1940s, the black caiman has